

ANTIOXIDATIVE LOW MOLECULAR WEIGHT EXTRACTIVES FROM TRIPLOID *POPULUS TOMENTOSA* XYLEM

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Triploid *Populus tomentosa* Carr. (Salicaceae) is a good alternative to meet the increasing need of the global pulp and paper industry. Meanwhile, the xylem of this species could be a useful bioresource to develop low molecular extractives with significant bioactive potential. In the present work, a phytochemical investigation on aqueous EtOH extractives of Triploid *P. tomentosa* xylem, by systematical performance of Sephadex LH-20 open column chromatography and Thin Layer Chromatography (TLC), resulted in the isolation of two phenolic acids (*p*-coumaric acid (I) and caffeic acid (II)), two flavonoids (apigenin (III) and luteolin (IV)), and three phenolic glucosides (salicortin (V), salireposide (VI) and populoside (VII)). The structure elucidation and determination of the isolated extractives were based on their spectroscopical data and physiochemical evidences. This was the first time to report the low molecular weight extractives of Triploid *P. tomentosa*. Various low molecular weight extractives from Triploid *P. tomentosa* xylem exhibited significant antioxidative activities by DPPH and hydroxyl radical scavenging assays.

Keywords: Triploid *Populus tomentosa* Carr.; Xylem; Low molecular weight extractives; Column chromatography; Spectroscopical data; Antioxidative activity

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INTRODUCTION

Chinese forest researchers have been making significant efforts to develop fast-growing trees due to the extreme shortage of xylem resource in the pulp and paper making industry. Triploid *Populus tomentosa* Carr., the cloned hardware poplar species from *Populus tomentosa* Carr., has been receiving the most attention (Zhang et al. 2010; Yang et al. 2006). Its chemical profile from a pulping point of view was reported (Yang et al. 2006). Zhang et al. (2010) have fractionally isolated and characterized the lignin and hemicelluloses of this cloned species previously. However, chemical constituents of low molecular weight extractives and biological activities of Triploid *P. tomentosa* have never been reported to date, though poplars have been widely used in folk medicines for the treatment of various diseases, such as diarrhea, pox, paralysis, variola, and pulmonary disorders (Si et al. 2010, 2009a, 2009b, 2009c; Zhang et al. 2006).

Reactive oxygen species, including superoxide anion, hydroxyl radical, and hydrogen peroxide, are chemically reactive free radicals derived from oxygen, which have been implicated in various of diseases due to the oxidative damage to DNA, lipids, and proteins, and which can result in failure of cellular functions (Brawek et al. 2010; Lane 2008). Fortunately, medical and cosmetic use or dietary consumption of antioxidative compounds will reduce oxidative damage (Hosoya et al. 2008). Some synthetic antioxidants show side effects; for example, butylated hydroxyanisole (BHA) has been reported to increase incidences of neoplasias (Tehila et al. 2007). Therefore, considerable effort has been directed towards identifying safe and natural antioxidants that can protect against oxidative stress. Natural compounds, such as phenolic acids, flavonoids, and phenolic glucosides, commonly found in many plants, have shown a wide range of biological and pharmacological activities, including antioxidative activity chelation of metals, anti-HIV, and antiplasmodial activities (Casano et al. 2010; Jung et al. 2010; Neo et al. 2010).

In the present study, a phytochemical study of 95% EtOH extractives of triploid *P. tomentosa* xylem, by repeated Sephadex LH-20 open column chromatography and Thin Layer Chromatography (TLC), led to the isolation of two phenolic acids (*p*-coumaric acid (**I**) and caffeic acid (**II**)), two flavonoids (apigenin (**III**) and luteolin (**IV**)), and three phenolic glucosides (salicortin (**V**), salireposide (**VI**), and populoside (**VII**)). The structures of the isolated phenolic compounds were elucidated and determined on the basis of their physiochemical and spectroscopical evidences (¹H- and ¹³C-NMR, as well as MS). The antioxidative activities of the isolated extractives of Triploid *P. tomentosa* xylem were assessed by DPPH and hydroxyl radical scavenging assays.

EXPERIMENTAL

Plant Materials

Xylem of Triploid *P. tomentosa* was obtained from Professor Qun Li's research group of Tianjin Key Laboratory of Pulp & Paper, College of Material Science and Chemical Engineering, Tianjin University of Science and Technology, China, in June of 2009, and was authenticated by Professor Zhong Liu, Tianjin Key Laboratory of Pulp & Paper. A voucher specimen (No. 2009-10) has been deposited at the herbarium of Tianjin Key Laboratory of Pulp & Paper.

Equipment and Reagents

Column chromatography was carried out with Sephadex LH-20 (Merck). Eluents were collected with a fraction collector (SBS-160). TLC analyses were conducted on DC-Plastikfolien Cellulose F (Merck) plates and developed with *t*-BuOH-HOAc-H₂O (3:1:1, v/v, solvent A) and HOAc-H₂O (3:47, v/v, solvent B). Visualization and detection were performed by UV light (254 and 365 nm) and then by spraying with 1% ethanolic FeCl₃ solution followed by heating.

¹H-NMR, ¹³C-NMR, DEPT, and correlation NMR spectra such as HMBC and HMQC were recorded in CD₃OD, (CD₃)₂SO, or (CD₃)₂CO with TMS as an internal standard using a Bruker Avance DPX 400 spectrometer at the operating frequency of 400

resubjected to Sephadex LH-20 column chromatography using MeOH-H₂O (2:1, v/v) for further purification to yield three subfractions, namely B₂₁, B₂₁, and B₂₃. The second (B₂₂, 3.8 g) and third subfractions (B₂₃, 2.9 g) were eluted with MeOH-H₂O (1:1 and 1:3, v/v) likewise, and compounds **I** (61 mg), **II** (43 mg), **III** (112 mg), and **IV** (88 mg) were obtained. Fraction B₃ was also applied to a Sephadex LH-20 column with EtOH-hexane (1:1, v/v) as eluting mixtures for further purification to give three subfractions that will be referred to as B₃₁ (1.2 mg), B₃₂ (7.2 g) and B₃₃ (0.6 g). When treated with MeOH-H₂O (1:1 and 1:4, v/v), compounds **V** (48 mg), **VI** (72 mg) and **VII** (36 mg) were obtained from B₃₂ as yellowish powders.

Antioxidative Activity Assays

DPPH radical scavenging assay

DPPH radical scavenging assay was carried out according to the procedure described by Blois (1958) with minor suitable modification. MeOH solutions (4 ml) of samples at different concentrations were added to a solution of DPPH (1.5×10^{-4} M, 1 ml) in MeOH. After mixing gently and standing at room temperature for 30 min, the optical density was measured at 517 nm with a UV-visible spectrophotometer (UV-1600, Rayleigh). The results were calculated by taking the mean of all triplicated values. IC₅₀ values were obtained through extrapolation from concentration of sample necessary to scavenge 50% of the DPPH free radicals. α -Tocopherol, an excellent natural antioxidant, was used as a positive control.

Hydroxyl radical scavenging assay

A hydroxyl radical scavenging assay was performed using a standard method (Chung et al. 1997). The Fenton reaction mixture consisted of 200 μ L of FeSO₄·7H₂O (10 mM), 2-deoxyribose (10 mM) and EDTA (10 mM). 200 μ L of sample and 1 mL of 0.1 M phosphate buffer (pH 7.4) were added to make up a total volume of 1.8 ml. Then, 200 μ L of 10 mM H₂O₂ was added and the reaction mixture was cultured at 37 °C for 4 h. After the incubation, 1 mL of 1% TBA and 1 mL of 2.8% TCA were added and the solution was placed in a boiling water bath for 10 min. After cooling, the solution was centrifuged for 5 min (3000 rpm) and the absorbance was measured at 532 nm with a UV-visible spectrophotometer (UV-1600, Rayleigh). Each assay was carried out in triplicate. α -Tocopherol was employed as a positive control. The hydroxyl radical scavenging activity of the sample was calculated using following formula,

$$\text{Scavenging activity (\%)} = \left[1 - \frac{A_1 - A_2}{A_0}\right] \times 100\% \quad (1)$$

where A_0 is the absorbance of the control (blank, without extract), A_1 is the absorbance in the presence of the extract, and A_2 is the absorbance without 2-deoxyribose).

Statistical Analyses

All results were presented as mean \pm SD, and the statistical analyses were subjected to one-way analysis of variation (ANOVA). The significance of difference between means was determined by Tukey's test ($P < 0.05$) for multiple comparisons.

RESULTS AND DISCUSSION

Low Molecular Weight Extractives Isolation and Identification

Based on a successive Sephadex LH-20 column chromatographic separation of a portion of BuOH soluble fraction that was extracted with 95% EtOH from xylem of Triploid *P. tomentosa* aided by TLC, seven yellowish low molecular weight natural extractives, as shown in Fig. 2, including two phenolic acids, *p*-coumaric acid (**I**), and caffeic acid (**II**) (Kim et al. 2002; Kwon et al. 1999), two flavonoids, apigenin (**III**) and luteolin (**IV**) (Agrawal 1989; Markham and Chari 1982), and three phenolic glucosides, salicortin (**V**), salireposide (**VI**), and populoside (**VII**) (Si et al. 2010, 2009a, 2009b, 2009c), were isolated, and their structure elucidation was established on the basis of their physiochemical and spectroscopic data. To the best of our knowledge, this was the first time of report natural compounds with low molecular weight from Triploid *P. tomentosa* xylem.

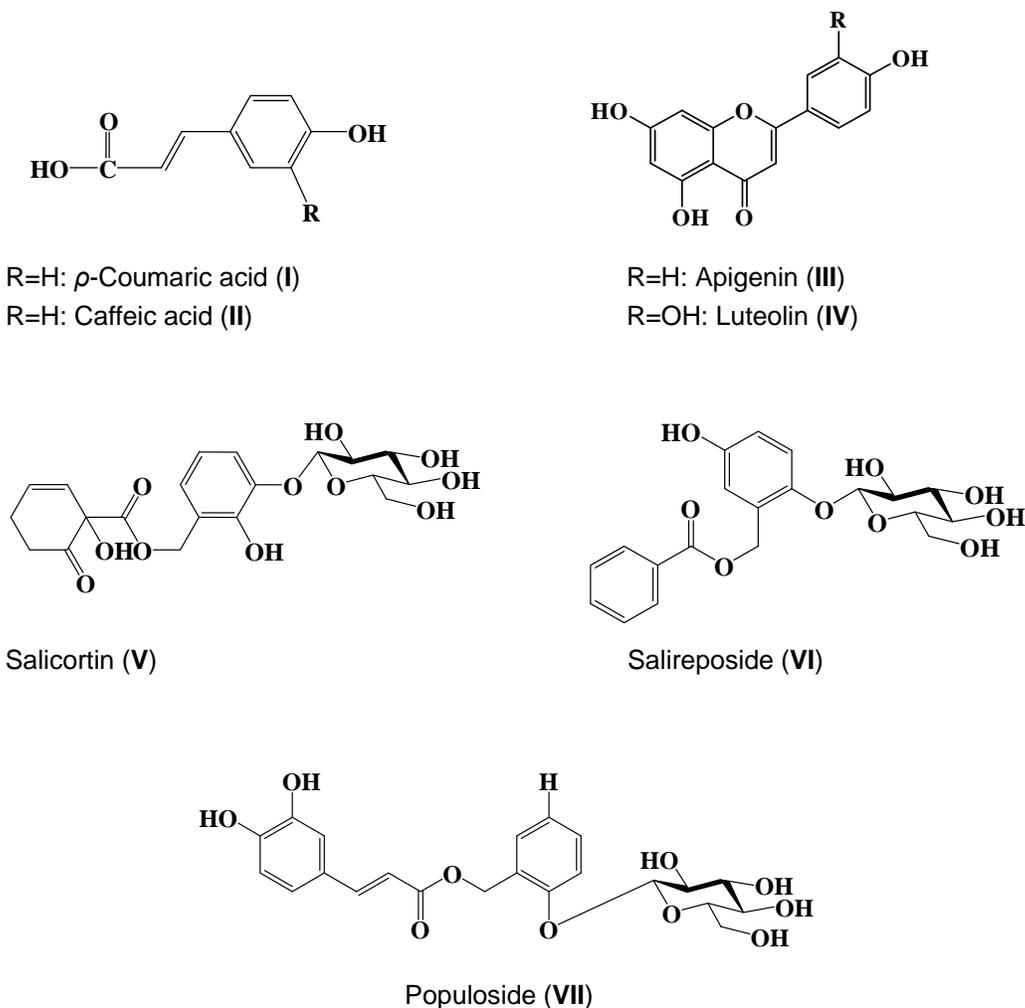


Fig. 2. Structures of the isolated low molecular weight extractives (I–VII) from Triploid *P. tomentosa* xylem

Compound I (p-Coumaric acid)

Yellowish amorphous powder; FeCl₃ test: Positive (dark brown); R_f: 0.86 (solvent A) and 0.40 (solvent B); EI-MS: Calculated for C₉H₈O₃ 164; Found [M]⁺ *m/z* 164, [M-OH]⁺ *m/z* 147 and [M-COOH]⁺ *m/z* 119. ¹H-NMR (400 MHz, δ, CD₃OD): 6.20 (1H, d, J=15.9 Hz, H-8), 6.80 (2H, d, J=8.2 Hz, H-3,5), 7.44 (2H, d, J=8.2 Hz, H-2,6), 7.60 (1H, d, J=15.9 Hz, H-7). ¹³C-NMR (100 MHz, δ, CD₃OD): 115.54 (C-8), 116.77 (C-3,5), 127.21 (C-1), 131.06 (C-2,6), 146.64 (C-7), 161.10 (C-4), 171.02 (C-9).

Compound II (Caffeic acid)

Yellowish amorphous powder; FeCl₃ test: Positive (dark brown); R_f: 0.76 (solvent A) and 0.33 (solvent B); EI-MS: Calculated for C₉H₈O₄ 180; Found [M]⁺ *m/z* 180, [M-OH]⁺ *m/z* 163 and [M-COOH]⁺ *m/z* 135. ¹H-NMR (400 MHz, δ, CD₃OD): 6.20 (1H, d, J=16.1 Hz, H-8), 6.77 (1H, d, J=8.0 Hz, H-5), 6.93 (1H, dd, J=2.0 Hz and J=8.0 Hz, H-6), 7.04 (1H, d, J=2.0 Hz, H-2), 7.53 (1H, d, J=16.1 Hz, H-7). ¹³C-NMR (100 MHz, δ, CD₃OD): 115.11 (C-8), 115.65 (C-5), 116.51 (C-2), 122.88 (C-6), 127.84 (C-1), 146.81 (C-3), 147.00 (C-7), 149.46 (C-4), 171.16 (C-9).

Compound III (Apigenin)

Yellowish amorphous powder; FeCl₃ test: Positive (dark brown); R_f: 0.77 (solvent A) and 0.05 (B); EI-MS: Calculated for C₁₅H₁₀O₅ 270; Found [M]⁺ *m/z* 270, [A₁+H]⁺ *m/z* 153, [A₁]⁺ *m/z* 152 and [B₁]⁺ *m/z* 118. ¹H-NMR (400 MHz, δ, (CD₃)₂SO): 6.20 (1H, d, J=1.7 Hz, H-6), 6.49 (1H, d, J=1.7 Hz, H-8), 6.79 (1H, s, H-3), 6.93 (2H, d, J=8.7 Hz, H-3',5'), 7.93 (2H, d, J=8.7 Hz, H-2',6'). ¹³C-NMR (100 MHz, δ, (CD₃)₂SO): 94.82 (C-8), 99.69 (C-6), 103.67 (C-3), 104.52 (C-10), 116.81 (C-3',5'), 122.01 (C-1'), 129.33 (C-2',6'), 158.16 (C-9), 162.03 (C-5), 162.30 (C-4'), 164.59 (C-2), 165.03 (C-7), 181.60 (C-4).

Compound IV (Luteolin)

Yellowish amorphous powder; FeCl₃ test: Positive (dark brown); R_f: 0.58 (solvent A) and 0.08 (solvent B); EI-MS: Calculated for C₁₅H₁₀O₆ 286; Found [M]⁺ *m/z* 286, [A₁]⁺ *m/z* 152, [A₁+H]⁺ *m/z* 153 and [B₁]⁺ *m/z* 134. ¹H-NMR (400 MHz, δ, (CD₃)₂CO): 6.26 (1H, d, J=1.9 Hz, H-6), 6.53 (1H, d, J=1.9 Hz, H-8), 6.59 (1H, s, H-3), 7.02 (1H, d, J=8.4 Hz, H-5'), 7.48 (1H, dd, J=2.0 Hz and J=8.4 Hz, H-6'), 7.52 (1H, d, J=2.0 Hz, H-2'). ¹³C-NMR (100 MHz, δ, (CD₃)₂CO): 94.72 (C-8), 99.72 (C-6), 104.21 (C-3), 105.35 (C-10), 114.14 (C-2'), 116.64 (C-5'), 120.16 (C-6'), 123.77 (C-1'), 146.49 (C-3'), 150.11 (C-4'), 158.79 (C-9), 163.38 (C-5), 164.92 (C-2), 165.16 (C-7), 183.07 (C-4).

Compound V (Salicortin)

Yellowish amorphous powder; FeCl₃ test: Positive (dark brown); R_f: 0.69 (solvent A) and 0.45 (solvent B); Positive FAB-MS: Calculated for C₂₀H₂₄O₁₀ 424; Found [M+Na]⁺ *m/z* 447 and [M+H]⁺ *m/z* 425. ¹H-NMR (400 MHz, δ, CD₃OD): 2.45–2.25 (2H, m, Ha-4", Ha-5"), 2.61–2.70 (1H, m, Hb-4"), 2.84–2.92 (1H, m, Hb-5"), 3.35–3.50 (4H, m, H-2',3', 4', 5'), 3.69 (1H, dd, J=5.0, 5.2 and 12.0 Hz, Ha-6'), 3.88 (1H, dd, J=1.8, 1.9 and 12.1 Hz, Hb-6'), 4.92 (1H, d, J=7.6 and 2.8 Hz, H-1'), 5.26 (1H, d, J=12.4 Hz, Ha-7), 5.37 (1H, d, J=12.4 Hz, Hb-7), 5.75 (1H, dt, J=1.6, 1.8, 1.7 and 9.7 Hz, H-2"), 6.14 (1H, dt, J=4.1, 3.4, 3.9 and 9.8 Hz, H-3"), 7.01 (1H, t, J=7.4 and 7.4 Hz, H-4), 7.20

(1H, d, J=8.1 Hz, H-6), 7.31 (2H, m, H-3, 5). ¹³C-NMR (100 MHz, δ , CD₃OD): 27.28 (C-4"), 36.91 (C-5"), 62.56 (C-6"), 64.44 (C-7), 71.37 (C-4'), 74.98 (C-2'), 78.06 (C-3'), 78.24 (C-5'), 79.24 (C-1"), 102.52 (C-1'), 116.67 (C-6), 123.46 (C-4), 126.16 (C-2), 129.38 (C-3"), 130.69 (C-5), 131.10 (C-3), 133.38 (C-2"), 157.01 (C-1), 171.58 (C-7"), 207.45 (C-6").

Compound VI (*Salireposide*)

Yellowish amorphous powder; FeCl₃ test: Positive (dark brown); R_f: 0.86 (solvent A) and 0.67 (B); Positive FAB-MS: Calculated for C₂₀H₂₂O₉ 406; Found [M+Na]⁺ *m/z* 429 and [M+H]⁺ *m/z* 407. ¹H-NMR (400 MHz, δ , CD₃OD): 3.37 (2H, m, H-4', 5'), 3.44 (2H, m, H-2',3'), 3.68 (1H, dd, J=5.0 and 12.0 Hz, H_a-6'), 3.87 (1H, d, J=11.4 Hz, H_b-6'), 4.78 (1H, d, J=7.5 Hz, H-1'), 5.42 (1H, d, J=13.0 Hz, H_a-7), 5.51 (1H, d, J=13.0 Hz, H_b-7), 6.72 (1H, dd, J=3.0 and 8.7 Hz, H-5), 6.86 (1H, d, J=3.0 Hz, H-3), 7.12 (1H, d, J=8.9 Hz, H-6), 7.48 (2H, t, J=7.8, 7.6 and 7.7 Hz, H-3",5"), 7.61 (1H, t, J=7.4 and 7.4 Hz, H-4"), 8.06 (2H, dd, J=1.4 and 7.8 Hz, H-2",6"). ¹³C-NMR (100 MHz, δ , CD₃OD): 62.62 (C-6'), 63.24 (C-7), 71.42 (C-4'), 75.06 (C-2'), 78.09 (C-3'), 78.18 (C-5'), 104.39 (C-1'), 116.25 (C-3), 116.62 (C-5), 119.32 (C-6), 128.76 (C-1"), 129.66 (C-3",5"), 130.68 (C-2",6"), 131.52 (C-2), 134.37(C-4"), 149.99 (C-4), 154.06 (C-1), 168.09 (C-7").

Compound VII (*Populoside*)

Yellowish amorphous powder; FeCl₃ test: Positive (dark brown); R_f: 0.80 (solvent A) and 0.44 (solvent B); Positive FAB-MS: Calculated for C₂₂H₂₄O₁₀ 448; Found [M+Na]⁺ *m/z* 471 and [M+H]⁺ *m/z* 449. ¹H-NMR (400 MHz, δ , CD₃OD): 3.3–3.5 (4H, br m, H-2', 3', 4', 5'), 3.70 (1H, dd, J=5.1 and 12.0 Hz, H_a-6'), 3.89 (1H, dd, J=1.8, 1.7 and 12.1 Hz, H_b-6'), 4.93 (1H, d, J=7.3 Hz, H-1'), 5.30 (1H, d, J=12.7 Hz, H_a-7), 5.42 (1H, d, J=12.8 Hz, H_b-7), 6.30 (1H, d, J=15.9 Hz, H-8"), 6.77 (1H, d, J=8.3 Hz, H-5"), 6.94 (1H, dd, J=2.1 and 8.2 Hz, H-6"), 7.04 (2H, m, H-4,2"), 7.23 (1H, br d, J=7.8 Hz, H-6), 7.29 (1H, td, J=1.6, 8.1 and 7.81 Hz, H-5), 7.36 (1H, dd, J=1.2, 0.9 and 7.4 Hz, H-3), 7.58 (1H, d, J=15.9 Hz, H-7"). ¹³C-NMR (100 MHz, δ , CD₃OD): 62.55 (C-6'), 62.62 (C-7), 71.34 (C-4'), 74.99 (C-2'), 78.04 (C-3'), 78.24 (C-5'), 102.92 (C-1'), 115.06 (C-8"), 115.18 (C-2"), 116.52 (C-5"), 116.71 (C-6), 123.08 (C-6"), 123.54 (C-4), 127.22 (C-1"), 127.74 (C-2), 130.41 (C-5), 130.68 (C-3), 146.83 (C-3"), 147.26 (C-7"), 149.66 (C-4"), 157.01 (C-1), 169.31 (C-9").

Antioxidative Activities of the Low Molecular Weight Extractives

Oxidative stress has been linked with the pathogenesis of many human diseases including cancer, atherosclerosis, and aging (Brawek et al. 2010; Danino et al. 2009; Si et al. 2009a). Currently, there is a growing interest in the research of natural products and plant extracts as antioxidants agents. It has been postulated and verified by researchers that such natural formulations are less toxic and have minimal side effects compared with those chemically synthesized (June et al. 2010; Neo et al. 2010). Therefore, the antioxidative activities of the low molecular weight extractives from Triploid *P. tomentosa* xylem were studied. As oxidative reactions can occur by many mechanisms, with formation of various reactive oxygen radicals, thus in this study, antioxidative activities of Triploid *P. tomentosa* xylem extractives were determined on the basis of two

classical radical scavenging procedures: DPPH and hydroxyl radical scavenging assays.

DPPH radical scavenging effect

DPPH scavenging assay is one of the most used methods to study the free radical scavenging ability of antioxidants (Blois 1958). Therefore, we firstly investigate the DPPH radical scavenging effects of the low molecular weight extractives from Triploid *P. tomentosa* xylem. As data summarized in Table 1, caffeic acid (**II**) with IC₅₀ value 5.93 μ M was clearly more efficient, meanwhile luteolin (**IV**), salireposide (**VI**), and populoside (**VII**) presented IC₅₀ values 6.81, 6.83, and 6.80 μ M, respectively, were very similar to the positive control α -tocopherol (IC₅₀ 6.82 μ M). However, ρ -coumaric acid (**I**), and apigenin (**III**) gave IC₅₀ values 8.89 and 7.89 μ M, respectively, were slightly less efficient, and salicortin (**V**, IC₅₀ value 17.88 μ M) clearly was not interesting in terms of DPPH radical scavenging.

Table 1. Free Radical Scavenging Effects of the Extractives from Triploid *P. tomentosa* Xylem

Sample	IC ₅₀ (μ M) ^a
I	8.89 \pm 0.02
II	5.93 \pm 0.01
III	7.89 \pm 0.02
IV	6.81 \pm 0.01
V	17.88 \pm 0.01
VI	6.83 \pm 0.01
VII	6.80 \pm 0.01
α -Tocopherol ^b	6.82 \pm 0.01

^a Values are mean \pm SD of three independent trials.

^b Positive control compound.

Hydroxyl radical scavenging effect

Among all of the reactive oxygen species, the hydroxyl radical plays the most important role in bacterial cell death. Oxidation is one of the key forms of damage (Kohanski et al. 2007). Accordingly, we investigated the antioxidant defense activities of low molecular weight extractives from Triploid *P. tomentosa* xylem on oxidation induced by hydroxyl radicals. From the results demonstrated in Fig. 3, generally speaking, all extractives isolated from Triploid *P. tomentosa* showed dose-dependent antioxidant effects of increasing potential at concentrations from 1 μ g/ml to 10 μ g/ml. Caffeic acid (**II**) at 1 μ g/mL and 10 μ g/mL showed 77% and 85% scavenging ability, respectively, which indicated superior antioxidative ability. Luteolin (**IV**, 49% and 58% at 1 μ g/ml and 10 μ g/ml, respectively), salireposide (**VI**, 54% and 64% at 1 μ g/ml and 10 μ g/ml, respectively), and populoside (**VII**, 52% and 61% at 1 μ g/ml and 10 μ g/ml, respectively) exhibited somewhat equivalent antioxidative potency as α -tocopherol (a positive control, 50% and 60% at 1 μ g/ml and 10 μ g/ml, respectively). By contrast, ρ -coumaric acid (**I**), and apigenin (**III**) showed weaker effects than α -tocopherol, while salicortin (**V**) exhibited almost no hydroxyl radical scavenging activity.

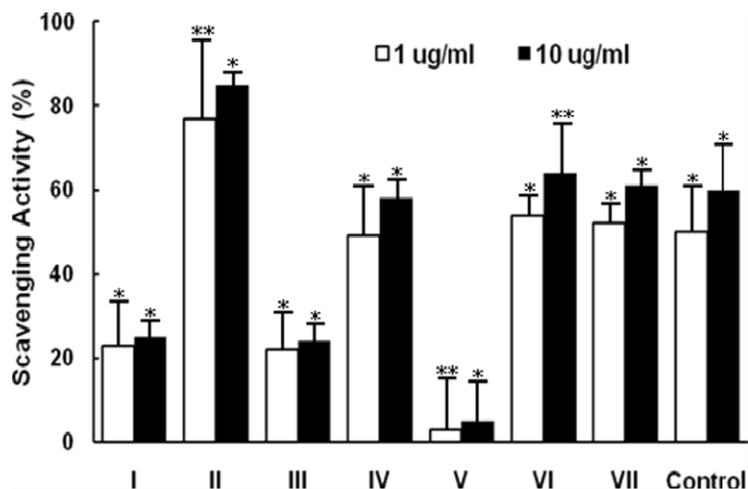


Fig. 3. Hydroxyl radical scavenging activity of the compounds **I-VII** isolated from Triploid *P. tomentosa* xylem (Values are mean \pm SD of three independent trials; α -Tocopherol was used as a positive control)

Relationships between the chemical structure and antioxidative activities of compounds have been determined in many antioxidative activity assays, and they vary with the protocols (Si et al. 2008; Wolfe and Liu 2008; Rice-Evans et al. 1996). However, some structure-activity relationships could still be concluded. For example, free hydroxyl groups acts as free radical-scavengers in the flavonoid compounds, and the structures that conferred flavonoids with the most antioxidant activity in the cellular antioxidant activity assays are a 3,4'-*O*-dihydroxyl group in the B-ring, a 2,3-double bond combined with a 4-keto group in the C-ring, and a 3-hydroxyl group (Wolfe and Liu 2008). For flavones, luteolin (**IV**) exhibits higher antioxidative activity than apigenin (**III**) (Wolfe and Liu 2008), which agrees well with our DPPH and hydroxyl radical scavenging assays. The facts above indicated that various low molecular weight extractives in Triploid *P. tomentosa* xylem exhibited significant antioxidative potency and could be an alternative to more toxic synthetic antioxidants presently used in food, pharmaceuticals, and cosmetic products.

CONCLUSIONS

1. Repeated Chromatographic separation of a BuOH-soluble fraction of Triploid *P. tomentosa* xylem resulted in the isolation of two phenolic acids, *p*-coumaric acid (**I**) and caffeic acid (**II**), two flavonoids, apigenin (**III**) and luteolin (**IV**), and three phenolic glucosides, salicortin (**V**), salireposide (**VI**), and populoside (**VII**). The structures of the isolated phenolic compounds were elucidated and determined on basis of their physiochemical and spectroscopic evidence. This was the first time of reporting the low molecular weight extractives from Triploid *P. tomentosa*, which will give rise to potential added value development of this fast-growing species.

2. Antioxidative properties of the extractives of Triploid *P. tomentosa* xylem were investigated by DPPH and hydroxyl radical scavenging assays. Compounds **II** exhibited more efficient DPPH and hydroxyl radical scavenging effects than the positive control α -tocopherol, and **IV**, **VI**, and **VII** showed the similar activities to the control. Compounds **I** and **III** revealed less efficient antioxidative potency when compared with α -tocopherol, and **V** presented almost no antioxidative capacity in DPPH and hydroxyl radical scavenging assays.
3. Triploid *P. tomentosa* xylem is a potential new source of natural antioxidants and could be developed for high value-added products.

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