Anaerobic Co-digestion of Pig Manure with Dried Maize Straw

Xiangyu Song, Keqiang Zhang, Bingya Han, Junfeng Liang, Zhongwei Zhai, and Lianzhu Du*

The anaerobic co-digestion of pig manure (PM) with dried maize straw (DMS) was studied at 35 °C with a volatile solid (VS) ratio (VS_PM/VS_DMS) of 1:2 in a continuously stirred tank reactor, and the digestions of mono-PM and mono-DMS were evaluated under the same conditions. The organic loading rates (OLRs) of 2, 3, and 4 g VS/L/d were studied and found to correspond to hydraulic retention times (HRTs) of 60, 40, and 30 d, respectively. Under the condition of long HRT and low OLR, PM could be degraded completely. The co-digestion of PM with DMS showed the most stable performance in TAN, whereas TAN in mono-PM increased with the increase of OLR. The specific methane yield (SMY) did not have a linear correlation with OLR, since HRT changed with different OLR. The maximum average SMY in the co-digestion reactor was 272 mL/g VS-fed at an OLR of 3 g VS/L/d and an HRT of 40 d. The SMY in mono-DMS digestion was the lowest and it decreased with the increase of OLR.

Keywords: Anaerobic co-digestion; Pig manure; Dried maize straw; Organic loading rates; Specific methane yield

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INTRODUCTION

With the development of the livestock industry, pig farming has become one of the most important agricultural industries in China. Pig breeding and pork consumption in China account for about half of the world’s total (Qin 2016). Extensive amounts of pig manure (PM) have become a major contributor to soil, water, and air pollution (Windisch 2001; Li and Chen 2005; Song et al. 2012). PM largely contains organic matter that is characterized by a suitable ratio of chemical oxygen demand (COD) to total solid (TS) for anaerobic digestion (AD) (Nuchdang and Phalakornkule 2012; Giuliano et al. 2013). AD is the most efficient practice for the management of PM because of the renewable biogas energy it provides and because of the suitability of the digested matter as a fertilizer for planting (Walsh et al. 2012; Xie et al. 2012; Giuliano et al. 2013; Muller et al. 2013). However, the C/N ratio of PM is within the range of 9.8 to 14.5 (Zhang et al. 2014a), which often leads to ammonia inhibition (Krylova et al. 1997; Hansen et al. 1998; Procházka et al. 2012; Wang et al. 2012b; Niu et al. 2014) because free ammonia molecules passively diffuse into cells, which causes a proton imbalance and/or a potassium deficiency that inhibits the growth of methanogens (Chen et al. 2008). To dilute toxic compounds, it is essential to maintain a proper C/N ratio (20 to 30). Some substrates characterized by high C/N ratio are often used to mix with PM to achieve a low ammonia content and proper C/N ratio (Okeh et al. 2014; Wang et al. 2015). In addition, co-digestion facilitates the
acquisition of a high level of organic matter, as well as provides missing nutrients to the microorganisms, which could result in high biogas yield (Zhang et al. 2013).

To date, most research involving the co-digestion of livestock with energy crops has mainly focused on proportion optimization. Lehtomäki et al. (2007) observed that 30% of crops in a system showed an increase in methane production by about 16% to 65% compared to digestion of manure alone. However, increasing the proportion of crops further to 40% resulted in a decrease in specific methane yield (SMY) by 4% to 12%. Giuliani et al. (2013) found that substitution of 25% energy crop with 25% agro-waste in co-digestion of PM with energy crops achieved the greatest methane production of 0.54 m³/kg VS. In addition to these studies, Linke et al. (2013) built a model to calculate the maximum methane production that could be achieved at different hydraulic retention time (HRT) by optimizing crop proportion in the co-digestion of cow manure with crops. For instance, it was found that the maximum methane yield of 388 L/kg at HRT of 60 days could be achieved only if choosing the proper crop proportion. The influence of different organic loading rates (OLR) on digester performance has also been studied. Xie et al. (2012) studied the OLRs of 1.0, 1.5, 2.0, and 3.0 g VS/L/d in anaerobic co-digestion of PM with dried grass silage, and it was found that tripling the OLR decreased methane production by 38%. Similarly, Lehtomäki et al. (2007) found that doubling the OLR from 2 to 4 g VS/L/d decreased the SMY by 16 to 26%. The optimization of the C/N ratio has also been studied. Ye et al. (2013) observed that the C/N ratio was 21.7 when the ratio of kitchen waste, PM with rice straw was 0.4:1.6:1, which resulted in the highest biogas yield of 674.4 L/kg VS. Wang et al. (2012b) investigated that the performance of pH and ammonia was more stable when C/N ratios were 25:1 and 30:1 than the other circumstances. Wang et al. (2015) also found that natural clinoptilolite could be used to control the C/N ratio in AD. However, no particular attention has been paid on the effects of different HRTs and OLRs on methane yield in AD. Dried maize straw (DMS) is a typical agricultural waste with a high C/N ratio (58 to 70) (Ouédraogo et al. 2007; Wang et al. 2013b). This characteristic makes DMS suitable for co-digestion with pig manure. In China, especially North China, most of the DMS is used as fertilizer or food for animals, and a large part is burned, which leads to a waste of resources and severe air pollution (Wang et al. 2011). Several studies have shown that the co-digestion of maize straw (MS) with animal waste is an effective method of managing MS (Dinuccio et al. 2010; Wang et al. 2012b; Yangin-Gomec and Ozturk 2013; Arici and Koçar 2015; Zou et al. 2016). However, the efficiency of integrating maize straw biomass into biogas production is very limited due to the resistance of its complex lignin structure to hydrolysis by bacteria (Jin et al. 2014; Khatri et al. 2015). Additionally, previous studies have mainly focused on silage MS, which is easier to degrade than DMS (TS, about 10% to 15%). Therefore, it is valuable and important to investigate the anaerobic co-digestion of PM with DMS in balancing OLR and HRT.

In the present study, a comparison of the digestion of mono-PM, mono-DMS, and the co-digestion of PM with DMS under mesophilic conditions (35 °C) was performed. The specific objective of this research was to investigate the effectiveness of DMS that was used to co-digest with PM and to determine the effect of different OLRs and HRTs on the digester stability and performance. The characteristics of methane production, pH, alkalinity, ammonia nitrogen, and volatile fatty acids (VFAs) were also examined in this study.
EXPERIMENTAL

Substrates and Inoculum
Both PM and DMS were collected from Yi Lilai Breeding Co. Ltd. (Tianjin, China). The PM was intraday fresh manure that was delivered to the laboratory in a plastic bucket. DMS was crushed to the size of approximately 1 mm and sealed in a plastic bag. Before feeding, PM and smashed DMS were stored in a freezer at 4 ± 1 °C. The inoculum sludge was obtained from a properly functioning pilot continuous stirred-tank reactor (CSTR) in the laboratory that ran the anaerobic digestion of pig manure under mesophilic conditions. The characteristics of the substrates and inoculum sludge are shown in Table 1.

Experimental Design
The digestion was performed in four CSTRs. The reactor was made of polymethyl methacrylate. The effective diameter, height, and volume of the reactor were 20 cm, 30 cm, and 7 L, respectively. At the top of the reactor was a stirrer device which operated for 2 h by a time relay at the speed of 50 rpm and stopped for 10 min. The gas acquisition and feeding ports were also located at the top of the reactor. The two sample connections were situated on the top and bottom sides of the device, respectively. An outlet was located at the center of the bottom. A double-layer structure maintained the reactor temperature at 35 ± 0.5 °C.

Digesters were seeded with 4 L of inoculum sludge, and tap water was added to a final working volume of 7 L. Prior to loading into the reactors, the sludge was starved for one week and then over the next 20 days acclimated with low concentrations of the substrate. The operation of the four CSTRs is shown in Table 2. PM1 was a contrast that operated the digestion of mono-PM with an OLR of 1 g VS/L/d and a HRT of 80 d. The operational parameters in PM1 remained constant during the entire experiment. The PM, PM+DMS, and DMS were loaded into the reactors of PM2, PM-DMS, and DMS, respectively, and then mixed with an OLR of 2 g VS/L/d and an HRT of 60 d. From the 69th day to the 149th day, the OLR and HRT of the reactors (PM2, PM-DMS, and DMS) were changed to 3 g VS/L/d and 40 d, and then from the 150th day to the 219th day they were changed to 4 g VS/L/d and 30 d, respectively. The ratio of VSPM/VSDMS in PM-DMS was 1:2, which maintained the ratio of C/N at around 26 in the mixed feedstock.

Analytical Methods
The feedstock was added to the reactors after discharging the digestate from the outlet each day. The digestate was collected every three days to measure pH, ammonia nitrogen (NH₄⁺-N), total alkalinity, and VFAs (acetate, propionate, isobutyric acid, butyrate, isovaleric acid, and valeric acid). The parameters (pH, NH₄⁺-N, total alkalinity, total Kjeldahl nitrogen (TKN), total organic carbon (TOC)) were analyzed in accordance with the standard methods (APHA 1998). To evaluate the VFAs, the pH of the samples was initially adjusted to 3.0 by using 5% H₂SO₄, and then centrifuged at 10,000 rpm for 10 min. The supernatants were filtered through 0.45 μm cellulose nitrate membrane filters and analyzed in a Thermo trace-1300 gas chromatograph, which was equipped with an FID detector. The temperatures of the injection inlet and detector were 200 °C. The carrier gas (helium) was maintained at a constant flow rate of 8.0 mL/min. The type of capillary column used was M12 (30 m × 0.53 mm × 1 μm, Thermo). The first stage program temperature rise was at a rate of 5 °C/min from the initial temperature of 110 °C to 150 °C.
Table 1. Characteristics of Pig Manure (PM), Dried Maize Straw (DMS), and Inoculum Sludge

<table>
<thead>
<tr>
<th></th>
<th>TS (%)</th>
<th>VS (%)</th>
<th>VS/TS (%)</th>
<th>TKN (mg/g TS)</th>
<th>TOC (mg/g TS)</th>
</tr>
</thead>
<tbody>
<tr>
<td>PM</td>
<td>30.46 ± 4.66</td>
<td>22.22 ± 5.17</td>
<td>72.34 ± 7.39</td>
<td>40.45 ± 6.72</td>
<td>434.43 ± 47.09</td>
</tr>
<tr>
<td>DMS</td>
<td>88.50 ± 0.02</td>
<td>80.72 ± 0.30</td>
<td>91.20 ± 0.35</td>
<td>11.13 ± 0.08</td>
<td>667.83 ± 4.60</td>
</tr>
<tr>
<td>Inoculum</td>
<td>4.62 ± 0.03</td>
<td>3.79 ± 0.06</td>
<td>82.03 ± 0.26</td>
<td>-</td>
<td>-</td>
</tr>
</tbody>
</table>

Note: average value (AV), standard deviation (SD)

Biogas was collected in a 20 L aluminum bag, and the gas volume was measured with a wet gas meter each day. The CH\textsubscript{4} and CO\textsubscript{2} content were measured every three days with a Thermo trace-1300 gas chromatograph that was equipped with a thermal conductivity detector, a 2 m × Φ 2 mm (diameter) stainless steel chromatographic column (PP-Q packed column), and a stationary phase (PorapakQ polymer beads). Helium was used as carrier gas at a constant pressure of 75 kPa. The temperature of the oven, injection inlet, and detector were 40 °C, 200 °C, and 200 °C, respectively.

Table 2. Operation Conditions of Reactors

<table>
<thead>
<tr>
<th>Reactors</th>
<th>Substrate</th>
<th>Stage</th>
<th>OLR (g VS/L/d)</th>
<th>HRT (d)</th>
</tr>
</thead>
<tbody>
<tr>
<td>PM1</td>
<td>PM</td>
<td>Whole stage</td>
<td>1</td>
<td>80</td>
</tr>
<tr>
<td>PM2</td>
<td>PM</td>
<td>I, II, and III</td>
<td>2, 3, and 4</td>
<td>60, 40, and 30</td>
</tr>
<tr>
<td>PM-DMS</td>
<td>PM + DMS</td>
<td>I, II, and III</td>
<td>2, 3, and 4</td>
<td>60, 40, and 30</td>
</tr>
<tr>
<td>DMS</td>
<td>DMS</td>
<td>I, II, and III</td>
<td>2, 3, and 4</td>
<td>60, 40, and 30</td>
</tr>
</tbody>
</table>

Note: stage I, 1-68 d; stage II, 69-149 d; stage III, 150-219 d

RESULTS AND DISCUSSION

Effect of Feedstock and OLR on Methane Production

Table 3 shows the average values of methane production and CH\textsubscript{4} content of PM1. The reactor of PM1 was conducted as a control to see the potential methane production of PM by itself. As the results showed, the average SMY in PM1 (295 mL/g VS\textsubscript{fed}) was the highest value compared to those of the other reactors at different OLRs, which suggested that long HRT with low OLR could make substrates degrade completely. PM has been demonstrated to be an excellent substrate for AD in various conditions (Xie et al. 2011; Zhang et al. 2014b). The SMY of mono-PM digestion in this experiment was lower than the reported 354.7 mL/g VS\textsubscript{fed} of Zhang et al. (2014a), but similar to that of 300 mL/g VS\textsubscript{fed} described by Hansen et al. (1998). However, long HRT with low OLR was meant to treat a small quantity of waste over a long period of time, which was regarded as poor efficiency to AD, so the experiment of 2 to 4 g VS/L/d proceeded in PM2.

In PM2, the average SMY at an OLR of 2 g VS/L/d and HRT of 60 d was 203 mL/g VS\textsubscript{fed}. The SMY increased with the increase of OLR. As time went by, the increase of OLR provided increasing nutrients for the microbe in the reactor, which increased the
activity of the microbe. Meanwhile, the difference between different batches of PM affected the methane production. Besides, when the OLR was increased to 3 g VS/L/d and 4 g VS/L/d, the average SMY increased by only 5% and 8%, respectively, while the SMY in PM2 during the OLR changed from 2 to 4 g. The VS/L/d was 22% to 31% lower than that of PM1, which suggested that the digestion of PM alone did not show a significant response with the increase of OLR.

![Graph](image)

**Fig. 1.** a) SMY and b) CH₄ content of the reactors

Considering that the C/N ratio of PM is not in the proper range for AD, the co-digestion of PM with DMS was carried out in the reactor denoted as PM-DMS with adjustment of the C/N ratio to 26. Methane production of the co-digestion of PM with DMS was higher than the mono-PM or mono-DMS digestion at the same OLR level. The average SMYs in PM-DMS were 7% to 28% and 34% to 193% higher, respectively, compared to that in PM2 and DMS, thereby showing the advantage of co-digestion over mono-digestion. In a similar experiment conducted by Lehtomäki *et al.* (2007), a 16% to 65% increase in methane production per digester volume was observed in a feedstock that included a 30% proportion of crops when compared to the digestion of manure alone. The
highest SMY values were observed at an OLR of 3 g VS/L/d in PM-DMS, with an average of 272 mL/g VS·fed, which were 25% and 7% higher than those of 2 g VS/L/d and 4 g VS/L/d OLR, respectively. The OLR at the first stage was the lowest. The lack of organic matter in the biomass resulted in low methane production, even though HRT was the longest. The increase in SMY achieved by co-digestion in the second stage was attributed to the increase in the biodegradability of DMS by co-digestion or synergism for methane potential. The higher biodegradability in PM-DMS could have been due to the strong microbial activity supported by the readily biodegradable organics in the PM, thereby contributing to a higher hydrolytic capacity of cellulose and hemicellulose in the DMS. Zhang et al. (2014b) reported that the co-digestion of PM and dewatered sewage sludge provided balanced amounts of nutrients in the mixed substrates and accelerated bacterial growth and hydrolysis, thereby enhancing methane formation. However, at the third stage, despite the high OLR, the HRT was very short, which was probably limited by the volume of the reactor, which in turn led to insufficient degradation (Lehtomäki et al. 2007). These observations indicated that the selection of an appropriate OLR and HRT combination was crucial to increasing methane production. Besides, as a substrate that has poorer degradability than MS silage, DMS in the co-digestion with PM of this experiment did not show any disadvantage, compared with the experiment that did by Arici and Koçar (2015), who observed that the maximum SMY was 215 mL/g VS·fed in the co-digestion of cattle manure and maize silage.

In DMS the average SMY values decreased by 8% and 47%, respectively, when the OLR increased from 2 to 3 and 4 g VS/L/d, possibly due to the limited degradation of DMS that resulted from the shielding effect, which was caused by the intense cross-linking of the lignin with cellulose and hemicellulose (Lehtomäki et al. 2007). Meanwhile, DMS floated to the up side of the reactor, which made it difficult to stir so that DMS could not contact with the microbes sufficiently.

The CH₄ content of the biogas is shown in Fig. 1b. The CH₄ percentage of PM-DMS was lower than that of PM1 and PM2, but higher than that of DMS. These discrepancies may be attributable to the characteristics of the different substrates. Proteins and fats which are abundant in PM are the main conversion components to methane (Petersen and Henius 1984; Astals et al. 2012). However, the poor degradability of lignin in DMS (Lehtomäki et al. 2007) decreases the methane potential. The CH₄ percentage in biogas produced by carbohydrates is lower than that produced by protein and fat (Moller et al. 2004).

**pH and Ammonia Nitrogen Measurements**

The pH is a key parameter that describes the stability of AD. No significant differences in pH among the four reactors were observed during the first stage (0 to 69 days).

For PM1, with the continuous loading at a low OLR, the pH values were within the range of 7.30 to 8.14. No significant difference in pH was observed between the reactors of PM1 and PM2, for they were both fed with PM alone (Fig. 2a).

The pH value of PM-DMS decreased from 7.48 to 7.24 at the 69th day when the OLR was increased to 3 g VS/L/d, which meant that the sudden increase in OLR accelerated the hydrolysis. With the stabilization of OLR, the pH fluctuated at around 7.4, which was consistent with the high buffering capacity and the normal growth of anaerobic microorganisms (Raposo et al. 2009). The decrease of pH in PM-DMS at the 69th day corresponded with the accumulation of VFA (Fig. 3). Furthermore, with progress in
digestion, a balance between acidogenesis and methanogenesis was achieved, and the performance of the digester gradually stabilized (Wang et al. 2013a).

For DMS, the pH rapidly decreased after increasing the OLR from 2 to 3 g VS/L/d and then stabilized at around 7.0 (Table 3). In AD, the observed change in pH was attributed to the degradation and production of VFAs and the conversion of ammonia nitrogen. The decrease of pH in DMS could have been caused by the hydrolysis acidification that resulted in the accumulation of VFA without a high buffering capacity. The increase in total ammonia nitrogen (TAN) could have led to an increase in pH given the chemical equilibrium between ammonium (NH$_4^+$) and free ammonia (NH$_3$), which could have been responsible for the high pH that corresponded to the observed elevated levels of ammonia nitrogen (Fig. 2b) in PM1 and PM2.

![Fig. 2. a) pH values and b) NH$_4^+$-N concentrations of the reactors during the experiment](image)

The total ammonia nitrogen can also play a critical role in the performance and stability of the anaerobic digestion of nitrogen-rich materials such as animal manure (Massé et al. 2014). The increase in TAN concentration provides a better buffering capacity (Walker et al. 2011), thereby affecting methane production. However, ammonia inhibition usually happens when the pH is above 7.4 and the TAN is between 1,500 and 3,000 mg/L, whereas inhibition will occur irrespective of pH when the TAN concentration exceeds 3,000 mg/L (Walker et al. 2011). According to Procházka et al. (2012), the optimal TAN
concentration for methanogens is 2,100 mg/L in the AD of PM, and the highest concentration that methanogens can tolerate is 4,200 mg/L.

In PM1, the TAN was stable at around 2,318 mg/L which was in the safety range of AD and corresponded with the stable performance in PM1 (Table 3).

The average TAN in PM2 gradually increased from 2,482 to 4,384 mg/L, with OLR increasing from 2 to 4 g VS/L/d, for the possible reason that ammonium is readily released during the mono-PM digestion for the relatively low C/N ratio. The average SMY values in PM2 were 203, 213, and 230 mL/g VS-fed (Table 3), respectively, at the three OLR levels, which suggested that no inhibition was observed even though the TAN was higher than the reported critical inhibition concentration (4,200 mg/L) when the pH was higher than 7.8. This could be attributed to the acclimation of the inoculum, which weakened the influence of ammonia (Chen et al. 2008).

The TAN of PM-DMS that co-digested PM with DMS gradually decreased from 1,572 to 1,154 mg/L, with OLR increasing from 2 to 3 g VS/L/d (Fig. 2b). In contrast, the TAN was stable at around 1,020 mg/L, and it did not decrease when the OLR increased to 4 g VS/L/d in PM-DMS, which showed that the co-digestion of PM with DMS could prevent the accumulation of ammonia.

For DMS, the average TAN gradually decreased, and it was below 300 mg/L after 102 days (Fig. 2b). When the OLR increased to 4 g VS/L/d, the TAN was stable below 50 mg/L. With the increase of OLR, the microbe in the digestion of DMS needed more nitrogen as a nutrient to degrade more substrate, but DMS characterized by the high C/N ratio could not provide enough nitrogen. According to Procházka et al. (2012), when the ammonia nitrogen concentration was lower than 500 mg/L, microorganisms were unable to acquire enough ammonia as a nutrient. So the insufficient of nitrogen might be the main reason for the decrease of TAN.

VFAs and TVFA/Alk

Acetic acid is the main intermediate in AD, and as much as two-thirds of the biologically generated methane is derived from acetate (Liu and Whitman 2008). Propionic acid is the most toxic organic acid and is not easily utilized by methanogens (Procházka et al. 2012). The concentrations of propionate, isobutyrate, butyrate, isovalerate, and valerate, together with acetate, were converted into total VFA and used in the calculation of the ratio of TVFA to alkalinity (TVFA/Alk), which is an important index for the evaluation of the stability of the AD process. The profiles of acetic acid, propionic acid, and TVFA/Alk for the four reactors are presented in Fig. 3.

For PM1, the average values of acetic acid and propionic acid were 86.67 mg/L and 28.13 mg/L when the OLR was 1 g VS/L/d, and the TVFA/Alk ratio was below 0.1. No particular imbalance was observed in PM1.

An increase in acetic acid and propionic acid concentration was observed in PM2 when the OLR increased from 2 to 3 g VS/L/d (Fig. 3), which may have been caused by the increase in organic matter in the biomass, leading to an acceleration of acidification. Figure 3c shows that the TVFA/alkalinity ratio in PM2 was below 0.1, which in turn indicated that the high alkalinity was providing a high buffering capacity (Astals et al. 2012) to maintain continuous digestion. When the OLR was increased to 4 g VS/L/d, the acetic acid and propionic concentrations in the PM2 increased, but the TVFA/alkalinity ratio remained below 0.1, which meant that the increase in VFA concentration was not sufficient to inhibit the anaerobic process, which in turn was demonstrated by an increase in SMY by 7% compared to that when using an OLR at 3 g VS/L/d.
For PM-DMS, the VFAs (acetate, isobutyrate, butyrate, isovalerate, and valerate) were almost depleted by the methane-producing bacteria during anaerobic digestion. The predominant conjugate base of carboxylic acid was acetate, which comprised more than 60% of the total VFA. Furthermore, isobutyrate and isovalerate, which are considered the best indicators of AD imbalance (Hill and Holmberg 1988; Kalamaras and Kotsopoulos

Fig. 3. a) Variations in the levels of acetic acid, b) propionic acid, and c) TVFA/Alk in the four reactors
2014), were almost totally depleted during the anaerobic process. The concentrations of isobutyrate and isovalerate showed a 15- and 10-fold decrease, respectively, compared to that of PM2 (data not shown). Meanwhile, the VFA/Alk ratio was below 0.1 during the experiment, which reflected the fact that the buffer capacity of the AD system was enhanced by the co-digestion of PM with DMS.

For DMS, the concentration of propionic acid was within the range of 50 to 250 mg/L when the OLR was increased to 4 g VS/L/d, which was still within the safe range (<900 mg/L) (Wang et al. 2009). The TVFA/alkalinity ratio also had a significant increase, yet remained within the safe range (0.3 to 0.4) according to Wang et al. (2012a) and Liu and Whitman (2008), except for a peak value of 0.43 that was observed at the 169th day. This could be explained by the poor buffering capacity caused by the extremely low TAN concentrations (Fig. 2). Therefore, higher risks for VFA (propionic acid) accumulation and low methane yield occurred at high OLRs during DMS digestion, which was characterized by the high C/N ratio and was indicated by a decrease in SMY when the OLR increased from 3 to 4 g VS/L/d.

**Table 3.** Characteristics of the Four Reactors Used in this Study

<table>
<thead>
<tr>
<th>Reactor</th>
<th>OLR (g VS/L/d)</th>
<th>DBP (L/d)</th>
<th>DMP (L/d)</th>
<th>CH₄ (%)</th>
<th>SMY (mL/g VS&lt;sub&gt;fed&lt;/sub&gt;)</th>
<th>NH₄⁺-N (mg/L)</th>
<th>Alkalinity (mg/L)</th>
<th>pH</th>
</tr>
</thead>
<tbody>
<tr>
<td>PM1</td>
<td>1</td>
<td>4.08 ± 0.20</td>
<td>2.08 ± 0.09</td>
<td>51.7 ± 1.28</td>
<td>295 ± 12</td>
<td>2318 ± 32</td>
<td>12163 ± 180</td>
<td>7.76 ± 0.01</td>
</tr>
<tr>
<td>PM2</td>
<td>2</td>
<td>6.22 ± 0.49</td>
<td>2.81 ± 0.24</td>
<td>47.0 ± 2.81</td>
<td>203 ± 18</td>
<td>2482 ± 65</td>
<td>15841 ± 522</td>
<td>7.61 ± 0.03</td>
</tr>
<tr>
<td></td>
<td>3</td>
<td>7.23 ± 0.81</td>
<td>4.27 ± 0.46</td>
<td>56.5 ± 1.92</td>
<td>213 ± 24</td>
<td>3756 ± 96</td>
<td>21427 ± 427</td>
<td>7.95 ± 0.02</td>
</tr>
<tr>
<td></td>
<td>4</td>
<td>11.90 ± 1.10</td>
<td>6.17 ± 0.54</td>
<td>51.1 ± 1.83</td>
<td>230 ± 20</td>
<td>4384 ± 53</td>
<td>18380 ± 287</td>
<td>7.90 ± 0.03</td>
</tr>
<tr>
<td>PM-DMS</td>
<td>2</td>
<td>7.16 ± 0.46</td>
<td>3.10 ± 0.30</td>
<td>42.8 ± 3.03</td>
<td>218 ± 22</td>
<td>1572 ± 55</td>
<td>10443 ± 318</td>
<td>7.45 ± 0.04</td>
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<tr>
<td></td>
<td>3</td>
<td>9.88 ± 0.71</td>
<td>4.87 ± 0.25</td>
<td>50.0 ± 1.33</td>
<td>272 ± 21</td>
<td>1154 ± 80</td>
<td>7706 ± 305</td>
<td>7.41 ± 0.03</td>
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<td></td>
<td>4</td>
<td>14.59 ± 0.39</td>
<td>6.77 ± 0.33</td>
<td>46.7 ± 1.81</td>
<td>254 ± 10</td>
<td>1020 ± 33</td>
<td>5874 ± 318</td>
<td>7.35 ± 0.05</td>
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<tr>
<td>DMS</td>
<td>2</td>
<td>6.08 ± 0.51</td>
<td>2.30 ± 0.24</td>
<td>38.1 ± 2.92</td>
<td>163 ± 17</td>
<td>1175 ± 98</td>
<td>7552 ± 630</td>
<td>7.49 ± 0.04</td>
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<td></td>
<td>3</td>
<td>5.76 ± 0.75</td>
<td>2.95 ± 0.35</td>
<td>48.9 ± 1.39</td>
<td>150 ± 18</td>
<td>284 ± 29</td>
<td>3616 ± 269</td>
<td>7.06 ± 0.04</td>
</tr>
<tr>
<td></td>
<td>4</td>
<td>5.94 ± 0.69</td>
<td>2.26 ± 0.19</td>
<td>39.5 ± 2.15</td>
<td>87 ± 8</td>
<td>22 ± 3</td>
<td>1723 ± 43</td>
<td>6.98 ± 0.03</td>
</tr>
</tbody>
</table>

Note: DBP, daily biogas production; DMP, daily methane production

**CONCLUSIONS**

1. The co-digestion of pig manure (PM) with dried maize straw (DMS) showed the advantage of ensuring the stability of anaerobic digestion (AD) and improving methane production by promoting a suitable C/N ratio and buffer capacity compared with the mono-PM digestion.

2. Under the condition of long hydraulic retention times (HRT) and low organic loading rates (OLR), PM could be degraded completely.

3. With the increase of OLR, no linear correlation to specific methane yield (SMY) was observed in this experiment. The maximum average SMYs obtained in mono-PM digestion, co-digestion of PM with DMS and mono-DMS digestion were at different conditions of OLR and HRT.

4. High OLR resulted in an increase in the concentrations of TAN in mono-PM digestion, and it resulted in decrease in TAN in the digestion of mono-DMS, while no significant changes of TAN was observed in the co-digestion of PM with DMS.

5. The DMS was not suitable for digestion alone, but it served effectively as an alternative substrate for co-digestion with PM.

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