

## Effect of Various Pretreatment Methods on Sugar and Ethanol Production from Cellulosic Water Hyacinth

Shahabaldin Rezanian,<sup>a,\*</sup> Hossein Alizadeh,<sup>b</sup> Junboun Park,<sup>a,\*</sup> Mohd Fadhil Md Din,<sup>c</sup> Negisa Darajeh,<sup>d</sup> Shirin Shafiei Ebrahimi,<sup>e</sup> Bidyut Baran Saha<sup>f,g</sup> and Hesam Kamyab<sup>h</sup>

Effects of acid, alkali, ionic liquid (IL), and microwave-alkali pretreatments on cellulosic water hyacinth (WH) were investigated based on the total reducing sugars (TRS) and ethanol production. For the first time, IL pretreatment with (1-Ethyl-3-methylimidazolium acetate ([EMIM][Ac])) was used for WH, and the efficiency was compared with the other methods. *Cellulase* and *Saccharomyces cerevisiae* were fermented together for 72 h. Based on the results, all pretreatment methods effectively increased the sugar content as well as the ethanol yield. Untreated WH had  $25 \pm 1.5$  mg/g of TRS, which was increased to  $157 \pm 8.2$  mg/g,  $95 \pm 3.1$  mg/g,  $51 \pm 4.2$  mg/g, and  $45 \pm 2.6$  mg/g via alkali, microwave-alkali, acid, and IL pretreatments, respectively. The highest TRS level of 402 mg/g was obtained in 24 h and  $6.2 \pm 0.4$  g/L of ethanol in 48 h of fermentation with the alkali-treated WH. The ethanol production was followed by other treatment methods of WH in the order of microwave-alkali, acid, and IL. The results indicated that the ethanol production from WH was related to the type of pretreatment as well as the TRS production.

**Keywords:** Water hyacinth; Ethanol production; Total reducing sugar; Pretreatment methods

**Contact information:** a: Department of Civil and Environmental Engineering, Seoul National University, Seoul, Republic of Korea; b: Bio-Protection Research Centre, Lincoln University, Lincoln 7647, New Zealand; c: Centre for Environmental Sustainability and Water Security (IPASA), Research Institute for Environmental Sustainability, Universiti Teknologi Malaysia, 81310, Johor Bahru, Malaysia; d: School of Biological Sciences, University of Canterbury, Christchurch 8140, New Zealand; e: School of Education, Universiti Teknologi Malaysia, Johor Bahru, Malaysia; f: International Institute for Carbon-Neutral Energy Research (WPI-I2CNER), Kyushu University, 744 Motoooka, Nishi-ku, Fukuoka 819-0395, Japan; g: Mechanical Engineering Department, Kyushu University, 744 Motoooka, Nishi-ku, Fukuoka 819-0395, Japan; h: Engineering department, Razak faculty of technology and Informatics, Universiti Teknologi Malaysia, Jalan sultan Yahya Petra, 56100, Kuala Lumpur;

\* Corresponding authors: shahab\_rezanian89@yahoo.com; junbpark@snu.ac.kr

## INTRODUCTION

Fossil fuel depletion in the near future can be predicted by the continuous over-utilization of natural energy sources (Jambo *et al.* 2016). Agricultural products, such as corn, wheat, and sugarcane, are defined as the first generation of raw materials for bioethanol production. Likewise, lignocellulosic biomass is the second generation, and algal biomass is the third generation (Baeyens *et al.* 2015). This energy source also has the potential to reduce greenhouse gas emissions originating from fossil fuels and by carbon sequestration (Tye *et al.* 2016). The use of lignocellulosic biomass as the feedstock for fermentable sugar production is the key element in sustaining the production of bioproducts like bioethanol (Mandade *et al.* 2016).

Water hyacinth is a type of lignocellulosic material that is potentially available in tropical regions (Sing and Bishnoi 2013; Rezanian *et al.* 2015). In an untreated biomass, cellulose, hemicelluloses, and lignin are linked to form a strong structure that is difficult to process (Rezanian *et al.* 2017). Because the lignin content of the lignocellulose can affect the ethanol production process, the selection of suitable biomass is important. The WH has high carbohydrate and low lignin content, an impressive growth rate, and no competition on land, which makes it a suitable lignocellulosic material for bioenergy generation (Rezanian *et al.* 2016). The economic potential of biofuel production can be improved by using low-lignin biomass in comparison to rich lignin biomass (Bhatt and Shilpa 2014). According to Singh and Bishnoi (2012), there are three main operations needed to convert lignocellulose to bioethanol: pretreatment (necessary to liberate cellulose and hemicellulose), cellulose hydrolysis to produce fermentable sugars, and the fermentation of reducing sugars. In this way, the most common and traditional microorganism used in industrial bioethanol production is the yeast *Saccharomyces cerevisiae* (Dionisi *et al.* 2015).

A major challenge for bioethanol production is finding a cost-effective pretreatment of lignocellulosic biomass (Singh *et al.* 2015). Different cost-effective pretreatment methods have been identified based on different types of lignocellulosic biomass and productivity (Srivastava *et al.* 2015). The effectiveness of these pretreatments depends on the physical structure and chemical composition of the biomass, as well as the pretreatment conditions (Sarto *et al.* 2019). The choosing of a pretreatment method can determine the success in overcoming this complicated structure and the factors that constrain the enzymatic hydrolysis of the lignocellulosic biomass (Sun *et al.* 2016). For instance, to catalyze the polysaccharide in the lignocelluloses, cellulase can serve an important catalytic enzyme with appropriate levels of stability in many ILs (Elgharabawy *et al.* 2016). In addition, thermal pretreatment, mild alkali pretreatment, and microbial combined dilute acid pretreatment are new approaches for ethanol production from cellulosic WH (Barua and Kalamdhad 2017; Narra *et al.* 2017; Zhang *et al.* 2018).

To increase the bioethanol yield, various pretreatment methods are applied to decrease the recalcitrance of lignocellulosic feedstock (Zheng *et al.* 2014). The supernatant from the lignocelluloses enzymatic hydrolysis contains both hexoses and pentoses sugars (Rezanian *et al.* 2018). As demonstrated by Cheng *et al.* (2014), the simultaneous processes of pretreatment and wet storage conserve 70% of carbohydrates and remove 40% of lignin with 325 mg/g reduced sugar yield from the WH biomass. Therefore, the aim of this study is to evaluate the effectiveness of acid, alkali, IL, and microwave-alkali pretreatments on the ethanol and total reducing sugars production from WH based on fermentation time. Meanwhile, the main novelty of this study was to use IL pretreatment with 1-ethyl-3-methylimidazolium acetate ([EMIM][Ac]) for WH which the efficiency was compared with the mentioned methods.

## EXPERIMENTAL

### Materials

#### *Water hyacinth preparation*

The plants were collected from a tank designed for propagation in the environmental lab (IPASA) at the Universiti Teknologi Malaysia, Johor. The WH leaves were separated and washed with distilled water, chopped, oven-dried, and then stored in

plastic bags for further usage. The chemicals used in this study, such as NaOH and H<sub>2</sub>SO<sub>4</sub>, were analytical grade reagents. The ionic liquid solution was bought from Sigma Aldrich Company (Kuala Lumpur, Malaysia).

## Methods

### *Different pretreatment methods*

In this study, four types of pretreatment methods were performed using acid, alkali, IL, and microwave-alkali. For the alkali treatment, 10 g of dried WH were soaked in an alkali solution for 1 h followed by a treatment time of 10 min at 150 °C inside of a furnace. Then, the hydrolysate was collected *via* a filtration method (Ahn *et al.* 2012). For the acid treatment, 10 g of WH were mixed with 90 mL of 4% dilute sulfuric acid (W/V) and then autoclaved at 120 °C for 15 min. Then, the treated biomass was neutralized with an addition of calcium hydroxide (3N) and filtered using a membrane filter (0.45 µm). The solid residues after pretreatment were washed with deionized water, then dried and stored for enzymatic saccharification (Fileto-Pérez *et al.* 2013).

The IL pretreatment was performed by mixing 51.0 mL of 1-ethyl-3-methylimidazolium acetate ([EMIM][Ac]) with 3 g of WH in a 17:1 ratio. The solution was heated and stirred at 120 °C and incubated for 180 min. To regenerate the WH, equal volumes of deionized water were added to the WH/solvent solution, which made the precipitation occur quickly. The sample was centrifuged and the supernatant containing IL was removed. Next, the precipitate was washed using deionized water. Finally, the regenerated WH was filtered using Whatman filter paper and oven-dried at 60 °C for 48 h in a vacuum (Gao *et al.* 2013a). The microwave-alkali pretreatment was conducted in a WX-4000 microwave digestion system (Hangzhou, China). In this pretreatment, the WH was treated with 1.0% (w/v) NaOH solution for 24 h at 45 °C in a shaker at 120 rpm, then heated for 1.0 min at 420 w in a microwave oven (SANYO Super Shower Wave 900W, Sanyo Denki America, Inc., Torrance, CA, USA) (Xia *et al.* 2013). FTIR analysis was performed using (Nicolet iS5, Thermo Fisher Scientific, Inc., USA). The spectra were obtained in the transmission mode and the range of 600 to 4000 cm<sup>-1</sup> at a resolution of 4 cm<sup>-1</sup> with 32 scans per each sample.

### *Preparation of yeast as inoculum*

A total of 5 g of yeast, *Saccharomyces cerevisiae* (*S. cerevisiae*) was added to 50 mL of distilled water and incubated at 37 °C with 150 rpm for 24 h. The potato dextrose agar medium (PDA) was prepared to culture the yeast cells. Then, *S. cerevisiae* was streaked on the PDA plates to grow the colonies and was kept in an incubator at 37 °C for 24 h. To enhance the growth rate and activity of *S. cerevisiae*, the best colonies in terms of size were screened and transferred to 50 mL of nutrient broth (NB) in a 250-mL Erlenmeyer flask at 30 °C.

Later, the plates were transferred to the shaker incubator in 37 °C for three days. A loopful of the colony was transferred to the test tube and mixed in 50 mL of distilled water. The tubes were centrifuged in 4000 rpm for 20 min (the temperature was held at -4 °C). The top layer (supernatant) was discarded and the pellet was collected to measure the optical density (OD) at 600 nm using a spectrophotometer (HACH DR6000; Loveland, CO, USA). The *S. cerevisiae* suspension cultures were used as an inoculum for fermentation (Mukhopadhyay and Chatterjee 2010).

*Simultaneous saccharification and fermentation (SSF)*

The SSF method was used for the enzymatic hydrolysis and fermentation of the treated WH biomass to produce ethanol. This process consisted of 5 g of the treated WH (previously autoclaved for 15 min at 121 °C), filter-sterilized cellulase (extracted from *Aspergillus niger*) solubilized in 0.1 M sodium phosphate, and 10 mL of *S. cerevisiae* in a Mendel medium. The initial pH of the fermentation medium was adjusted to 5.0, and the SSF was carried out at 37 °C in 250-mL conical flasks with 80 mL of fermentation medium and a final volume of 100 mL with agitation for 72 h. The sampling was completed at 12 h and approximately 10 mL of each sample was taken. The samples were centrifuged at 4000 rpm for 15 min and the pellet was discarded. The supernatant was separated and used to determine the ethanol and reducing sugar content (Eshtiaghi *et al.* 2012).

*Analytical procedures*

The WH composition (cellulose, hemicellulose, and lignin content) was measured according to Di Blasi *et al.* (1999). The 3,5-dinitrosalicylic acid (DNS) method was used to determine the total sugar reduction in the enzymatic hydrolysate of the acid-treated biomass (Miller 1959). The amount of ethanol during fermentation was measured using gas chromatography (GC) (Model 7890, Hewlett Packard, Waldbronn, Germany) with an FID detector and column HP20 M. The temperatures of the oven, column inlet, and detector were 200 °C, 300 °C, and 300 °C, respectively. The carrier gas was helium with a flow rate of 25 mL/min (Ang and Abd-Aziz 2015). The ethanol concentration in the samples was identified by comparing the area of the respective sample against the area of a reference sample containing only the standard ethanol. All of the experiments were conducted in triplicate. The theoretical yield value and volumetric ethanol productivity were calculated based on Eqs. 1 to 3:

$$\text{Volumetric ethanol productivity (Qp)} = \frac{\text{Ethanol Concentration}}{\text{Fermentation Time}} \quad (1)$$

$$\text{Theoretical Yield Value } (\eta, \%) = \frac{\text{Ethanol Yield}}{0.51} \times 100 \quad (2)$$

$$\text{Ethanol Yield } \left(\frac{\text{g}}{\text{g}}\right) = \frac{\text{Fermentation Medium (mL)}}{\text{Gram Biomass} \times 1000} \times \text{Ethanol Concentration} \quad (3)$$

*Theoretical maximum reducing sugar and ethanol yield*

For the calculation of the theoretical maximum reducing sugar yield, it was assumed that the cellulose and hemicellulose in the WH were completely hydrolyzed into hexose (glucose) and pentose (xylose). The weight ratio of the glucose molecule to cellulose was 1.111 (180/162), whereas that of xylose to hemicellulose was 1.136 (150/132). The reaction formula of cellulose conversion transformed into glucose, and the hydrolyze percentage was calculated based on Hu and Wen (2008).

**RESULTS AND DISCUSSION****Composition of Untreated WH**

The untreated WH consisted of  $16.4 \pm 0.35\%$  cellulose,  $32.7 \pm 0.14\%$  hemicellulose,  $5.7 \pm 0.05\%$  lignin, and  $10.7 \pm 0.1\%$  ash content. This demonstrated that WH could be a favorable carbohydrates source. However, Table 1 shows that previous studies obtained different amounts of carbohydrates and lignin from the WH biomass.

Various geographic conditions, different parts of the WH, different growth states, and methods for drying biomass could be reasons behind these differences.

**Table 1.** Chemical Composition of Untreated Water Hyacinth

Composition of WH	(Gao <i>et al.</i> 2013b)	(Cheng <i>et al.</i> 2014)	(Zhang <i>et al.</i> 2015)	(Das <i>et al.</i> 2016)	(Ruan <i>et al.</i> 2016)	This Study
Cellulose (%)	18.3	24.15	18.07	24.7	24.5	16.4 ± 0.35
Hemicellulose (%)	23.3	27.23	28.21	32.2	34.1	32.7 ± 0.14
Lignin (%)	17.7	12.39	7.03	3.2	8.6	5.7 ± 0.05

*Total reducing sugar from different pretreatment methods*

Manivannan and Narendhirakannan (2015) observed that the amount of glucose, xylose, and TRS range from 7 mg/g to 41 mg/g of WH. To evaluate the effectiveness of different pretreatment methods, the TRS was measured before and after pretreatment, as shown in Table 2.

**Table 2.** Reducing Sugars by Different Pretreatment Methods

WH Biomass	Untreated	Acid Treated	Alkali Treated	IL Treated	Microwave-alkali Treated
Total Reducing Sugars (mg/g of WH)	25 ± 1.5	51 ± 4.2	157 ± 8.2	45 ± 2.6	95 ± 3.1

The amount of total reducing sugars (hexoses and pentoses) released from the substrate equated to the effectiveness of the pretreatment method (Das *et al.* 2016). Based on Table 2, the highest rate of TRS obtained was from the alkali pretreatment at 157 mg/g of WH. This pretreatment enhanced the sugar ratio more than six times more in comparison to the untreated WH (25 mg/g). Similarly, Harun *et al.* (2011) obtained 24.7 mg sugar/g dry matter of sugars in untreated WH. In a study by Xu *et al.* (2016), 99 mg/g yield of TRS from untreated WH was produced during hydrolysis. The acid, IL, and microwave-alkali pretreatments produced 51 mg/g, 45 mg/g, and 95 mg/g WH of TRS, respectively. In conclusion, the highest TRS was obtained by the alkali-treated WH, followed by microwave-alkali-, acid-, and IL-treated WH, respectively.

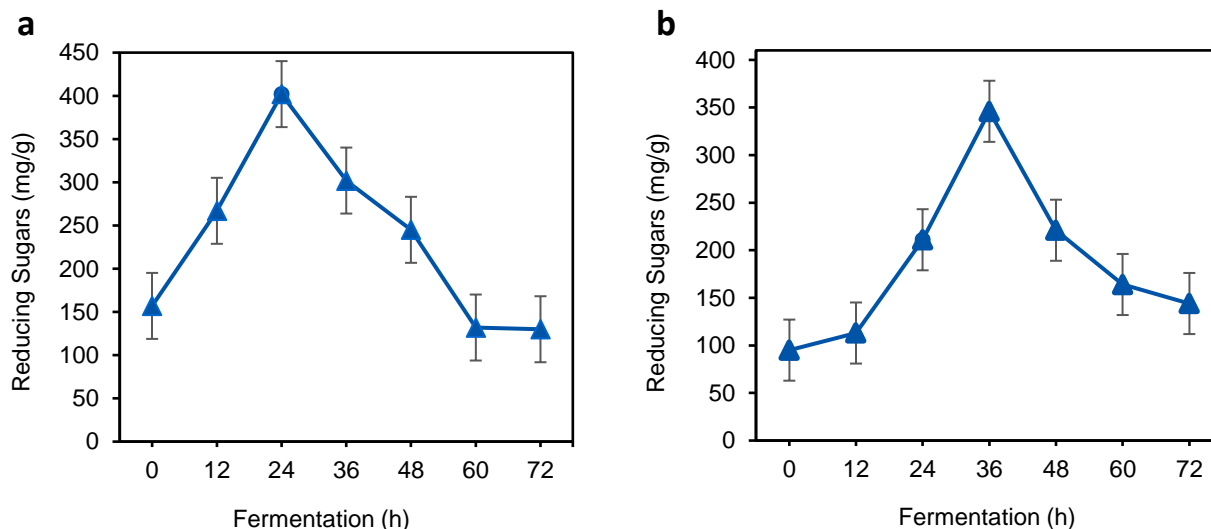
*TRS production during fermentation*

During fermentation, the majority of reducing sugars were converted to ethanol through the *S. cerevisiae* activity. Initially, the TRS was increased by the *cellulase* activity, and then decreased due to the sugar consumption by *S. cerevisiae* to produce ethanol. The high TRS production was due to breakage of the cellulosic structure by the *cellulase*, which resulted in the exposure of more sugars to microorganisms. In addition, the treated biomass in all pretreatment methods showed an increase in sugar amount in comparison to the untreated biomass. The pretreatments were effective in increasing the TRS during fermentation for ethanol production.

*TRS from alkali and microwave-alkali treated WH*

Based on Fig. 1a and Table 2, the highest reducing sugar was obtained from the alkali-treated WH. Initially, 157 mg/g of reducing sugars were obtained from the alkali-

treated sample and then were increased up to 402 mg/g after 24 h of fermentation. The reduction was observed afterwards due to the sugar consumption by the microorganisms until the end of the experiment. Similarly, Ganguly *et al.* (2013) observed 465 mg/g of TRS after an alkali pretreatment of WH. The high level of TRS was due to the availability of large amounts of monosaccharide (cellulose and hemicellulose) after the alkali pretreatment.

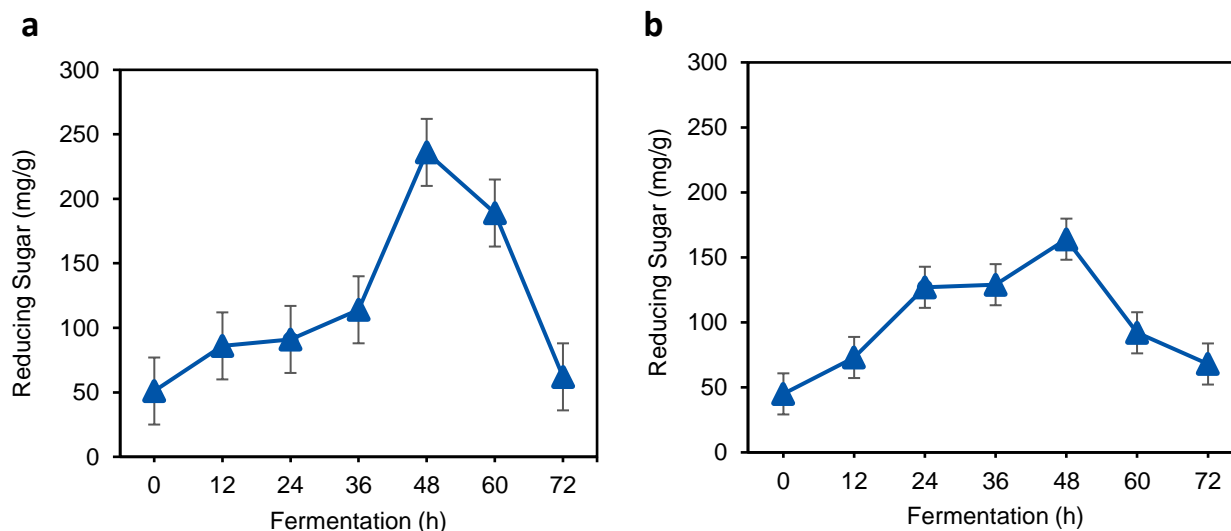


**Fig. 1.** TRS of alkali-treated and microwave-alkali-treated WH in fermentation: a) alkali-treated and b) microwave-alkali-treated

As shown in Fig. 1b, a sharp increment of TRS occurred from 95 mg/g to 346 mg/g in 36 h by the microwave-alkali-treated WH. The radiation successfully enhanced the amount of reducing sugars in the WH biomass. The TRS decreased sharply until the end of the fermentation at 72 h based on the *S. cerevisiae* activity in the fermentation flask and reached 144 mg/g at 72 h. Meanwhile, Zhang *et al.* (2015) obtained only up to 99.12 mg/g of TRS from the microwave-alkali-treated WH. Aswathy *et al.* (2010) observed the maximum reducing sugar yield from the hydrolysis of alkali-treated WH, which was 639.4 mg/g. In a study by Xia *et al.* (2013), the maximum reducing sugar yield of 483 mg/g from WH substrate, with 94.6% of the theoretical reducing sugar yield, was obtained using a microwave-alkali pretreatment. As stated by Singh and Bishnoi (2013), the alkali-treated WH has a higher amount of monosaccharide compared to the acid and oxidative pretreatment. They observed the reducing sugar amount was 5.1 g/L, 6.2 g/L, 9.3 g/L, 13.5 g/L, 11.3 g/L, and 8.8 g/L after 12 h, 24 h, 48 h, 60 h, 72 h, 96 h, and 120 h of fermentation, respectively.

#### *TRS from acid and IL treated water hyacinth*

As shown in Fig. 2a, the highest TRS from acid-treated WH was obtained after 48 h of fermentation at 236 mg/g of WH. After 48 h, the TRS was reduced to 62 mg/g at 72 h fermentation, which implied sugar consumption by microbes. Zhang *et al.* (2015) obtained 197.60 mg/g of TRS in hydrolysates from the acid-treated WH. Meanwhile, 3.79 g/L to 5.27 g/L of total reducing sugars was obtained from the alkali-treated WH using *S. cerevisiae* during fermentation (Manivannan and Narendhirakannan 2015).



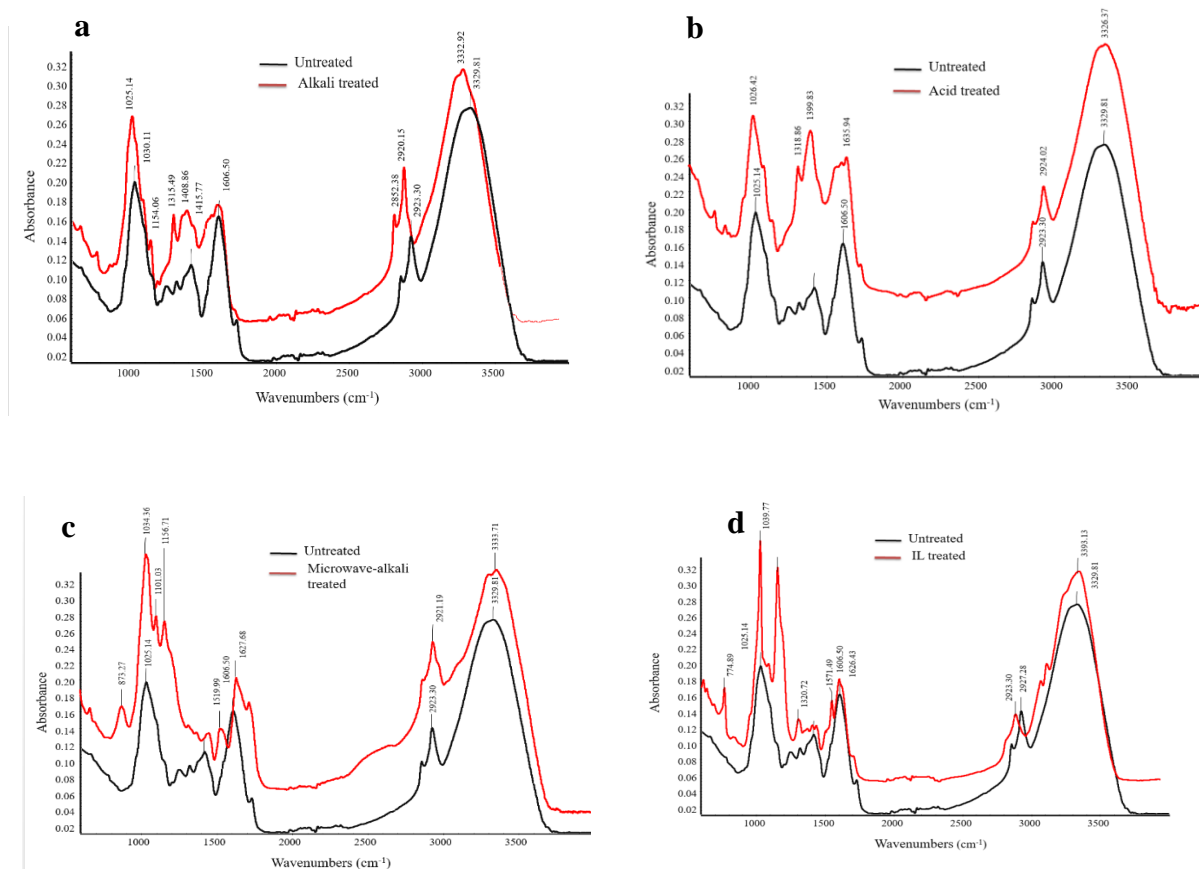
**Fig. 2.** TRS of acid-treated and IL-treated WH in fermentation: a) acid-treated and b) IL-treated

Similar to other pretreatment methods, the reduction of TRS in the IL-treated sample occurred between 48 h and 72 h. During this stage, the depletion of sugars occurred in the fermentation due to microorganism activity. The highest TRS was 164 mg/g at 48 h, and it was reduced to 68 mg/g at the end of fermentation at 72 h (Fig. 2b). In this study, only 164 mg/g of TRS were obtained from the IL-treated WH, which could have been related to the level of lignin in the substrate or type of IL. In contrast, high levels of TRS at 584 mg/g were obtained from that IL-treated WH 1-butyl-3-methylimidazolium acetate (BMIMA) (Guragain *et al.* 2011).

#### FTIR analysis

For better validation of the effects of various pretreatment methods in the structure of WH, the analysis of FTIR spectra before and after the pretreatment is shown in Fig. 3. After alkali pretreatment of WH, in the range of  $1000\text{ cm}^{-1}$  to  $1500\text{ cm}^{-1}$ , five peaks were appeared  $1025\text{ cm}^{-1}$ ,  $1154\text{ cm}^{-1}$ ,  $1315\text{ cm}^{-1}$ ,  $1408\text{ cm}^{-1}$ , and  $1415\text{ cm}^{-1}$  (Fig. 3a). These peaks represented the structural features of cellulose and hemicelluloses in WH biomass. Based on the results of this study, the alkali pretreatment increased the amount of cellulose and hemicellulose in WH. In addition, a significant drop of band intensity was observed at  $2915\text{ cm}^{-1}$ , which indicated the reduction in the hydrogen bonding of cellulose in WH structure. In contrast, the absorption band at  $2940\text{ cm}^{-1}$  after alkali pretreatment and enzymatic hydrolysis of WH was observed, as reported by Das *et al.* (2015)

Based on FTIR spectra of acid treated WH, no peaks in the range of  $> 1000$  were observed. This means that the acid pretreatment only increased the cellulose of WH and was not effective in lignin removal (Fig. 3b). Although, Cheng *et al.* (2014) found the peak intensity of  $894\text{ cm}^{-1}$ , which confirms the increase of cellulose after acid treatment of WH due to the removal of amorphous hemicellulose. In addition, because of the complete hydrolysis of hemicellulose, the peak intensity of  $1739\text{ cm}^{-1}$  was remarkably decreased after acid pretreatment. FTIR spectra showed significant reduction of band intensity at  $1321\text{ cm}^{-1}$  (C–H deformation in hemicelluloses) due to the removal of hemicelluloses.



**Fig. 3.** FTIR analysis of different pretreatments a) Alkali treated WH b) Acid treated WH c) Microwave-alkali treated WH d) IL treated WH

FTIR spectra of microwave-alkali treated WH showed that the disruption of lignin structure (Fig. 3c). Furthermore, the peak intensity of  $1630\text{ cm}^{-1}$  in C=O of acetyl groups of lignin dropped due to the lignin cleavage. Furthermore, it also confirmed that the microwave irradiation was effective for complete hydrolysis of tough WH structure (de Souza Moretti *et al.* 2014).

In FTIR spectra of IL treated WH there was a sharp band at  $774\text{ cm}^{-1}$ , which corresponds to the C1 group frequency and can be attributed to the  $\beta$ -glycosidic linkages (1 $\rightarrow$ 4) between xylose units in hemicelluloses. Another peak appeared at  $1,240\text{ cm}^{-1}$  corresponding to (C–O stretching in lignin and hemicellulose), which was indicative of the removal of hemicellulose (Fig. 4d).

Similarly, 2922, 1427, and  $1373\text{ cm}^{-1}$  bands were related to the characteristic of cellulose and did not substantially change due to the protection of cellulose during IL pretreatment (Xia *et al.* 2013). Furthermore, IL pretreatment had more contribution in disruption of  $\beta$ -glycosidic linkages and hemicelluloses reduction. Recently, Singh *et al.* (2018) showed that treatment of WH using different imidazolium-based ILs such as [1-alkyl]-3-methylimidazolium bromide, [Cnmim]Br, resulted in high levels of cellulose disruption.



## Ethanol Production from Differently Treated WH

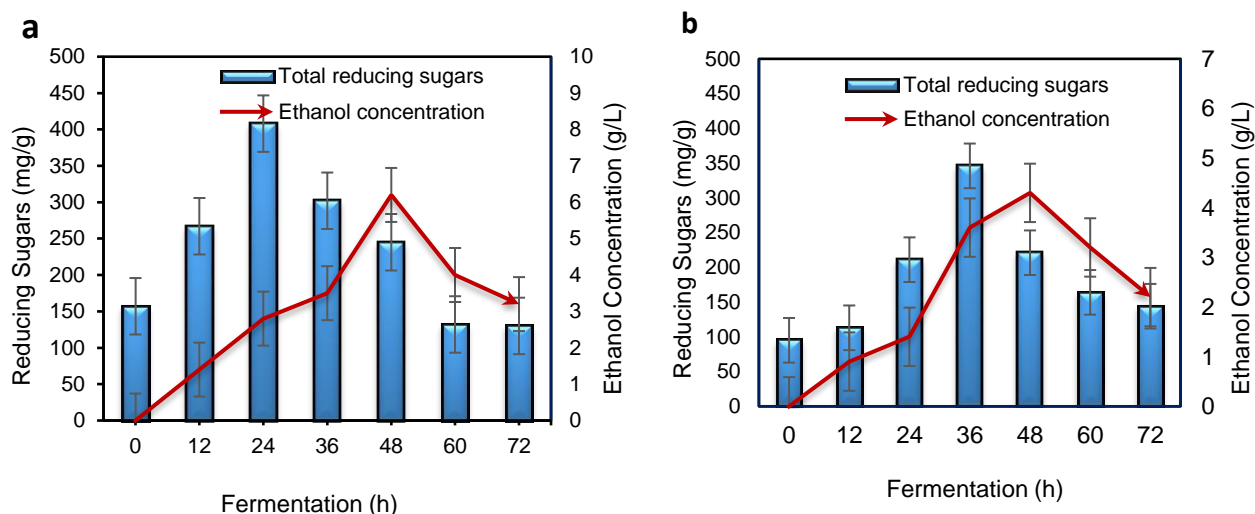
### Ethanol production from alkali- and microwave-alkali treated WH

Alkali and microwave-alkali pretreatment are two effective methods for ethanol production from WH. Table 3 shows a summary of some studies that used alkali treated WH in different NaOH concentration by using *S. cerevisiae*.

**Table 3.** Ethanol production from alkali-treated WH in different studies

NaOH concentration	Yeast type	Fermentation (h)	Ethanol yield (g/L)	Reference
1.0%	<i>S. cerevisiae</i> and <i>Pachysolen tannophilus</i>	48	5.2	(Mukhopadhyay and Chatterjee 2010)
5.0%	<i>S. cerevisiae</i>	36	4.3 ± 0.8	(Singh and Bishnoi 2013)
5.0%	<i>S. cerevisiae</i>	48	3.07	(Ganguly <i>et al.</i> 2013)
5.0%	<i>S. cerevisiae</i>	72	6.76	(Das <i>et al.</i> 2015)

As shown in Fig. 4a and Table 2, the alkali-treated WH had the highest TRS at 402 mg/g and the highest ethanol was produced at 48 h of fermentation by  $6.2 \pm 0.4$  g/L.

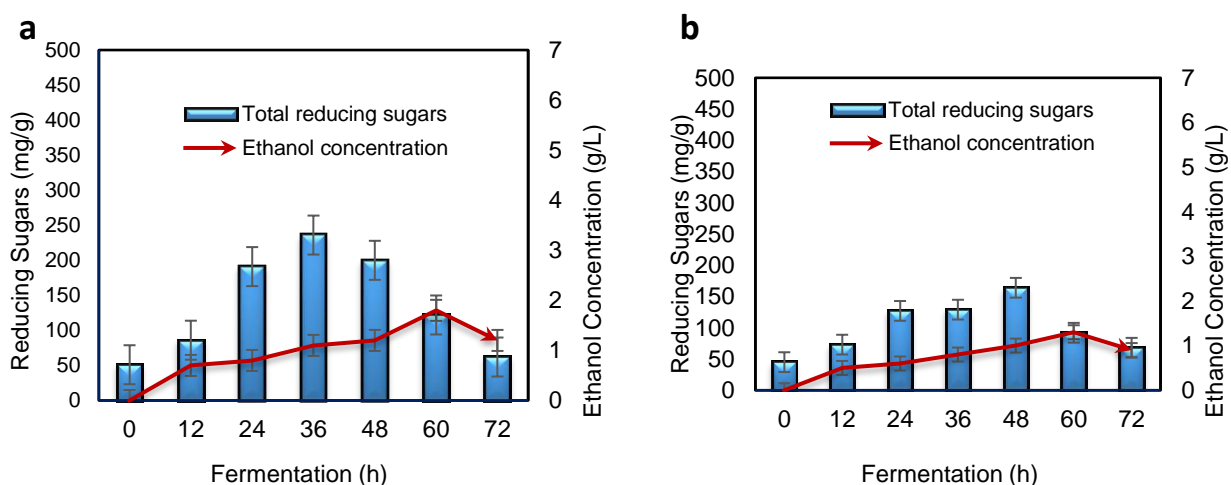


**Fig. 4.** Ethanol production: a) alkali-treated WH and b) microwave-alkali-treated WH

Das *et al.* (2015) found that the higher ethanol yield by *P. stipitis* was due to the utilization of both pentose and hexose. Ethanol production based on sugar consumption in the alkali-treated WH is shown in Fig. 4a. The ethanol production from the microwave-alkali-treated WH based on TRS is shown in Fig. 4b. By using microwave-alkali-treated WH,  $4.3 \pm 0.5$  g/L of ethanol with an ethanol yield of 0.86 g/5g of WH and 0.059 of volumetric ethanol productivity in 48 h of fermentation was produced. Meanwhile, the highest amount of TRS was 346 mg/g in 36 h. The ethanol production *via* microwave-alkali increased sharply after 24 h to 48 h. Therefore, the constant reduction occurred due to the low rate of TRS production during fermentation.

### Ethanol production from acid and IL treated WH

Figure 5a shows the ethanol production from the acid-treated WH based on TRS and fermentation time. The acid-treated WH produced  $1.8 \pm 0.8$  g/L of ethanol with a 0.025 productivity and yield of 0.03 g/g of WH in 60 h of fermentation. In addition, the sugar level was drastically decreased because of the TRS consumption by the microbes for ethanol production. In this study, 4% (w/v)  $\text{H}_2\text{SO}_4$  was applied and 236 mg/g of TRS were obtained at 36 h of fermentation. In previous studies, 33.3 g/L of total sugars were obtained in optimized conditions by Reales-Alfaro *et al.* (2013) and 21 g/g by Nigam (2002) after pretreatments with sulfuric acid. These findings further support the idea that the amount of sugars released from the substrate depends on the pretreatment method efficiency and the enzyme activity. As *cellulase* cleaves the cellobiose units to liberate glucose due to high  $\beta$ -glucosidases activity about 21.39 (U/gDS), it effectively enhanced the TRS production and subsequently some end products such as ethanol. As reported by Alvira *et al.* (2010), the presence of ILs residues could have a significant negative effect on *cellulase* activity. Therefore, to prevent the decrease of final sugars concentrations, ILs removal is necessary.



**Fig. 5.** Ethanol production: a) acid-treated WH and b) IL-treated WH

In this study, the lowest ethanol yield and TRS production were obtained from the IL-treated WH. Hence, there are not many published studies using IL pretreatment for ethanol production from WH. It could hinder the enzymatic hydrolysis as well, as reported by Guragain *et al.* (2011). The reason could be due to a high moisture content of WH, which leads to a reaction of water molecules with IL solvents during fermentation. Besides, some small molecules such as fructose and lignin can be dissolved in IL solutions during the pretreatment process (Gao *et al.* 2013a).

Based on Fig. 5b, the IL-treated WH produced  $1.3 \pm 0.9$  g/L of ethanol with a volumetric productivity of 0.018 and a yield of 0.02 g/5g of WH. The highest ethanol production was obtained at 60 h, and the highest TRS was obtained at 48 h. As demonstrated by Guragain *et al.* (2011), by using a dilute acid pretreatment on WH, the hydrolysis yields of glucose and total reducing sugars were 2.1 and 3.3 times higher than IL pretreatments. Glucose and total sugars yield of the IL pretreatments were 332 mg/g and 584 mg/g of WH, respectively. One of the reasons for such results could have been due to the solubilization of the majority of all hemicelluloses in dilute acid pretreatments.

### Theoretical maximum reducing sugar and ethanol yield

In this study, a maximum theoretical yield of reducing sugar for untreated WH was  $(18.2 + 37.2) = 0.554$  g/g of untreated WH biomass. Theoretically, 0.51 kg of ethanol per kilogram can be obtained from glucose and xylose. Therefore, the maximum theoretical ethanol yield of WH was  $(0.554 \times 0.51) = 0.282$  g/g of raw biomass of WH. However, Das *et al.* (2015) state the maximum theoretical ethanol yield as 0.3659 g/g of WH, which was 0.294 g/g of WH as reported by Cheng *et al.* (2014). This is comparable to ethanol yields of other lignocellulosic materials, such as rice straw with 0.326 g/g (Imman *et al.* 2015). Table 4 shows the ethanol productivity and yield by different types of pretreatment methods. Conditions of fermentation, species, and carbohydrate content of WH notably contributed to ethanol production.

**Table 4.** Ethanol Production from Differently Treated WH

Different Pretreatments	Highest Level of TRS (g/g)	Highest Ethanol (h)	Ethanol Production (gp/L)	Volumetric Ethanol Productivity	Yield (g Ethanol/ 5 g WH)
Alkali	402	48	$6.2 \pm 0.4$	0.086	0.12
Microwave-alkali	346	48	$4.3 \pm 0.5$	0.059	0.086
Acid	234	60	$1.8 \pm 0.8$	0.025	0.03
Ionic Liquid	164	60	$1.3 \pm 0.9$	0.018	0.02

## CONCLUSIONS

1. The IL pretreatment with 1-ethyl-3-methylimidazolium acetate ([EMIM][Ac]) was not effective for sugar and ethanol production from WH.
2. The most effective pretreatment method was alkali followed by microwave-alkali, acid, and IL.
3. FTIR analysis showed a decrease of the band at  $1430\text{ cm}^{-1}$  in all treated samples, which confirmed the effective disruption of the crystalline structure of WH.
4. The highest ethanol yield was  $6.2 \pm 0.4$  g/L *via* alkali-treated biomass with 0.086 volumetric ethanol productivity and an ethanol yield of 0.012 g/5g of WH in 48 h.
5. The acid, microwave-alkali, and IL-treated WH produced  $4.3 \pm 0.5$  g/L,  $1.8 \pm 0.8$  g/L, and  $1.3 \pm 0.9$  g/L of ethanol, respectively.

## ACKNOWLEDGMENTS

This research was supported by a grant (2016R1D1A1A02937267) provided by Korean National Research Foundation (KNRF) which is part of project individual basic science & engineering research program (SGER).

## REFERENCES CITED

- Ahn, D. J., Kim, S. K., and Yun, H. S. (2012). "Optimization of pretreatment and saccharification for the production of bioethanol from water hyacinth by *Saccharomyces cerevisiae*," *Bioprocess Biosystem Engineering* 35(1-2), 35-41. DOI: 10.1007/s00449-011-0600-5
- Alvira, P., Tomás-Pejó, E., Ballesteros, M. J., and Negro, M. J. (2010). "Pretreatment technologies for an efficient bioethanol production process based on enzymatic hydrolysis: a review," *Bioresource Technology* 101(13), 4851-4861. DOI: 10.1016/j.biortech.2009.11.093
- Ang, S. K., and Abd-Aziz, S. (2015). "Potential uses of xylanase-rich lignocellulolytic enzymes cocktail for oil palm trunk (OPT) degradation and lignocellulosic ethanol production," *Energy & Fuels* 29(8), 5103-5116. DOI: 10.1021/acs.energyfuels.5b00891
- Aswathy, U. S., Sukumaran, R. K., Devi, G. L., Rajasree, K. P., Singhanian, R. R., and Pandey, A. (2010). "Bio-ethanol from water hyacinth biomass: An evaluation of enzymatic saccharification strategy," *Bioresource Technology* 101(3), 925-930. DOI: 10.1016/j.biortech.2009.08.019
- Baeyens, J., Kang, Q., Appels, L., Dewil, R., Lv, Y., and Tan, T. (2015). "Challenges and opportunities in improving the production of bio-ethanol," *Progress in Energy and Combustion Science* 47, 60-88. DOI: 10.1016/j.peccs.2014.10.003
- Barua, V. B., and Kalamdhad, A. S. (2017). "Effect of various types of thermal pretreatment techniques on the hydrolysis, compositional analysis and characterization of water hyacinth," *Bioresource Technology* 227, 147-154. DOI: 10.1016/j.biortech.2016.12.036
- Bhatt, S. M., and Shilpa (2014). "Lignocellulosic feedstock conversion, inhibitor detoxification and cellulosic hydrolysis: A review," *Biofuels* 5(6), 633-649. DOI: 10.1080/17597269.2014.1003702
- Cheng, J., Wang, X., Huang, R., Liu, M., Yu, C., and Cen, K. (2014). "Producing ethanol from water hyacinth through simultaneous saccharification and fermentation with acclimatized yeasts," *BioResources* 9(4), 7666-7680. DOI: 10.15376/biores.9.4.7666-7680
- Das, S. P., Gupta, A., Das, D., and Goyal, A. (2016). "Enhanced bioethanol production from water hyacinth (*Eichhornia crassipes*) by statistical optimization of fermentation process parameters using Taguchi orthogonal array design," *International Biodeterioration & Biodegradation* 109, 174-184. DOI: 10.1016/j.ibiod.2016.01.008
- Das, S., Bhattacharya, A., Haldar, S., Ganguly, A., Gu, S., Ting, Y. P., and Chatterjee, P. K. (2015). "Optimization of enzymatic saccharification of water hyacinth biomass for bio-ethanol: Comparison between artificial neural network and response surface methodology," *Sustainable Materials and Technologies* 3, 17-28. DOI: 10.1016/j.susmat.2015.01.001
- de Souza Moretti, M. M., Bocchini-Martins, D. A., Nunes, C. D. C. C., Villena, M. A., Perrone, O. M., da Silva, R., Boscolo, M., and Gomes, E. (2014). "Pretreatment of sugarcane bagasse with microwaves irradiation and its effects on the structure and on enzymatic hydrolysis," *Applied Energy* 122, 189-195. DOI: 10.1016/j.apenergy.2014.02.020

- Di Blasi, C., Signorelli, G., Di Russo, C., and Rea, G. (1999). "Product distribution from pyrolysis of wood and agricultural residues," *Industrial & Engineering Chemistry Research* 38(6), 2216-2224. DOI: 10.1021/ie980711u
- Dionisi, D., Anderson, J. A., Aulenta, F., McCue, A., and Paton, G. (2015). "The potential of microbial processes for lignocellulosic biomass conversion to ethanol: A review," *Journal of Chemical Technology and Biotechnology* 90(3), 366-383. DOI: 10.1002/jctb.4544
- Elgharbawy, A. A., Alam, M. Z., Moniruzzaman, M., and Goto, M. (2016). "Ionic liquid pretreatment as emerging approaches for enhanced enzymatic hydrolysis of lignocellulosic biomass," *Biochemical Engineering Journal* 109, 252-267. DOI: 10.1016/j.bej.2016.01.021
- Eshtiaghi, M. N., Yoswathana, N., Kuldiloke, J., and Ebadi, A. G. (2012). "Preliminary study for bioconversion of water hyacinth (*Eichhornia crassipes*) to bioethanol," *African Journal of Biotechnology* 11(21), 4921-4928.
- Fileto-Pérez, H. A., Rutiaga-Quiñones, J. G., Aguilar-González, C. N., Paéz, J. B., López, J., and Rutiaga-Quiñones, O. M. (2013). "Evaluation of *Eichhornia crassipes* as an alternative raw material for reducing sugars production," *BioResources* 8(4), 5340-5348. DOI: 10.15376/biores.8.4.5340-5348
- Ganguly, A., Halder, S., Laha, A., Saha, N., Chatterjee, P. K., and Dey, A. (2013). "Effect of alkali pretreatment on water hyacinth biomass for production of ethanol," *Advanced Chemical Engineering Research* 2(2), 40-44.
- Gao, J., Chen, L., Yan, Z., and Wang, L. (2013a). "Effect of ionic liquid pretreatment on the composition, structure and biogas production of water hyacinth (*Eichhornia crassipes*)," *Bioresource Technology* 132, 361-364. DOI: 10.1016/j.biortech.2012.10.136
- Gao, J., Chen, L., Yuan, K., Huang, H., and Yan, Z. (2013b). "Ionic liquid pretreatment to enhance the anaerobic digestion of lignocellulosic biomass," *Bioresource Technology* 150, 352-358. DOI: 10.1016/j.biortech.2013.10.026
- Guragain, Y. N., De Coninck, J., Husson, F., Durand, A., and Rakshit, S. K. (2011). "Comparison of some new pretreatment methods for second generation bioethanol production from wheat straw and water hyacinth," *Bioresource Technology* 102(6), 4416-4424. DOI: 10.1016/j.biortech.2010.11.125
- Harun, M. Y., Radiah, A. D., Abidin, Z. Z., and Yunus, R. (2011). "Effect of physical pretreatment on dilute acid hydrolysis of water hyacinth (*Eichhornia crassipes*)," *Bioresource Technology* 102(8), 5193-5199. DOI: 10.1016/j.biortech.2011.02.001
- Hu, Z., and Wen, Z. (2008). "Enhancing enzymatic digestibility of switchgrass by microwave-assisted alkali pretreatment," *Biochemical Engineering Journal* 38(3), 369-378. DOI: 10.1016/j.bej.2007.08.001
- Imman, S., Arnthong, J., Burapatana, V., Champreda, V., and Laosiripojana, N. (2015). "Fractionation of rice straw by a single-step solvothermal process: Effects of solvents, acid promoters, and microwave treatment," *Renewable Energy* 83, 663-673. DOI: 10.1016/j.renene.2015.04.062
- Jambo, S. A., Abdulla, R., Azhar, S. H. M., Marbawi, H., Gansau, J. A., and Ravindra, P. (2016). "A review on third generation bioethanol feedstock," *Renewable & Sustainable Energy Reviews* 65, 756-769. DOI: 10.1016/j.rser.2016.07.064
- Mandade, P., Bakshi, B. R., and Yadav, G. D. (2016). "Ethanol from Indian agro-industrial lignocellulosic biomass: An energy evaluation," *Clean Technologies and Environmental Policy* 18(8), 2625-2634. DOI: 10.1007/s10098-016-1179-y

- Manivannan, A., and Narendhirakannan, R. T. (2015). Bioethanol production from aquatic weed water hyacinth (*Eichhornia crassipes*) by yeast fermentation,” *Waste and Biomass Valorization* 6(2), 209-216. DOI: 10.1007/s12649-015-9347-6
- Miller, G. L. (1959). “Use of dinitrosalicylic acid reagent for determination of reducing sugar,” *Analytical Chemistry* 31(3), 426-428.
- Mukhopadhyay, S. B., and Chatterjee, N. C. (2010). “Bioconversion of water hyacinth hydrolysate into ethanol,” *BioResources* 5(2), 1301-1310. DOI: 10.15376/biores.5.2.1301-1310
- Narra, M., Divecha, J., Shah, D., Balasubramanian, V., Vyas, B., Harijan, M., and Macwan, K. (2017). “Cellulase production, simultaneous saccharification and fermentation in a single vessel: A new approach for production of bio-ethanol from mild alkali pre-treated water hyacinth,” *Journal of Environmental Chemical Engineering* 5(3), 2176-2181. DOI: 10.1016/j.jece.2017.04.043
- Nigam, J. N. (2002). “Bioconversion of water-hyacinth (*Eichhornia crassipes*) hemicellulose acid hydrolysate to motor fuel ethanol by xylose-fermenting yeast,” *Journal of Biotechnology* 97(2), 107-116. DOI: 10.1016/S0168-1656(02)00013-5
- Reales-Alfaro, J. G., Trujillo-Daza, L. T., Arzuaga-Lindado, G., Castaño-Peláez, H. I., and Polo-Córdoba, Á. D. (2013). “Acid hydrolysis of water hyacinth to obtain fermentable sugars,” *CT&F-Ciencia, Tecnología y Futuro* 5(2), 101-111.
- Rezania, S., Din, M. F. M., Kamaruddin, S. F., Taib, S. M., Singh, L., Yong, E. L., and Dahalan, F. A. (2016). “Evaluation of water hyacinth (*Eichhornia crassipes*) as a potential raw material source for briquette production,” *Energy* 111, 768-773. DOI: 10.1016/j.energy.2016.06.026
- Rezania, S., Din, M. F. M., Mohamad, S. E., Sohaili, J., Taib, S. M., Yusof, M. B. M., and Ahsan, A. (2017). “Review on pretreatment methods and ethanol production from cellulosic water hyacinth,” *BioResources* 12(1), 2108-2124. DOI: 10.15376/biores.12.1.2108-2124
- Rezania, S., Din, M. F. M., Taib, S. M., Mohamad, S. E., Dahalan, F. A., Kamyab, H., and Ebrahimi, S. S. (2018). “Ethanol production from water hyacinth (*Eichhornia crassipes*) using various types of enhancers based on the consumable sugars,” *Waste and Biomass Valorization* 9(6), 939-946. DOI: 10.1007/s12649-017-9883-3
- Rezania, S., Ponraj, M., Din, M. F. M., Songip, A. R., Sairan, F. M., and Chelliapan, S. (2015). “The diverse applications of water hyacinth with main focus on sustainable energy and production for new era: An overview,” *Renewable & Sustainable Energy Reviews* 41, 943-954. DOI: 10.1016/j.rser.2014.09.006
- Ruan, T., Zeng, R., Yin, X. Y., Zhang, S. X., and Yang, Z. H. (2016). “Water hyacinth (*Eichhornia crassipes*) biomass as a biofuel feedstock by enzymatic hydrolysis,” *BioResources* 11(1), 2372-2380. DOI: 10.15376/biores.11.1.2372-2380
- Sarto, S., Hildayati, R., and Syaichurrozi, I. (2019). “Effect of chemical pretreatment using sulfuric acid on biogas production from water hyacinth and kinetics,” *Renewable Energy* 132, 335-350. DOI: 10.1016/j.renene.2018.07.121
- Singh, A., and Bishnoi, N. R. (2012). “Optimization of ethanol production from microwave alkali pretreated rice straw using statistical experimental designs by *Saccharomyces cerevisiae*,” *Industrial Crops and Products* 37(1), 334-341. DOI: 10.1016/j.indcrop.2011.12.033
- Singh, A., and Bishnoi, N. R. (2013). “Comparative study of various pretreatment techniques for ethanol production from water hyacinth,” *Industrial Crops and Products* 44, 283-289. DOI: 10.1016/j.indcrop.2012.11.026

- Singh, J., Suhag, M., and Dhaka, A. (2015). "Augmented digestion of lignocellulose by steam explosion, acid and alkaline pretreatment methods: A review," *Carbohydrate Polymers* 117, 624-631. DOI: 10.1016/j.carbpol.2014.10.012
- Singh, J., Sharma, R., Ghosh, P., Kumar, A., and Khan, M. L. (2018). "Imidazolium based ionic liquids: A promising green solvent for water hyacinth biomass deconstruction," *Frontiers in Chemistry* 6, 548. DOI: 10.3389/fchem.2018.00548
- Srivastava, N., Rawat, R., Singh Oberoi, H., and Ramteke, P. W. (2015). "A review on fuel ethanol production from lignocellulosic biomass," *International Journal of Green Energy* 12(9), 949-960. DOI:
- Sun, S., Sun, S., Cao, X., and Sun, R. (2016). "The role of pretreatment in improving the enzymatic hydrolysis of lignocellulosic materials," *Bioresource Technology* 199, 49-58. DOI: 10.1016/j.biortech.2015.08.061
- Tye, Y. Y., Lee, K. T., Abdullah, W. N. W., and Leh, C. P. (2016). "The world availability of non-wood lignocellulosic biomass for the production of cellulosic ethanol and potential pretreatments for the enhancement of enzymatic saccharification," *Renewable & Sustainable Energy Reviews* 60, 155-172. DOI: 10.1016/j.rser.2016.01.072
- Xia, A., Cheng, J., Song, W., Yu, C., Zhou, J., and Cen, K. (2013). "Enhancing enzymatic saccharification of water hyacinth through microwave heating with dilute acid pretreatment for biomass energy utilization," *Energy* 61, 158-166. DOI: 10.1016/j.energy.2013.09.019
- Xu, F., Chen, L., Wang, A., and Yan, Z. (2016). "Influence of surfactant-free ionic liquid microemulsions pretreatment on the composition, structure and enzymatic hydrolysis of water hyacinth," *Bioresource Technology* 208, 19-23. DOI: 10.1016/j.biortech.2016.02.027
- Zhang, Q., Wei, Y., Han, H., and Weng, C. (2018). "Enhancing bioethanol production from water hyacinth by new combined pretreatment methods," *Bioresource Technology* 251, 358-363. DOI: 10.1016/j.biortech.2017.12.085
- Zhang, Q., Weng, C., Huang, H., Achal, V., and Wang, D. (2015). "Optimization of bioethanol production using whole plant of water hyacinth as substrate in simultaneous saccharification and fermentation process," *Frontiers in Microbiology* 6, 1411-1419. DOI: 10.3389/fmicb.2015.01411
- Zheng, Y., Zhao, J., Xu, F., and Li, Y. (2014). "Pretreatment of lignocellulosic biomass for enhanced biogas production," *Progress in Energy and Combustion Science* 42, 35-53. DOI: 10.1016/j.pecs.2014.01.001

Article submitted: August 9, 2018; Peer review completed: November 18, 2018; Revised version received and accepted: November 23, 2018; Published: November 30, 2018.  
DOI: 10.15376/biores.14.1.592-606