

Essential Oil Extracted from Wood of *Phoebe bournei* (Hemsl.) Yang: Chemical Constituents, Antitumor, Antibacterial, and Hypoglycemic Activities

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The essential oil was extracted from the wood of *Phoebe bournei* (Hemsl.) Yang by a hydrothermal method and then analyzed by gas chromatography-mass spectrometry. The wood oil was comprised mainly of α -copaene (5.44%), α -muurolene (7.32%), δ -cadinene (11.44%), and 1s-calamenene (5.18%). *P. bournei* wood essential oil exhibited significant antifungal properties against *Candida albicans*, *Trichophyton rubrum*, *Epidermophyton floccosum*, and *Microsporium gypseum*, and it showed good potential antitumor activity against leukemia cell lines. Although *P. bournei* (Hemsl.) Yang wood essential oil exhibited weaker activity against the four tested bacteria, it had some effect on promoting glucose uptake by adipocytes.

Keywords: Essential oil; Chemical constituents; Antitumor activity; Antifungal activity; Hypoglycemic activity

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INTRODUCTION

Phoebe bournei (Hemsl.) Yang, an evergreen tree in the Lauraceae family, is a world-renowned precious tree species used for timber production and landscaping and a national second-class protected plant (Liu *et al.* 2014b). Historically, *P. bournei*, *C. camphora* (L.) Presl, *C. ovata* G. Don, and *C. glavca* (Thunb.) Oerst were called the four famous woods (Chen and Xie 2014). *P. bournei* is dry and straight, with a magnificent tree crown, vertical wood texture, fine structure, no warping and deformation, and resistance to insects and corrosion, making it a good material for high-grade buildings and fine furniture (Gao *et al.* 2016). It is also the main tree species of *Phoebe jinsinan*, one of the ancient exclusive royal woods, known as "Emperor Wood" (He *et al.* 2013).

Due to its enormous economic value and wide range of uses, *P. bournei* has suffered severe damage (Liu *et al.* 2014a). Currently, it is only sporadically distributed in mountainous, evergreen broad-leaved forests 200 to 1000 meters above sea level in Zhejiang, Fujian, Guangxi, and Guizhou provinces of China (Zhang and Iwatsuki 2013). Plant essential oil is a secondary metabolite of plants and is widely used in pest control, cosmetics, food, medicine and other fields (Bedini *et al.* 2016; Perdones *et al.* 2016; Salahfatnassi *et al.* 2016). Studies have shown that *Phoebe zhennan* plays an important role in the development of natural aromatic resources. *Phoebe zhennan* seed oil can be used as raw material for cosmetics production (Ren *et al.* 1990) and its wood essential oil has certain bactericidal and antitumor activity. According to the ancient Chinese medical book

"Compendium of Materia Medica" "Taiping Shenghui Fang", Nanmu is used as a medicine, which can treat stomach diseases, cholera, tendons, otitis media (otitis), athlete's foot and other diseases. At present, studies on *P. bournei* are limited to biological characteristics (Zhou *et al.* 2005; Zhang *et al.* 2012), wood properties (Jiang *et al.* 2008), nursery techniques (Huang *et al.* 2016; Zhang *et al.* 2016a, b), and artificial forest management (Shao *et al.* 2014; Huang *et al.* 2017; Yu and Chen 2017). Research on the chemical constituents and biological activity of the essential oil and other aspects had not been carried out yet.

The wood structures of *Phoebe* Ness and its closely related species are very similar. It is difficult to identify the species accurately by traditional methods. However, there is a high demand for the identification of the species of the jinsinan wood in the timber market (Yang *et al.* 2018). Also there is a need for reliable forensic identification of the species. Studies have shown that there are differences in the chemical composition of the *P. bournei* wood and other species in the same genus (Ding *et al.* 2017b; Wang *et al.* 2017; Xue and Zhao 2017). This study tested and analyzed the chemical composition of *P. bournei* essential oil, in order to provide a reference for the identification of *P. bournei*, regulate the trade of jinsinan wood, and promote the prosperity and development of the market.

EXPERIMENTAL

Extraction of Essential Oil

The experimental raw material *P. bournei* was provided by Yiming Wood, Rongshui County, Guangxi Province. The age of the tree was 1700 years. Wood samples were collected at 1.3 m above the ground. The moisture content of the wood was 16.8%.

The essential oil was extracted from the wood of *P. bournei* by a hydrothermal method. First, a crushed sample of *P. bournei* weighing 200 ± 0.01 g was accurately measured using an analytical balance and added to a 2000 mL round-bottomed flask. Next, distilled water (1200 mL) was introduced, and several glass beads were added to prevent bumping. The extraction apparatus was assembled and heated for 5 h (until the volume of the essential oil remained constant). After settling for 0.5 h, the volume of the extract was recorded, with three replications.

Instrumentation

The instruments used were an Agilent 7890A-5975C gas chromatography–mass spectrometry (GC–MS) system (Hercules, CA, USA), an Agilent 7890A gas chromatography (GC) system, a steam distiller supplied by Shuniu Glassware Co., Ltd (Chengdu, China), a Sartorius CP224S analytical balance (Sartorius, Göttingen, Germany), and a multifunctional enzyme marker Multiskan FC (Thermo, Waltham, MA, USA).

Gas Chromatography Analysis

An Agilent HP-5 chromatography column (30 m × 0.25 mm × 0.50 μm) was used. The sample injection volume was 1 μL, the split ratio was 50:1, the detector temperature was 250 °C, and the carrier gas used was ultra-pure helium. The oven temperature program was as follows: 120 °C for 3 min, increased to 140 °C at 5 °C/min over 5 min, then increased to 160 °C at 2 °C/min over 10 min. The flow rate was 1 mL/min, and the machine was equipped with a flame ionization detector. The components in the essential oil were identified by retention indices that were determined with C7–C20 alkane standards as the

reference and that were confirmed by GC–MS.

Gas Chromatography–Mass Spectrometry Analysis

The gas chromatography conditions are detailed above.

The ion source for the mass spectrography was electron ionization. The ionization energy was 70eV, the auxiliary heating zone was 280 °C, the ion source was 230 °C, and the quadrupole temperature was 150 °C. The data acquisition mode was set to full scan, with a mass scanning range of 50 to 550 m/z and a solvent delay of 3 min. The GC–MS spectra were analyzed *via* a computer-based automatic search, the automated and manual analyses of a mass spectral library (NIST), and a literature search to identify each component. Finally, the relative contents of each constituent were calculated using the peak area normalization method.

Antibacterial Assay of the Extracted Essential Oil

Escherichia coli ATCC25922, *Staphylococcus aureus subsp. aureus* ATCC29213, *Salmonella enterica subsp. enterica* ATCC14028, *Pseudomonas aeruginosa* ATCC27853, *Candida albicans*, *Epidermophyton floccosum*, *Trichophyton rubrum*, and *Pseudomonas aeruginosa* were obtained from the China General Microbiological Culture Collection Center (CGMCC). Streptomycin was obtained from HyClone Co. (Los Angeles, USA), and penicillin G sodium was obtained from Biosharp Co. (Hefei, China). Terbinafine, amphotericin B, and DMSO were obtained from Sigma (San Francisco, CA, USA). Chloramphenicol was purchased from Scientific Research Special (San Francisco, CA, USA).

First, the samples were diluted to specific concentrations and added to the individual wells of a 96-well microplate. Next, the bacterial solutions were added and diluted to obtain a final concentration of 5×10^5 CFU/mL, and then cultured for 24 h at 37 °C. A microplate photometer was used to measure the optical density (OD) below 625 nm. Each treatment was the same as the three controls, and the mean value was taken. Control cultures, blank media, bacterial cultures, and penicillin-streptomycin and penicillin G sodium-positive controls were prepared. Inhibition rates were calculated based on Eq. 1 (NCCLS 2002a,b).

$$\text{Inhibition rate} = (1 - \text{OD value of sample} / \text{OD value of control well}) \times 100\% \quad (1)$$

Antifungal Assay of the Extracted Essential Oil

For the antifungal experiment, the samples to be tested were diluted by adding fungal liquid to each well of the 96-well plate. The final concentration of *Candida albicans* was 1×10^5 CFU/mL. *Epidermophyton floccosum* and *Trichophyton rubrum* were incubated at 37 °C for 24 h. The final concentration of *Plasmodium* sp. was 5×10^5 CFU/mL, incubated at 25 °C for 5 days, and the OD value at 625 nm was measured with a microplate reader. The experiment included a blank control medium, *Candida albicans*, *Epidermophyton floccosum*, *Trichophyton rubrum*, and the *Pseudomonas aeruginosa* control, along with the amphotericin B and terbinafine positive drug controls. The inhibition rates were calculated as shown in Eq. 1 (Liu 2003).

Antitumor Assay of the Extracted Essential Oil

The leukemia cell line HL-60, the lung cancer cell line A549, the hepatoma cell line SMMC-7721, the breast cancer cell line MCF-7, and the colon carcinoma cell line

SW480 were used. All cell lines were stored in the Kunming Institute of Botany. Cisplatin (DDP) and Taxol were also provided by the institute. All reagents were of analytical grade.

Dulbecco's modified Eagle medium (DMEM) culture medium containing 10% fetal bovine serum was used to formulate a single-cell suspension culture. Approximately 3000 to 15000 cells per well were seeded into the 96-well plates, with a volume of 100 μ L in each well. Adherent cells were cultured 12 to 24 h in advance. Next, 20 μ L of the supernatant from the samples was removed and dissolved in dimethyl sulfoxide. The essential oil samples were rescreened using five concentration gradients: 100, 20, 4, 0.8, and 0.16 μ g/mL. Each well had a final volume of 200 μ L, each treatment was performed in triplicate, and the mean value was taken. The samples were incubated at 37 °C for 48 h. After the incubation period, the media from the adherent cell cultures were discarded, and a solution of 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium (MTS) (20 μ L) and culture medium (100 μ L) were added to each well. For the cultured suspension cells, 100 μ L of the supernatant was discarded from each well; then, MTS solution (20 μ L) was added to each well. The cultures were further incubated for another 2 to 4 h. After the reactions had reached completion, the microplate photometer was employed to measure the OD value below 492 nm. DDP and Taxol were used as positive controls for each measurement. The cell growth curves were plotted with the concentration and cell viability on the x- and y-axes, respectively. The two-point Reed-Muench method was employed to calculate the half-maximal inhibitory concentration (IC₅₀) values of the compounds (Li *et al.* 2017) were calculated as shown in Eq. 1.

Hypoglycemic Assay of the Extracted Essential Oil

3T3-L1 adipocytes (adipocyte lines were preserved in Kunming Institute of Botany), insulin, berberine, and MTS were provided by Kunming Institute of Botany (Kunming, China). After 3T3-L1 cells were induced to differentiate into adipocytes, the cells were digested and seeded in 96-well plates and cultured overnight. After the cells were washed once with a low-glucose medium (glucose concentration 1800 mg/L), 200 μ L of low-sugar medium containing different drugs was added. The control group was incubated with DMSO and 100 μ M insulin and berberine, respectively, and the test sample group was incubated at a final concentration of 40 μ g/mL. Three replicate wells were used per sample.

After incubation for 24 h, 10 μ L of cell culture medium was aspirated, and the glucose concentration in the culture fluid was measured by the glucose oxidase-peroxidase method. At the same time, 20 μ L of MTS was added to the cell rest medium and incubated at 37 °C for 2 h. The absorbance at 492 nm was used to judge whether the compound was toxic to adipocytes. The glucose concentration and glucose consumption rate were calculated by Eqs. 2 and 3, respectively (Zhang *et al.* 2011).

$$\text{Glucose concentration (mmol/L)} = \frac{\text{sample absorbance}}{\text{standard absorbance}} \times \text{calibrator concentration} \quad (2)$$

$$\text{Glucose consumption (\%)} = \frac{\text{initial concentration} - \text{experimental concentration}}{\text{initial concentration}} \times 100 \quad (3)$$

RESULTS AND DISCUSSION

Analysis of the Main Chemical Constituents of *P. bournei* Wood

Under the experimental conditions, the yield of essential oil from the wood of *P. Bournei* was 0.9 mL/100 g. By GC-MS analysis, a total of 46 components were isolated from the wood samples. A total of 34 compounds were identified, and the sum of the relative contents accounted for 88.38% of the total sample content. The GC-MS spectra were analyzed *via* a computer-based automatic search, the automated and manual analyses of a mass spectral library (NIST), and a literature search to identify each component. Finally, the relative contents of each constituent were calculated using the peak area normalization method. Table 1 shows the results.

In this study, the chemical constituents of the essential oil from the *P. bournei* were identified, and their relative contents were measured. Under the experimental conditions, as shown in Table 1, the main constituents of the essential oil extracted from *P. bournei* were enol compounds. The sesquiterpene content of the wood sample was 84.46%, the monoterpene content was 0.64%, and the other component compounds were 3.28%. The main constituents of wood oil included α -copaene (5.44%), α -muurolene (7.32%), δ -cadinene(11.44%), and 1s-calamenene (5.18%).

The main volatile components of *Phoebe* wood from different production areas were basically the same, while the chemical compositions of different species of *Phoebe* differed greatly. The main chemical constituents of *P. bournei* wood essential oil were α -copaene, α -muurolene, and δ -cadinene. The main chemical components of *Phoebe hui* Cheng ex Yang wood were 3,7-guaiadiene, calarene, and torreyol (Ding *et al.* 2017b). The main components of *Phoebe zhennan* wood oil were agarospirol, guaiol, and γ -eudesmol (Ding *et al.* 2017a). It could be seen that the major components of the three were indeed very different from each other. Hence, it would be feasible to identify the *Phoebe* wood by chemical composition, and this is more accurate and objective than the traditional identification.

Study on the Bioactivity of the Essential Oil

Analysis of the antimicrobial effects of the essential oil extracted from the wood of P. bournei

The positive drugs Penicillin G Sodium and Streptomycin have an IC₅₀ of less than 2 μ g/mL for the four bacteria. Figure 1 shows that *P. bournei* wood oil had an inhibitory effect on the four tested bacteria; at all concentrations, *P. bournei* wood oil demonstrated the best activity against *P. aeruginosa*. Moreover, in the remaining three bacteria, when the concentration of *P. bournei* wood oil was less than 32 μ g/mL, the inhibitory effect of *P. bournei* wood oil against *S. aureus* and *S. enteritidis* was lower than that of *E. coli*. When the concentration was more than 32 μ g/mL, the inhibitory effect of *P. bournei* wood oil against *S. aureus* and *S. enteritidis* was better than that of *E. coli*.

The concentration of essential oil ranged from 8 μ g/mL to 128 μ g/mL, and the inhibitory rates of wood essential oil from *P. bournei* on the four tested bacteria showed an increasing trend, and had the best inhibitory effect against *P. aeruginosa*. The IC₅₀ of the essential oil to four bacteria was greater than 128 μ g/mL, indicating that the inhibition effect of *P. bournei* wood essential oil against *E. coli* ATCC25922, *Staphylococcus aureus* golden subspecies ATCC29213, *Salmonella enterica* subspecies ATCC14028, and *Pseudomonas aeruginosa* ATCC27853 is relatively weak, which is the same as the results of *Phoebe hui* and *Phoebe zhennan* wood essential oil (Ding *et al.* 2017a,b).

Table 1. The Chemical Composition of *P. bournei* Wood Oil

No.	Compound	KI	Area
1	a-copaene	1515	1.08
2	camphor	1552	0.60
3	isopinocampyon	1582	1.25
4	nopinone	1618	1.22
5	caryophyllene	1624	0.48
6	myrtenal	1662	0.55
7	pinocarveol	1685	2.35
8	p-menth-2-ene-1,8-diol	1700	0.33
9	verbenol	1707	2.18
10	a-terpineol	1721	1.97
11	endo-borneol	1728	0.80
12	verbenone	1746	6.38
13	p-mentha-1,5-dien-8-ol	1775	1.67
14		1786	0.46
15	d-cadinene	1816	2.93
16	myrtenol	1855	1.76
17		1849	0.56
18	calamenene	1855	3.95
19	carveol	1862	0.66
20	p-cymen-8-ol	1871	1.54
21		1893	0.75
22	a-agarofuran	1912	1.15
23		1971	0.83
24	caryophyllene oxide	2017	0.87
25		2074	1.06
26		2087	1.53
27	a-cedrene	2094	2.69
28	elemol	2103	2.30
29	10-eip-γ-eudesmol	2130	3.38
30		2151	1.09
31	a-eudesmol	2170	3.32
32		2182	0.82
33	agaruspirol	2207	17.55
34	bicyclo[4,4,0]dec-1-en,2-isopropyl-5-methyl-9-methylene	2213	1.25
35	calarene	2237	10.07
36	guaiazolene	2245	2.10
37		2252	4.99
38	b-eudesmol	2252	4.99
39	oxo-a-ylangene	2287	2.41
40		2318	0.92
41		2328	1.02
42	cycloisolongifolene	2359	2.20
43	caryophylla-3,8(13)-dien-5.beta.-ol	2363	0.78
44		2372	0.97
45	4-isopropyl-6-methyl-1,2,3,4-tetrahydronaphthalen-1-one	2376	0.71
46	5-HydroxyCalamenene	2613	0.93

Note: KI values (retention indices) are calculated from the retention times relative to those of the C7–C30

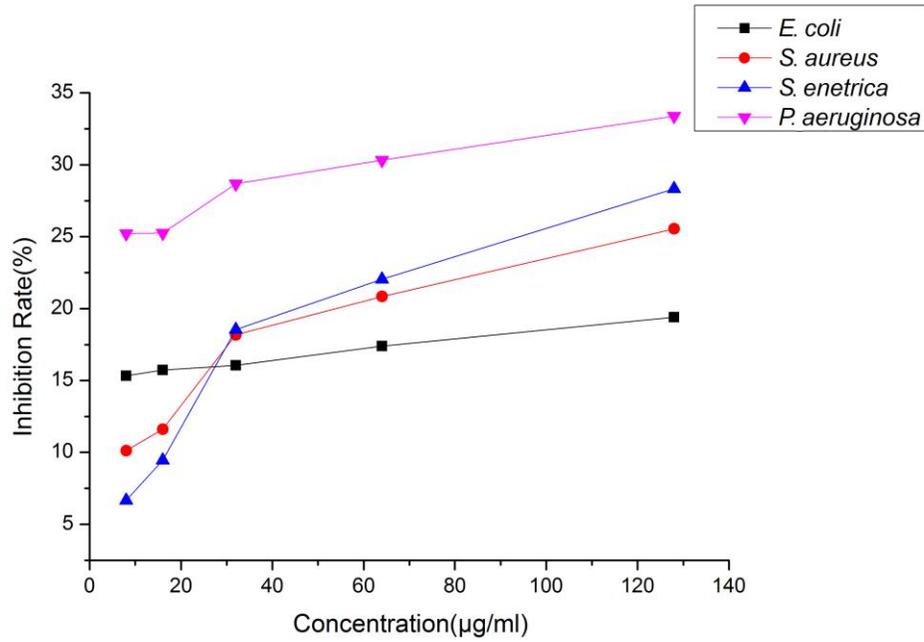


Fig. 1. Inhibition of four bacteria by different concentrations of the wood essential oil from *P. bournei*

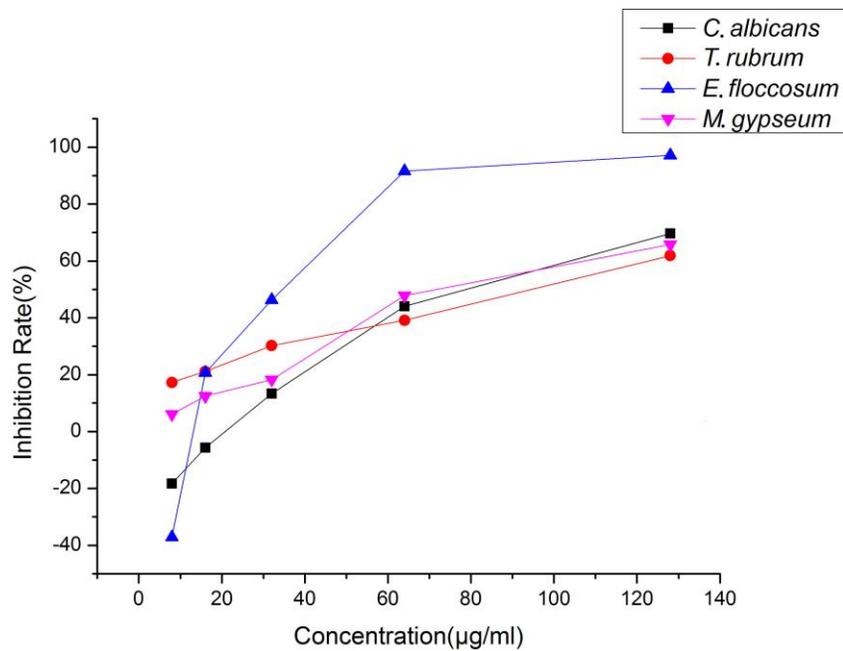


Fig. 2. Inhibition of four fungi by different concentrations of the wood essential oil from *P. bournei*

Analysis of antifungal activities of the essential oil extracted from wood of P. bournei

The positive drugs Amphotericin B and Terbinafine have an IC₅₀ of less than 0.0125 µg/mL for the four fungi. Figure 2 shows that *E. floccosum* showed the most significant response to the change in wood essential oil concentration of *P. bournei*, and the inhibition rate varied significantly with the change in essential oil; different concentrations of *P. bournei* wood essential oil always inhibited the *T. rubrum* and *M. gypseum*. However, *P. bournei* wood essential oil demonstrated a promoting effect on *E. floccosum* and *C. albicans* at low concentrations: when the wood essential oil concentration was higher than 16 µg/mL, the wood essential oil showed a certain degree of inhibition on the four test fungi, and in particular, the inhibitory effect on the *E. floccosum* was significantly better than that of the other three fungi. However, when the concentration was lower than 16 µg/mL, the inhibitory effect on the *E. floccosum* and *C. albicans* was rapidly transformed into a significant promoting effect.

With the concentration of essential oil ranging from 8 µg/mL to 128 µg/mL, the inhibitory rates of *P. bournei* wood essential oil on the four tested fungi showed an increasing trend. Wood essential oil exhibited significant inhibitory activity against *Candida albicans* ATCC10231, *Trichophyton rubrum* ATCC4438, *Epidermophyton floccosum* CBS566.94, and *Microsporium gypseum* CBS118893, with the IC₅₀ values of 75 µg/mL, 82.6 µg/mL, 33.6 µg/mL, and 69.60 µg/mL, respectively. Attention could be paid to the development of related antifungal active substances of *P. bournei* wood essential oil.

Analysis of the antitumor effects of the essential oil extracted from wood of P. bournei

Table 2 shows the results obtained from the *in vitro* comparison of the IC₅₀ values of the samples. As shown, the essential oil from wood of *P. bournei* had good activity against leukemia cell lines, with an IC₅₀ value of 42.7 µg/mL. The essential oil from the wood of *P. bournei* exhibited poor activity against lung cancer, liver cancer, breast cancer, and colon cancer cell lines. In particular, the toxic activity of the lung cancer cells was the worst, with an IC₅₀ values of more than 50 µg/mL.

Table 2. Effect of the Essential Oil on the IC₅₀ of Five Cancer Cells

	Leukemia Cell Line HL-60	Lung Cancer Cell Line A549	Hepatoma Cell Line SMMC-7721	Breast Cancer Cell Line MCF-7	Colon Carcinoma Cell Line SW480
IC ₅₀ ±SD (µg/mL)					
Wood	42.7±1.84	>100	88.1±2.79	54.9±0.58	75.6±2.10
DDP	2.3±0.81	4.7±0.26	6.9±0.59	8.2±0.57	8.1±1.44
Taxol	<0.0068	<0.0068	<0.0068	<0.0068	<0.0068

P. bournei wood essential oil exhibited good activity against leukemia cell lines, and exhibited the worst activity against lung cancer cell lines. This is the same as the results for *Phoebe hui* and *Phoebe zhennan* wood essential oil (Ding *et al.* 2017a,b). Further attention could be paid to the research and development of antitumor-related products.

Analysis of the hypoglycemic activity of the essential oil extracted from wood of P. bournei

Table 3 shows the effects of essential oil extracted from the wood of *P. bournei* on cell glucose uptake. In the adipocyte glucose uptake model, wood essential oil acted at a concentration of 40 µg/mL for 24 h, and the adipocyte glucose consumption rate was increased. That is, wood essential oil played a certain role in the promotion of glucose uptake by adipocytes. It also showed a certain potential for hypoglycemic activity, although

there was still a certain gap between the oil and the mature hypoglycemic drugs. Because essential oil is a mixture, the concentration of effective components within the volume was relatively low, and thus the wood essential oil of *P. bournei* was basically non-toxic to adipocytes at this concentration. Thus, there is still a certain potential for exploring its effects on hypoglycemic activity.

Table 3. Effects of *P. bournei* Wood Essential Oil on Glucose Consumption Rate

Sample	Final Concentration	Glucose Consumption Rate (%)
DMSO (Control)		16.23±1.57
Insulin	0.1 µM	38.22±1.18
Berberine (235.324)	10 µM	31.23±1.77
Wood	40 µg/mL	20.49±1.35

CONCLUSIONS

1. The main components of *Phoebe bournei* wood essential oil are α -copaene (5.44%), α -muurolene (7.32%), δ -cadinene (11.44%), and 1s-calamenene (5.18%). The composition of wood essential oil is different from that of *Phoebe zhennan* and *Phoebe hui*, so it is feasible to use the chemical composition of *P. bournei* as a marker to identify *P. bournei*.
2. *P. bournei* wood essential oils exhibited some inhibitory effect on the eight tested bacteria and fungi relative to the control, and with the increase of essential oil concentration, the inhibition rate showed an increasing trend. The essential oil had weaker activity against *Escherichia coli* ATCC25922, *Staphylococcus aureus subsp. aureus* ATCC29213, *Salmonella enterica subsp. enterica* ATCC14028, and *Pseudomonas aeruginosa* ATCC27853. Among the four tested fungi, the wood essential oil of *P. bournei* had significant inhibitory activity against *Candida albicans* ATCC10231, *Trichophyton rubrum* ATCC4438, *Epidermophyton floccosum* CBS566.94, and *Microsporum gypseum* CBS118893.
3. *P. bournei* wood essential oil exhibited good activity against leukemia cell lines, and exhibited the worst activity against lung cancer cell line. *P. bournei* wood essential oil acted at a concentration of 40 µg/mL for 24 h, which had a certain role in promoting glucose uptake by adipocytes.

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