

Fast-responsive Temperature-sensitive Hydrogels

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Temperature fast-responsive and magnetic poly(*N*-isopropylacrylamide-co-acrylamide) (CMX-MNP-PNIPAm/Fe₃O₄) hydrogels were developed using carboxymethyl xylan (CMX) as a pore-forming agent and a NaCl solution as the reaction medium, followed by fabricating Fe₃O₄ nanoparticles *in situ* within the hydrogel matrix. It was found that NaCl played a role in the phase separation and was used as the electrolyte to shield CMX molecular chains. The obtained hydrogels exhibited a fast, temperature-responsive behavior, and the water retention was less than 15% for 1 min under 60 °C. The prepared hydrogels showed enhanced mechanical properties and magnetic properties due to the presence of Fe₃O₄ particles. The lower critical solution temperature of the hydrogels was in the range of 35 to 39 °C, which was acquired through adjusting the amount of hydrophilic monomer (AM). The magnetic and thermosensitive hydrogel had the attractive photothermal conversion ability and could be heated to 40 °C within 2 min, and to 69 °C within 7 min under near infrared irradiation.

Keywords: Temperature fast-responsive hydrogels; Carboxymethyl xylan; Pore-forming agent; Magnetic property; Photothermal conversion

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INTRODUCTION

Recently, intelligent hydrogels have attracted considerable attention because of their multiple environmental response behaviors, including sensitivity to temperature, pH, salt, and biomolecules (Peng *et al.* 2011b; Dai *et al.* 2016). These attributes make them have great potential in the biomedical field as targeted drug release agents, super-absorbents, and biosensors. Temperature-sensitive hydrogels are intelligent materials that can respond to temperature according to the reversible change in volume (swelling/deswelling) *via* absorbing or releasing water or other liquids (Higgins *et al.* 2016). Poly(*N*-isopropylacrylamide) (PNIPAm) hydrogel, as a typical temperature-sensitive hydrogel, has the characteristics of thermal shrinkage (showing a lower critical solution temperature, LCST) due to hydrophilic amino groups and hydrophobic isopropyl groups within the hydrogel (Ashraf *et al.* 2016). Below the LCST, PNIPAm hydrogels are swollen and hydrophilic, while as the temperature exceeds the LCST, they exhibit a collapse and dehydration because the hydrophile/hydrophobicity balance in the hydrogel network structure is destroyed (Shekhar *et al.* 2016).

However, traditional PNIPAm hydrogels have a poor response rate to temperature change because the dense surface of hydrogels hinders the discharge of internal water molecules in the deswelling process, which limits their application in many fields (Shibayama and Nagai 1999). The response rate of temperature is an important factor in the evaluation of temperature-sensitive hydrogels. A great deal of effort has been devoted to improving the temperature response rate of PNIPAm hydrogels, and some effective

methods have been developed, including phase-separation and pore-forming agent methods (Zhang and Zhuo 2000; Cheng *et al.* 2003). For example, the PNIPAm hydrogels in some inorganic salts or saccharides solution could result in a phase-separation and then enhance the rate of shrinkage. Aqueous sucrose solutions were used to prepare temperature-sensitive PNIPAm hydrogels with faster response rates and higher swelling ratios. The response rate of hydrogels prepared in the 0.15 M sucrose solution was obviously faster than that of conventional hydrogels prepared in pure water (Watsons). The former lost at least 80% water in 10 min, while the latter lost only approximately 15% water within 10 min and took 24 h to reach the equilibrium. This improved property was ascribed to the porous structure of hydrogels, which was caused by the phase separation in the sucrose solution (Zhang *et al.* 2003a). The fast response PNIPAm hydrogels were prepared in the aqueous glucose solution. The hydrogels prepared in the 1.0 M glucose solution released 95% water in 5 min, whereas the conventional hydrogels prepared without glucose solution lost approximately 40% water in 20 min. These properties were due to the glucose solution impact, which caused the phase separation and formed a porous structure during hydrogels preparation (Cheng *et al.* 2003). The aqueous NaCl solution was applied as the reaction medium to synthesize temperature-sensitive PNIPAm hydrogels with fast, response behavior. Hydrogels prepared in the 1.0 M NaCl solution released at least 95% water in 5 min, while the gels prepared in pure water released approximately 50% water in 120 min. The salt influence led to the phase separation and resulted in heterogeneous macroporous structures during the polymerizations (Cheng *et al.* 2003).

A pore-forming agent is another effective method to improve the temperature-sensitive rate of PNIPAm hydrogels. Fast-responsive PNIPAm hydrogels were prepared with polyethylene glycol (PEG) as the pore-forming agent. The response rate of hydrogels prepared in the presence of PEG was significantly faster than that of hydrogels prepared in the absence of PEG. The former released approximately 95% water in 3 min and the later lost only 40% water in 68 min because PEG acted as the pore-forming agent, which resulted in producing a large amount of macroporous structures during the polymerizations and macroporous hydrogels could absorb more water and exhibit a fast response rate (Zhang and Zhuo 2000). A fast-responsive PNIPAm hydrogel with macroporous structure was fabricated using carboxymethyl cellulose (CMC) as the pore-forming agent (Wu *et al.* 1992). Hydrogels prepared in the presence of CMC reached to the swelling equilibrium (no longer absorb water) in only a few minutes, which was at least 10-fold faster than that of hydrogels prepared in the absence of CMC. The faster response was attributed to the existence of CMC, which resulted in many open pores by occupying extra volume during the polymerization of hydrogels. After the polymerization, CMC was washed out from the hydrogels, and macroporous structure was formed within the hydrogels.

Xylan is an abundant, nontoxic, biodegradable, widely available, and natural polysaccharide (Ebringerová and Heinze 2000) that can be modified to form multi-functional polymers by different methods, such as oxidation, carboxymethylation, esterification, and so on. Recently, a series of hemicelluloses-based intelligent hydrogels were prepared by introducing functional xylan. These synthesized hydrogels exhibited controllable swelling abilities, temperature sensitivity, pH sensitivity, and magnetic properties (Peng *et al.* 2012; Gao *et al.* 2014, 2015), which make them promising materials for a variety of applications. However, there are few studies on xylose or xylan derivative as the pore-forming agent to prepare intelligent hydrogels.

Carboxymethylation enables xylan to have anionic properties by introducing more $-\text{COO}^-$ groups on its main chain. In an aqueous salt solution, the repulsion between

carboxymethyl xylan (CMX) molecular chains became weaker and promoted the curl of CMX molecular chains, which was beneficial to producing a large amount of porous structures in the preparation of hydrogels. In this work, carboxymethyl xylan was prepared, and for the first time used as the pore-forming agent to prepare temperature fast-responsive magnetic hydrogels (CMX-MNP-PNIPAm/Fe₃O₄) in the NaCl solutions. The impact of CMX as the pore-forming agent was studied on the hydrogel's properties as well as the NaCl solution. The characteristics of the prepared hydrogels were determined *via* scanning electron microscopy (SEM), carbon-13 nuclear magnetic resonance (¹³C-NMR) spectrometry, thermogravimetric analysis (TGA), vibrating sample magnetometry (VSM), differential scanning calorimetry (DSC), X-ray diffraction (XRD), and Raman spectra. The photothermal conversion properties of the obtained hydrogels were also investigated.

EXPERIMENTAL

Materials

Beech wood xylan (M_w of 100,000 to 130,000 g/mol) was obtained from Sigma Aldrich (Karlsruhe, Germany). Acrylamide (AM, 98%), NIPAm (98%), chloroacetic acid (99%), ammonium persulfate (99%), tetramethylethylenediamine (TEMED, 99%), *N,N'*-methylenebisacrylamide (MBA, 98%), FeSO₄•7H₂O, FeCl₃•6H₂O, NaOH, and NaCl were supplied by Aladdin Reagent Co., Ltd. (Shanghai, China). Glacial acetic acid (99%), ammonium hydroxide (NH₃•H₂O, 25% to 28%), ethanol, and H₂O₂ were received from Guangzhou Chemical Reagent Company (Guangzhou, China). All the reagents were used without further purification.

Methods

Preparation of carboxymethyl xylan

The type of preparation procedure of CMX was conducted according to the authors' previous work (Ren *et al.* 2008). A total of 6.6 g of xylan was dissolved in 25 mL of distilled water with vigorous magnetic stirring at 80 °C for 70 min. After the solutions were cooled to 30 °C, 2 mL of the NaOH solution (12.5 mol/L) was added and stirred for 20 min. The 11-mL sodium chloroacetate solution (0.1 mol/L) and the 2-mL NaOH solution (12.5 mol/L) were added into the above solution, and the reaction mixture solution was irradiated at 65 °C for 20 min with a 300 W microwave (VIP 200; Whirlpool, Michigan, America). After reaction, the mixture was first cooled down, then neutralized with diluted acetic acid, and dialyzed in dialysis bag (M_w of 3000) for a week. The purified carboxymethyl xylan was freeze-dried and preserved. The degree of substitution (DS) of the CMX was 0.85, which was measured by the acidometric titration method.

Preparation of magnetic temperature fast-responsive hydrogels

A total of 2 g of NIPAm were dissolved in different concentration NaCl solutions, and different amounts of CMX, AM, and MBA were added into the mixtures in a water bath at 10 °C for magnetic stirring. After bubbling N₂ for 20 min, a certain amount of ammonium persulfate was first added in the mixtures as the initiator, and after 10 min TEMED was added to end the reaction. Subsequently, the mixture was poured into a glass mould at 25 °C for 6 h. After reaction, the mould was removed and the obtained hydrogels were washed with distilled water. Subsequently, the prepared hydrogels were cut into cubes and immersed in the mixture solution (40 mL) containing 2 mmol of FeCl₃•6H₂O and 1

mmol of $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$ for 6 h under a nitrogen atmosphere. Then, hydrogels loaded with iron ions were transformed into 30 mL of $\text{NH}_3 \cdot \text{H}_2\text{O}$ for 30 min under a nitrogen atmosphere. Finally, black hydrogels were obtained and washed with distilled water to remove excess ions and impurity. The detailed synthesis conditions are shown in Table 1.

Table 1. Preparation Conditions of the CMX-MNP-PNIPAm Hydrogels

Entries	NIPAm (g)	CMX (g)	MBA (g)	AM (g)	NaCl (mol/L)	H ₂ O (mL)
MBA _{0.05}	2	0.12	0.05	0.1	0.4	24
MBA _{0.10}	2	0.12	0.1	0.1	0.4	24
MBA _{0.15}	2	0.12	0.15	0.1	0.4	24
MBA _{0.20}	2	0.12	0.2	0.1	0.4	24
AM _{0.05}	2	0.12	0.1	0.05	0.4	24
AM _{0.10}	2	0.12	0.1	0.1	0.4	24
AM _{0.20}	2	0.12	0.1	0.2	0.4	24
AM _{0.40}	2	0.12	0.1	0.4	0.4	24
NaCl ₀	2	0.12	0.1	0.05	--	24
NaCl _{0.2}	2	0.12	0.1	0.05	0.2	24
NaCl _{0.4}	2	0.12	0.1	0.05	0.4	24
NaCl _{0.6}	2	0.12	0.1	0.05	0.6	24
CMX ₀ /NaCl ₀	2	--	0.1	0.05	--	24
CMX ₀ /NaCl _{0.2}	2	--	0.1	0.05	0.2	24
CMX ₀ /NaCl _{0.4}	2	--	0.1	0.05	0.4	24
CMX ₀ /NaCl _{0.6}	2	--	0.1	0.05	0.6	24
CMX/MBA/AM/NaCl	2	0.12	0.2	0.05	0.6	24

Characterization of the hydrogels

Surface morphology of hydrogels and images of Fe_3O_4 nanoparticles in hydrogels were recorded using SEM (Quanta 200; FEI Company, Hillsboro, USA) and field emission (FE)-SEM (JSM-7000F; JEOL Co., Ltd., Beijing, China) with an accelerating voltage of 10 kV and 20 kV, respectively. The ^{13}C -NMR of xylan, CMX, and hydrogels were analyzed using a Bruker AV-III 400 M (Bruker, Karlsruhe, Germany). The solvent and the internal standard were D_2O and trimethylsilane, respectively. The XRD patterns of hydrogels were tested on a Bruker diffractometer with $\text{Cu K}\alpha$ radiation (Bruker, Karlsruhe, Germany) in the range of 20° to 80° . The magnetic hydrogels were measured using VSM (LakeShore-7307; Lakeshore Company, Columbus, OH, USA) at room temperature. The Raman spectra (HORIBA HR 800; HORIBA H.J.Y Company, Paris, France) of hydrogels were recorded at 632.8 nm radiation from an argon ion laser with an output power of 20 mW and a typical acquisition time of 10 s. The TGA experiments of hydrogels were performed using a TGA Q500 instrument (New Castle, DE, USA) under a nitrogen flow of 20 mL/min from 50 to 600 °C at a heating rate of 10 °C/min. The DSC experiments for LCST determination were from -10 °C to 60 °C with a heating rate of 2 °C/min on a DSC-Q2000 apparatus (TA Instruments, Newcastle, DE, USA) under a nitrogen flow of 25 mL/min. The compressive strength of hydrogels was determined by a universal material testing machine (Instron 3300; Instron Corporation, Norwood, MA, USA).

Swelling degree and photothermal conversion properties of hydrogels

The swelling degree of hydrogels was measured based on a gravimetric method. The dehydrated hydrogel samples were immersed in distilled water in the temperature range of 20 °C to 60 °C. When the samples reached an equilibrium swelling state, the swollen hydrogels were removed from the water, the water was wiped off with filter paper, and the material was immediately weighed. The determination of all samples was conducted in three parallel measurements. The swelling ratio (SR), equilibrium swelling ratio (Seq), and water retention (WR) were calculated as follows (Sun *et al.* 2013),

$$SR = (W_t - W_o) / W_o \quad (1)$$

$$S_{eq} = (W_t - W_o) / W_o \quad (2)$$

$$WR = (W_t - W_o) / (W_{eq} - W_o) \quad (3)$$

where W_t is the weight (g) of swollen hydrogels, W_o is the weight (g) of dried hydrogels, and W_{eq} is the equilibrium weight (g) of swollen hydrogels.

The CMX-MNP-PNIPAm/Fe₃O₄ hydrogel (0.9 × 0.4 × 0.3 cm) was placed in a cuvette that contained 0.6 mL water. The photothermal conversion properties of the hydrogels were measured using near infrared (NIR) irradiation (GKGY-NIR-200; Hengaode Instrument Co., Ltd., Beijing, China), which was performed at 808 nm with a radiant power of 2 W. The reversibility of photothermal changes was investigated by the swelling-deswelling of the CMX-MNP-PNIPAm/Fe₃O₄ hydrogels.

RESULTS AND DISCUSSION

¹³C -NMR Spectra of Native Xylan, CMX, and Hydrogel

Figure 1 shows the ¹³C-NMR spectra of CMX (a), CMX-MNP-PNIPAm hydrogels (b), and native xylan (c). As shown in Fig. 1a, these peaks at 101.77, 76.48, 73.79, 72.81, and 63.09 ppm are assigned to C-1, C-4, C-3, C-2, and C-5 of the β-D-xylpyranosyl units of xylan, respectively (Peng *et al.* 2011a). Compared with the spectrum of xylan (Fig. 1c), two new signals at 68.6 ppm and 178.34 ppm were observed, which were attributed to the -CH₂ of the carboxymethyl and carboxyl groups, respectively (Lazik *et al.* 2002). These results demonstrated that the carboxymethylation occurred on xylan.

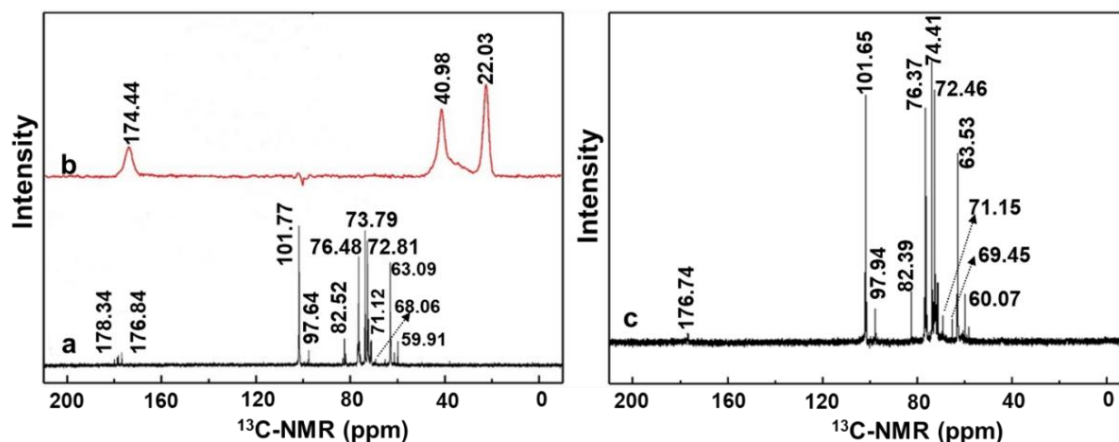


Fig. 1. The ¹³C-NMR spectra of CMX (a), CMX-MNP-PNIPAm hydrogels (b), and native xylan (c)

In Fig 1a, signals at 176.84, 97.64, 82.52, and 59.91 ppm are the characteristic signals of C-6, C-1, C-4, and the methoxyl group of the 4-*O*-methyl-D-glucosyluronic acid residue, respectively (Sun *et al.* 1998). In Fig. 1b, the signals at 22.03 and 40.98 ppm are attributed to the methyl carbon of *N*-isopropyl (NIPAm) and the vibration of the -CH group (Lo *et al.* 2005). The peak at 174.44 ppm (Fig. 1b) was ascribed to the carbonyl group (C=O), which was associated with acrylamide and NIPAm in hydrogels (Lazik *et al.* 2002). Compared with the spectrum of CMX in Fig. 1a, the characteristic signals of CMX did not appear in Fig. 1b. These changes confirmed that prepared hydrogels did not contain CMX. It was also shown that CMX was eluted as a pore-forming agent during the procedure of synthetic hydrogels.

Morphology Analysis of Hydrogels

For Fig. 2a through 2d, it is clearly apparent that hydrogels with pore sizes of 2 μm to 10 μm were successfully prepared by changing the crosslinking agent amount. With an increase in the amount of MBA, the pore sizes of hydrogels decreased due to improving the crosslinking density. The porous structure of hydrogels became more uniform. This illustrated that the pore sizes of hydrogels could be controlled by adjusting the MBA amount. Figure 2e and 2f show the deswelling SEM images of MBA_{0.1} hydrogels. After deswelling, the network structure of hydrogels shrunk, and then the surface of the hydrogels wrinkled. After magnification (Fig. 2f), the hydrogels surface was a three-dimensional porous structure consisting of very small holes. Figures 2a and 2g show the surface appearance of the traditional PNIPAm hydrogels without CMX and CMX-MNP-PNIPAm hydrogels (with CMX), respectively. Compared with Fig. 2g, the image in Fig. 2a exhibits a larger pore size and a more stereoscopic network structure at the same preparation conditions. This results indicates that CMX acts as a pore-forming agent in the preparation of the hydrogel and promotes the formation of pore.

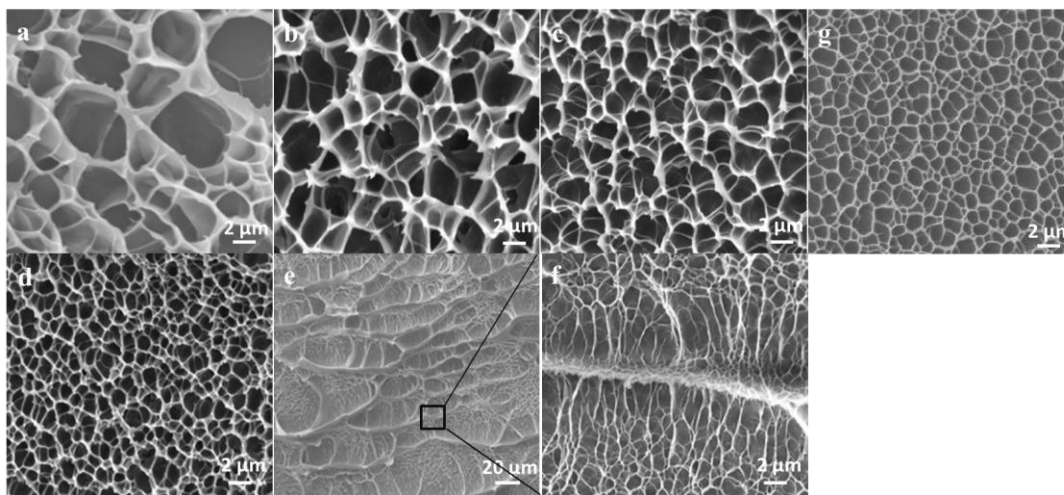


Fig. 2. The SEM images of CMX-MNP-PNIPAm hydrogels: (a) MBA_{0.05}; (b) MBA_{0.1}; (c) MBA_{0.15}; (d) MBA_{0.2}; (e) deswelling MBA_{0.1}; (f) the magnification of deswelling MBA_{0.1} and (g) traditional PNIPAm hydrogels without CMX

Photographs of CMX-MNP-PNIPAm hydrogels before the fabrication of Fe₃O₄ nanoparticles that were synthesized in different NaCl concentrations without washing with deionized water to remove unreacted chemicals are shown in Fig. 3. It was obvious that all

hydrogels were light yellow, which was attributed to the presence of carboxymethyl hemicellulose (Kong *et al.* 2016). The transparent hydrogels (NaCl_0) were obtained in an aqueous solution without sodium chloride. However, hydrogels synthesized in sodium chloride aqueous solution were semi-transparent ($\text{NaCl}_{0.2}$) or even opaque ($\text{NaCl}_{0.4}$, $\text{NaCl}_{0.6}$), indicating that increased NaCl concentrations caused increases in the opacity degree of the hydrogels. This was due to the existence of pore-forming agent and the NaCl solutions. In the synthesis process of hydrogels, the repulsion from the CMX chain was depressed in the NaCl solutions to facilitate the curliness of the CMX chain, and the pore-forming agent worked. Meanwhile, the PNIPAm chains curled and intertwined with each other, finally becoming a nucleus, which was due to the fact that the formed PNIPAm chains could not dissolve extendedly in the NaCl solutions (Zhang *et al.* 2003b). The agglomeration of the nucleus led to forming a macroporous structure in hydrogels. Therefore, the phase separation easily occurred when the NaCl solution concentration exceeded 0.2 mol/L.

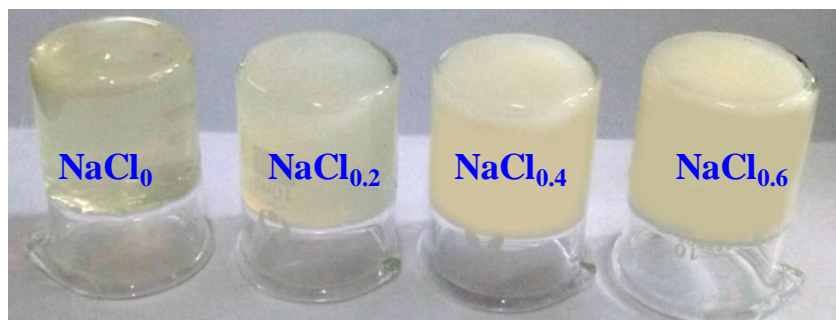


Fig. 3. Digital photograph of CMX-MNP-PNIPAm hydrogels before incorporating Fe_3O_4 nanoparticles

Magnetic Particle Analysis in Hydrogels

As shown in Fig. 4a, CMX-MNP-PNIPAm/ Fe_3O_4 hydrogels had apparent diffraction peaks at $2\theta = 30.25^\circ$, 35.70° , 43.16° , 53.94° , 57.42° , and 62.83° , which are ascribed to the (220), (311), (400), (422), (511), and (440) planes of Fe_3O_4 , respectively (Zhu *et al.* 2014). The Raman spectrum in Fig. 4b shows a single peak at 670 cm^{-1} , which was consistent with the characteristic peak of Fe_3O_4 (Shebanova and Lazor 2003), but is different from that of Fe_2O_3 , which has three prominent peaks around 350, 500, and 700 cm^{-1} (Wang *et al.* 2009). Therefore, Fe_3O_4 particles were *in-situ* prepared within the network of CMX-MNP-PNIPAm hydrogels. The hysteresis loop and the SEM image of CMX-MNP-PNIPAm/ Fe_3O_4 hydrogels are shown in Fig. 4c and 4d, respectively. Clearly, the saturation magnetization of CMX-MNP-PNIPAm/ Fe_3O_4 hydrogels was 10.78 to 12.51 emu/g. This value was increased with the increase of crosslinking concentration, which was due to the influence of the crosslinking concentration on the magnetism of Fe_3O_4 by affecting the pore structure of hydrogels (Gao *et al.* 2014). When the pore size of the hydrogels network was less than $2\text{ }\mu\text{m}$, the anisotropy of Fe_3O_4 nanoparticles began to grow (Gnanaprakash *et al.* 2007). Moreover, the hysteresis loop of the prepared hydrogels did not decrease obviously. The hydrogels exhibited apparent superparamagnetism. Therefore, it was demonstrated that the crosslinking concentration of hydrogels had an apparent effect on the Fe_3O_4 particles by affecting the pores structures of hydrogels. In Fig. 4d, the Fe_3O_4 particles were uniformly distributed in the hydrogels' network, and the size was approximately 10 nm.

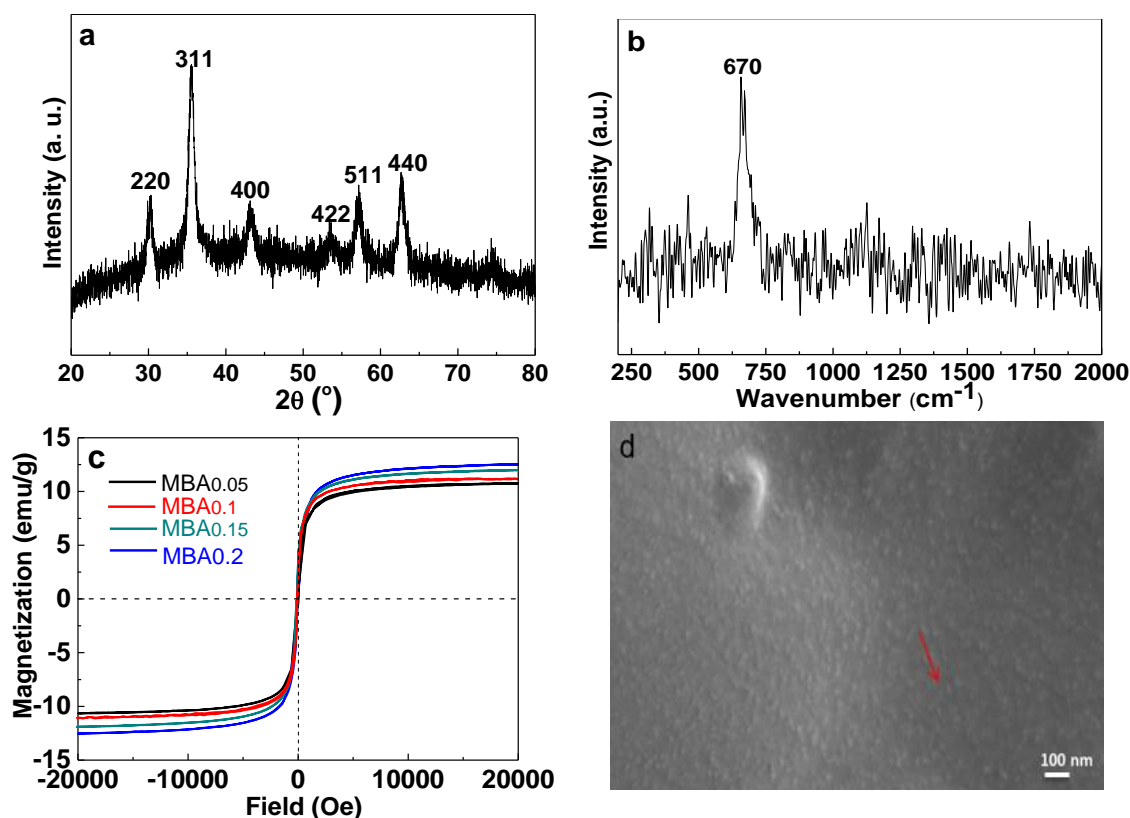


Fig. 4. The XRD spectra (a) and Raman spectra (b) of CMX-MNP-PNIPAm/Fe₃O₄ (MBA_{0.2}); hysteresis loop (c) of CMX-MNP-PNIPAm/Fe₃O₄ hydrogels with different MBA amounts, and SEM image (d) of CMX-MNP-PNIPAm/Fe₃O₄ hydrogels

Thermogravimetric and Mechanical Property Analysis of Hydrogels

Thermogravimetric analysis of the CMX-MNP-PNIPAm/Fe₃O₄ hydrogels is shown in Fig. 5. It was obvious that the weight loss of CMX and hydrogels occurred at approximately 200 °C to 300 °C and 300 °C to 400 °C, respectively. This illustrated that hydrogels did not contain CMX, which was consistent with the analysis of the above-mentioned Fig. 1. The weight loss at 200 °C to 300 °C was mainly attributed to the degradation of molecular chains of CMX, and the mass loss in the range of 250 °C to 300 °C was ascribed to the dehydroxylation and decarboxylation of xylan (Peng *et al.* 2015). The weight loss at 300 °C to 400 °C was ascribed to the cleavage of the carbon backbone in hydrogels (Ghorpade *et al.* 2018). When the temperature was above 400 °C, the weight of the sample related to carbonization gradually decreased and leveled off. Similar TG curves were observed for CMX-MNP-PNIPAm/Fe₃O₄ hydrogels with different MBA amounts, which demonstrated that MBA had no effect on the thermal stability of hydrogels. These changes showed that CMX-MNP-PNIPAm/Fe₃O₄ hydrogels had a good thermal stability compared to CMX.

In addition to thermal stability of hydrogels, strength is also an important criterion. Figure 6 displays the compressive stress-strain curve of CMX-MNP-PNIPAm/Fe₃O₄ hydrogels. Clearly, the compressive stress of hydrogels increased with the amount of MBA crosslinking agent. When the amount of MBA was the maximum, the compressive stress of the MBA_{0.2} sample could reach 180 kPa, and the corresponding compressive strain reached 66%. One possible explanation was that Fe₃O₄ nanoparticles were fabricated in the

hydrogel network by co-precipitation and dispersed more evenly in the hydrogels, resulting in improved compression strength (Dai *et al.* 2016). For Fig. 6b, the CMX-MNP-PNIPAm/Fe₃O₄-hydrogels had good compression properties.

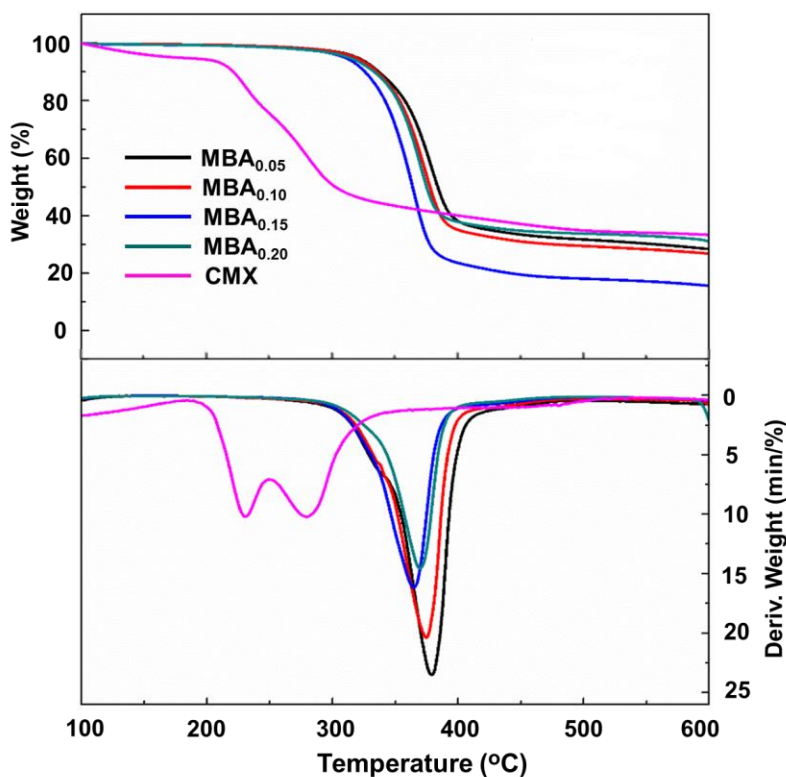


Fig. 5. TG curves of CMX-MNP-PNIPAm/Fe₃O₄ hydrogels and CMX

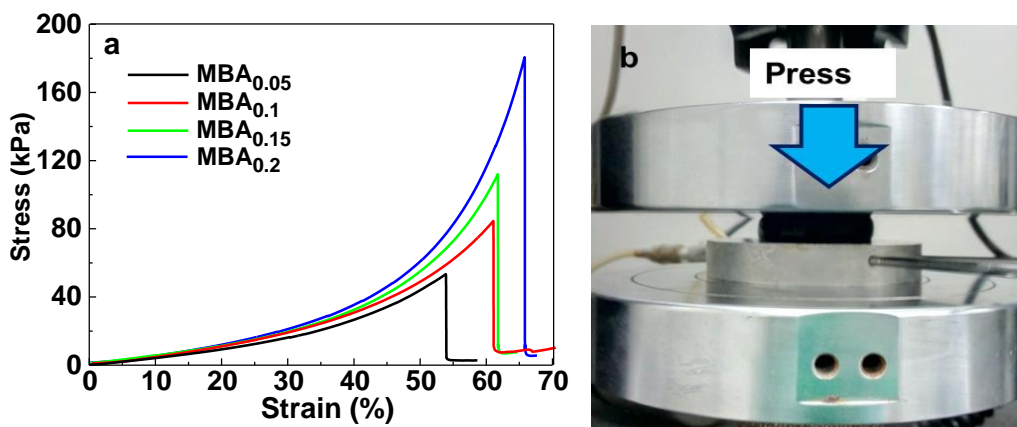


Fig. 6. The compressive strength of the CMX-MNP-PNIPAm/Fe₃O₄ hydrogels (a) and the compressive test of CMX-MNP-PNIPAm/Fe₃O₄ hydrogels (MBA_{0.2}) (b)

Deswelling Behavior of Hydrogels

The SR of CMX-MNP-PNIPAm/Fe₃O₄ hydrogels with different MBA amounts as a function of different temperatures is shown in Fig. 7a. The SR of all hydrogels decreased with an increase in the temperature in 60 min. Especially, when the temperature from 36

°C rose to 38 °C, the SR of hydrogels decreased sharply at different MBA amounts. This illustrated that the volume of hydrogels shrank rapidly when the temperature exceeded 38 °C. Figure 7b shows the DSC curves of the CMX-MNP-PNIPAm/Fe₃O₄ hydrogels with different AM amounts. The DSC curves of prepared hydrogels showed a rising trend with an increase of the temperature from 5 °C to 60 °C, which was attributed to the production of evaporation of enthalpy in the hydrogels when the temperature increased (Hou *et al.* 2008). All of the hydrogels showed an obvious peak near 36 °C to 38 °C, which was due to the production of the phase transition enthalpy caused by the phase transition of hydrogels. The corresponding temperature of the peak was the LCST of hydrogels (Ma *et al.* 2012), and the LCST gradually rose following the increase in the AM amount. The LCST of CMX-MNP-PNIPAm/Fe₃O₄ hydrogels were 35.83 °C, 36.70 °C, 37.84 °C, and 38.60 °C, corresponding to 0.05-M, 0.1-M, 0.2-M, and 0.4-M AM, respectively. Clearly, the LCST of CMX-MNP-PNIPAm/Fe₃O₄ hydrogels could be controlled by adjusting the AM amounts. This was because hydrophilic acrylamide was introduced into the hydrogels network, increasing the hydrophilic group ratio of the hydrogels network, further forming more hydrogen bonds, and requiring more energy to break the hydrogen bonds (Wang *et al.* 2013). At near 0 °C, the hydrogel had a remarkable endothermic peak corresponding to the phase transition of free water (Tamai *et al.* 1996), indicating that there was a glass transition between -10 °C to 0 °C after the swelling of hydrogels in distilled water.

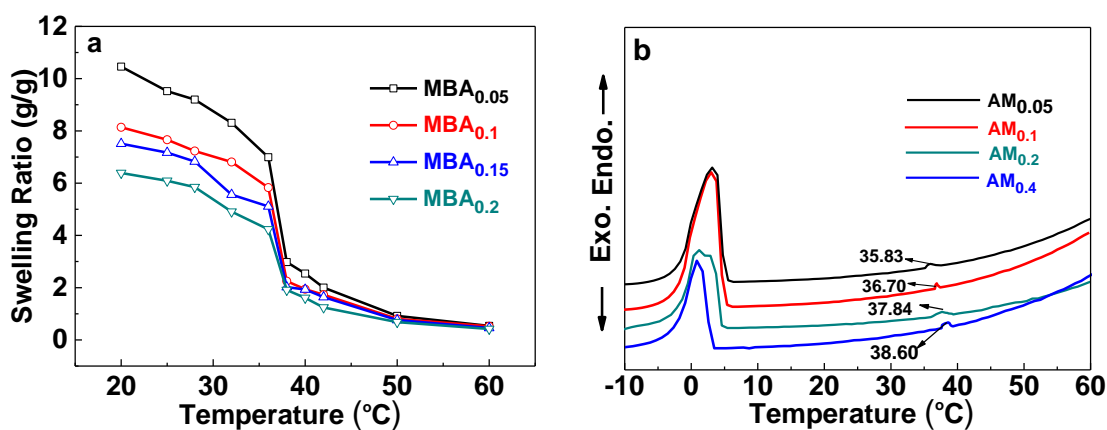


Fig. 7. The equilibrium swelling ratio (a) and the DSC curves (b) of CMX-MNP-PNIPAm/Fe₃O₄ hydrogels with different MBA amounts

The swelling behavior of CMX-MNP-PNIPAm/Fe₃O₄ hydrogels at 20 °C and deswelling behavior at 60 °C were studied. The effect of MBA amounts on the equilibrium swelling rate of hydrogels is shown in Fig. 8a. The equilibrium swelling rates of hydrogels first increased and then leveled off with the increase of time and decreased with the increase of MBA amounts at 20 °C. When MBA was 0.05 g, 0.1 g, 0.15 g, and 0.2 g, the equilibrium swelling rate of CMX-MNP-PNIPAm/Fe₃O₄ hydrogels was 10.47 g/g, 8.12 g/g, 7.52 g/g, and 6.44g/g, respectively, indicating that the crosslinking agent had a positive influence on the crosslinking density and accordingly had an impact on the SR of hydrogels. The deswelling behavior of the CMX-MNP-PNIPAm/Fe₃O₄ hydrogels was shown in different NaCl concentrations at 60 °C (Fig. 8b). With the increase of NaCl concentration, the capacity of hydrogels to hold water clearly decreased. When the concentration of NaCl was above 0.2 M, the water retention rate of hydrogels was less than 15% in 1 min. In addition, the water retention rate of hydrogels prepared without NaCl was clearly higher than that of

the samples prepared in the NaCl solution. This change illustrated that the NaCl had a positive effect on the deswelling behavior of the hydrogels. In the NaCl medium, the repulsion from the CMX chain was depressed by the shielding effect of the small molecular electrolytes to facilitate the curliness of the CMX chain (Annaka *et al.* 2000). Additionally, the monomers and crosslinking agents were clustered around carboxymethyl xylan to form a macroporous structure in hydrogels. After the removal of CMX, the heterogeneous porous hydrogels were obtained, which demonstrated that the CMX played a key role as the pore-forming agent in the preparation of hydrogels in NaCl solutions (Cheng *et al.* 2003).

The deswelling behavior of hydrogels at different AM amounts is shown in Fig. 8c. The water retention rate of hydrogels increased with the increase of AM amounts. When the AM amount was 0.05 M, the water retention rate of the hydrogels was less than 15% in 50 s. This was because water molecules in hydrogels were blocked and difficult to spread due to forming more hydrogen bonds by introducing more hydrophilic AM into the hydrogels network. This trend was consistent with the conclusion obtained from a previous study that the water retention of hydrogels increased with the increase of AM in 6000 s (Gao *et al.* 2015).

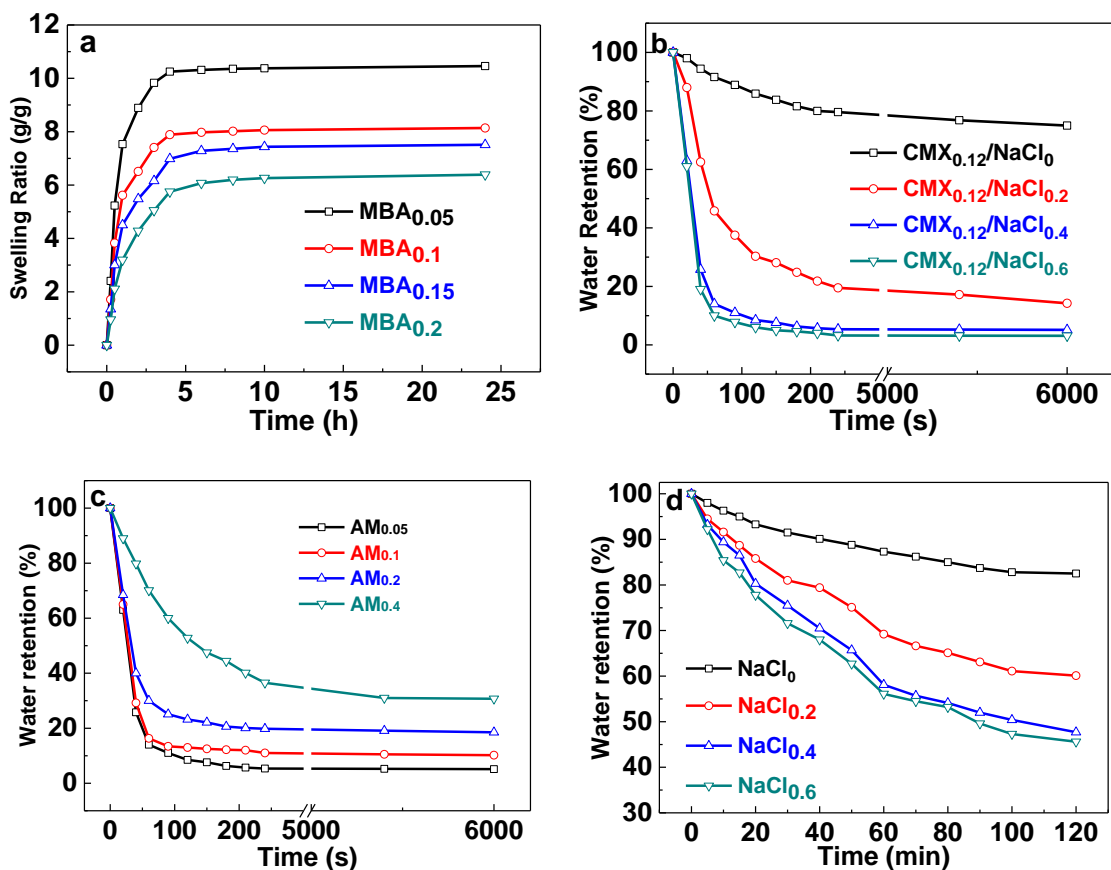


Fig. 8. The swelling behavior and deswelling behavior of CMX-MNP-PNIPAm/Fe₃O₄ hydrogels: (a) different MBA amounts at 20 °C; (b) different NaCl concentrations at 60 °C; (c) different AM amounts at 60 °C; and (d) the influence of different NaCl concentrations on deswelling behavior of the hydrogel at 60 °C without CMX

Figure 8d depicts the deswelling of hydrogels in different concentrations of NaCl solutions in the absence of CMX. Clearly, hydrogels prepared in the absence of NaCl and CMX showed a slow deswelling rate and a high water retention rate of 85% in 120 min. However, the deswelling rate of prepared hydrogels increased with the concentration of NaCl in the absence of CMX. The deswelling rate of hydrogels prepared with the addition of CMX and NaCl solutions were remarkably faster than that of hydrogels without CMX. For example, the hydrogel prepared with CMX in 0.6 M NaCl solution lost at least 85% water within 20 s, whereas the hydrogel without CMX lost only approximately 15% in 120 min. This improved property demonstrated that the CMX played an important role in the process of preparing hydrogels in NaCl solutions. The shielding effect of the NaCl small molecule made the molecular chain of CMX curl into particles that were easily removed during the washing of hydrogels. Thus, CMX acted as the pore-forming agent during the hydrogels preparation (Wu *et al.* 1992).

Photothermal Properties of Hydrogels

The photothermal properties of CMX-MNP-PNIPAm/Fe₃O₄ hydrogels were investigated in Fig. 9. In Fig. 9a, a small piece of CMX-MNP-PNIPAm/Fe₃O₄ hydrogels was placed in a quartz-type cell that contained 0.6 mL of water. Then, they were exposed to the NIR laser of 808 nm (2 W) for 2 min. The volume phase transition of hydrogels was observed under a laser irradiation in 2 min, which was due to the temperature of hydrogels being above its LCST under the photothermal effect induced by black Fe₃O₄ particles in the hydrogels (Zhu *et al.* 2014).

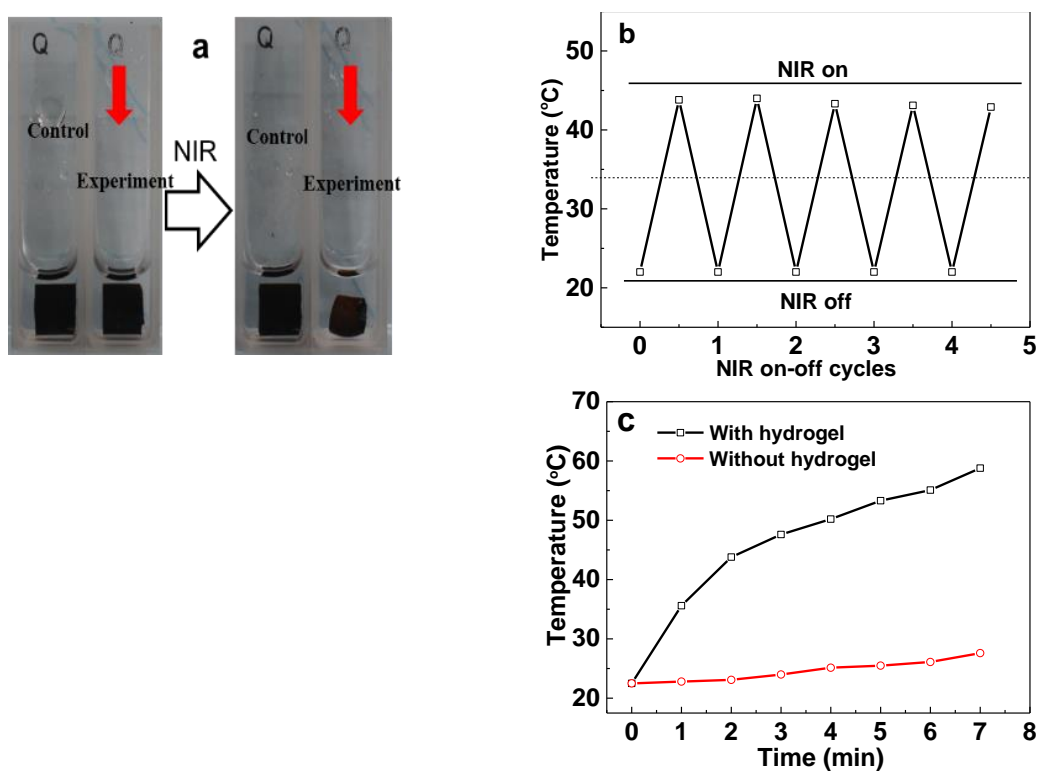


Fig. 9. The volume phase transition of hydrogels (CMX/MBA/AM/NaCl in Table 1) under the NIR irradiation (a); the change of the water temperature under the NIR on-off cycles (b); and change of water temperature under the NIR irradiation (c) (808 nm, 2 W)

After removing the laser, hydrogels recovered from a shrunken state to a swollen state like the original one, suggesting that the photothermal reversibility of hydrogels was excellent. To further confirm the photothermal reversibility of the hydrogel, the NIR irradiation experiment was repeated several times, and the results are shown in Fig. 9b. When the laser was removed after 2 min of irradiation, the temperature of the water was immediately measured with a probe. Hydrogels could be controlled to be shrunken/swollen by adjusting the NIR laser on/off, and the water temperature followed the change, suggesting that the phase transitions of the hydrogels were completely reversible. In Fig. 9c, the water temperature of CMX-MNP-PNIPAm/Fe₃O₄ hydrogels increased with the increase of irradiation time. The NIR-induced temperature increased to 59 °C in 7 min, whereas no obvious temperature variation was observed without the hydrogel at the same laser irradiation. These changes demonstrated that the prepared CMX-MNP-PNIPAm/Fe₃O₄ hydrogels could absorb and transform NIR laser into heat, and the volume phase transition under NIR irradiation was reversible. More importantly, the *in-situ* formation of Fe₃O₄ nanoparticles during the preparation of hydrogels not only endows the hydrogels with photothermal properties, but also endows them with magnetism, which is conducive to their further utilization and recovery.

CONCLUSIONS

1. Temperature fast-responsive, magnetic carboxymethylxylan-magnetic nanoparticle-poly(*N*-isopropylacrylamide)-Fe₃O₄ (CMX-MNP-PNIPAm/Fe₃O₄) hydrogels were successfully prepared using carboxymethyl xylan as the pore-forming agent in the NaCl solution. The obtained hydrogels exhibited a fast, temperature-responsive behavior and the water retention was less than 15% for 1 min under 60 °C.
2. The CMX played an important role in pore-making at the process of preparing hydrogels. The CMX-MNP-PNIPAm hydrogels with CMX showed intensive aperture networks structure and larger pore size compared with the hydrogels without CMX.
3. The lower critical solution temperature (LCST) of the CMX-MNP-PNIPAm/Fe₃O₄ hydrogels could be controlled by adjusting the hydrophilic monomer (AM) amounts. Hydrogels also showed good photothermal transformation performance and had great compression properties. The hydrogels could be heated to 40 °C within 2 min, and to 69 °C within 7 min under near infrared irradiation.

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