Rot Resistance of Tropical Wood Species Affected by Water Leaching

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Initial water leaching affects were studied relative to the rot resistance of 21 tropical wood species to the brown-rot fungus Coniophora puteana and the white-rot fungus Trametes versicolor. The rot resistance of tropical woods, determined by their weight loss (Δm) at 6-weeks fungal attack, was differently influenced by the initial water leaching, as follows: in the 1st group of eight durable species (dark red meranti, macassar ebony, cerejeira, merbau, santos rosewood, zebrano, wengé, and karri) there occurred a very significant decrease of their high rot resistance (with statistical confidence in the range 99 to 99.9% for different species); in the 2nd group of 10 durable species (ipé, yellow balau, doussié, bubinga, ovengol, padouk, iroko, blue gum, maçaranduba, and makoré) there did not occur a decrease of their high rot resistance; and in the 3rd group of three less durable species (okoumé, tineo, sapelli) a weaker rot resistance did not change due to leaching. The durable tropical woods (18 species from the 1st and 2nd groups) were more resistant to the brown-rot fungus $(\Delta m_{\text{C.p.-average}} = 0.72\%)$ than the white-rot fungus $(\Delta m_{\text{T.v.-average}} = 1.07\%)$. However, this phenomenon was no longer apparent after water leaching $(\Delta m_{\text{C.p.-Leached-average}} = 2.61\%; \Delta m_{\text{T.v.-Leached-average}} = 2.32\%).$

Keywords: Tropical woods; Water leaching; Rot resistance; Coniophora puteana; Trametes versicolor

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INTRODUCTION

The natural durability of individual wood species in moisture and temperature conditions suitable for biodegradation processes is closely related to their molecular structure, in particular, to the type, amount, and distribution of their biocidal extracts present in cell walls and lumens (Wong et al. 2005; Taylor et al. 2006; Kirker et al. 2013; Nascimento et al. 2013; Amusant et al. 2014; Sablík et al. 2016; Lopes et al. 2018). Extracts in heartwood zones of the majority of tropical wood species range from 10% to 20%, while in European species such values would be in the range 4% to 10%. According to the EN 350 (2016) standard, several tropical wood species belong to the category of very durable species (e.g., cumaru, doussié, ipé, jarrah, and maçaranduba) or durable species (e.g., bubinga, karri, lotofa, ovengol, and mahogany). Their natural durability is not only influenced by an individual wood species, but also by the location of the tree's heartwood zone, its age, and growth conditions (Amusant et al. 2014; Sundararaj et al. 2015). For example, the outer heartwood obtained from older trees is usually more resistant to rotting than the outer heartwood from younger trees (Scheffer and Cowling 1966). This is due to the gradual formation of larger quantities of biocidal extracts in the heartwood (Magel et al. 1994).

Regarding the practical application of different wood species, the following principle is valid: a required durability of wooden products (*i.e.*, natural and technologically increased) should be assigned based on the conditions of their use (EN 460 1994; Baar and Gryc 2012; EN 335 2013; Moya and Berrocal 2015; Pánek and Reinprecht 2016). For example, in exterior applications, the following woods should preferably be used: wood species with non-leachable or low-leachable biocidal extracts, wood that is chemically preserved with fixable and water-stable biocides, or thermally/chemically-modified wood that has a water-stable and biologically resistant structure.

Organic and inorganic wood extracts are mainly localized in the epithelial cells (bituminous channels) and parenchyma cells, but also in the lumens and cell walls of vessels, fibres, and tracheids. Extracts present in wood can be leached out by various solvents: polar (*e.g.*, water), medium polar (*e.g.*, ethanol, acetone), and nonpolar (*e.g.*, hexane, toluene). Heartwood zones of durable wood species contain extracts of various chemical structures, *e.g.*, monoterpenes, diterpenes, terpenoids, fats, waxes, and polyphenols such as stilbenes, tannins, flavonoids, flobafenes, rubrenolide, and rubrynolide (Rodrigues *et al.* 2010; Valette *et al.* 2017). After the initial leaching or evaporation of biocidal extracts, the original biological resistance of very durable, durable, and medium durable wood species decreases.

Several tropical wood species have a strong biological resistance to decaying fungi (Wong *et al.* 2005) and insects (Yalcin *et al.* 2018). However, this resistance is not always permanent, due to leaching or evaporation of biocidal extracts. This indirectly means that high rot resistance of durable tropical wood species can be reduced in a permanently moist environment due to water leaching of their biocidal extracts.

The aim of this work was to investigate the impact of initial water leaching of selected tropical wood species on their resistance to the brown-rot fungus *Coniophora puteana* and the white-rot fungus *Trametes versicolor*. The results from this study will have great importance in the practical use of tropical wood species in adverse environments.

EXPERIMENTAL

Materials

Tropical and reference woods

In this experiment, 21 tropical wood species (Table 1) and 2 reference European wood species, beech (*Fagus sylvatica* L.) and Scots pine (*Pinus sylvestris* L.), were used. Tropical woods used in the experiment, with names by Association Technique Internationale des Bois Tropicaux (ATIBT) in France, were obtained from the trading company JAF Holz, Ltd. (Špačince, Slovakia), in the form of naturally dried and conditioned boards with a moisture content of $13 \pm 2.5\%$. The classes of durability, according to EN 350 (2016), of the tropical wood species used are between 1 "very durable" (Table 1).

Micro-specimens with dimensions of 25 mm \times 25 mm \times 3 mm (longitudinal \times radial \times tangential) were prepared from the heartwood zone of the tropical wood species and the sapwood zone of the reference wood species. The micro-specimens had no biological damage, knots, or other inhomogeneities. The experiment was performed with 16 micro-specimens from each tropical wood species and with 84 micro-specimens from each reference wood species.

Table 1. 7	Fropical	Woods	Used in	the Ex	<i>cperiment</i>
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Species	Species	Density ⁴	Class of Durability as			
Common Name	Scientific Name	(kg/m³)	per EN	350 (2016)		
			Field	Laboratory		
Inó	Handroanthus corratifolius	068 (26)	1 1 1	Test		
ipe	(Vahl) S. O. Grose ³	900 (20)	I			
Okoumé	<i>Aucoumea klaineana</i> Pierre	566 (27)	4	4 to 5		
Tineo	Weinmannia trichosperma Cav.	646 (31)				
Dark red meranti	Shorea curtisii Dyer ex. King	592 (10)	2 to 4	2		
Yellow balau ¹	Shorea laevis Ridl.	925 (55)	2			
Macassar ebony	Diospyros celebica Bakh.	1013 (82)				
Doussié	Afzelia bipindensis Harms	889 (18)	1			
Cerejeira	Amburana cearensis A. C. Sm.	651 (10)	3			
Bubinga	<i>Guibourtia demeusei</i> J. Léonard	830 (13)	2			
Ovengol	<i>Guibourtia ehie</i> J. Léonard	755 (27)	2			
Merbau	Intsia bijuga O. Ktze.	837 (50)	1 to 2	1		
Santos rosewood	Machaerium scleroxylon Tul.	904 (6)				
Zebrano	Microberlinia brazzavillensis Chev.	718 (23)				
Wengé	<i>Millettia laurentii</i> De Wild.	823 (37)	2	1		
Padouk	<i>Pterocarpu</i> s soyauxii Taub.	647 (37)	1			
Sapelli	Entandrophragma cylindricum Sprague	631 (38)	3	3 to 4		
Iroko	<i>Milicia excelsa</i> C. C. Berg	551 (16)	1 to 2			
Karri	Eucalyptus diversicolor F. Muell.	804 (30)	2			
Blue gum	Eucalyptus globulus Labill.	760 (59)	1 to 2	1		
Maçaranduba ²	<i>Manilkara bidentata</i> A. Chev.	916 (19)	1			
Makoré	<i>Tieghemella heckelii</i> Pierre	570 (25)	1			
¹ Other commonly used name is Bangkirai; ² its other commonly used name is						
Massaranduba; ³ registered in The IUCN Red List of Threatened Species™ (IUCN 2017);						
⁴ Mean values of densities are from 16 micro-specimens, at which standard deviations are						
in parentheses (Vidholdová and Reinprecht 2019)						

Methods

Water Leaching

Eight micro-specimens from each tropical wood species, and 42 micro-specimens from each reference wood species (with an input moisture content of $12\% \pm 2\%$), were leached in distilled water over 14 d, using 9 leaching cycles at a 1:5 hydro module ratio, according to conditions described in the EN 84 (1997) standard.

Fungal attack

Micro-specimens of the individual tropical and reference wood species (both native and initially water-leached) were subjected to attack by the brown-rot fungus *Coniophora puteana* (Schumacher ex Fries) Karsten, strain BAM Ebw. 15 (Bundesanstalt für Materialforschung und -prüfung, Berlin, Germany) or to attack by the white-rot fungus *Trametes versicolor* (Linnaeus ex Fries) Pilat, strain BAM 116 (Bundesanstalt für Materialforschung und -prüfung, Berlin, Germany).

Before the fungal attacks, the native and water-leached micro-specimens were conditioned to achieve a moisture content of $12\% \pm 1\%$, weighed with an accuracy of 0.001 g (m_i), and sterilized with a 30 W germicidal lamp (Chirana Medical, a.s., Stará Turá, Slovakia) at a temperature of 22 ± 2 °C for 20 min on each side.

The fungal attacks of micro-specimens were performed in glass Petri dishes with a diameter of 100 mm. Two replicates of the same tropical wood species, and one replicate of the reference wood species, were placed into one dish on plastic mats under which a fungal mycelium had already been grown on a sterilized and a solidified layer of 4.5 wt% malt agar medium (HiMedia, Ltd., Mumbai, India) with a thickness from 3 mm to 4 mm. Malt agar medium was sterilized in autoclave (Autoclave PS 121V, Chirana, Brno, Czech Republic) at a temperature of 118 °C for 15 minutes. The incubation of micro-specimens in fungal mycelia lasted 6 weeks at a temperature of 24 ± 2 °C, and a relative humidity of 90% \pm 5% according the rapid screening test (Van Acker *et al.* 2003).

After the fungal attacks, the micro-specimens were carefully cleaned of surface fungal mycelia, slowly conditioned in the laboratory to their moisture content of $12\% \pm 1\%$, and weighed again with an accuracy of 0.001 g (m_f). The weight loss (Δm) of micro-specimens was calculated as a percentage from their weights (determined in grams) in the conditioned state before (m_i) and after (m_f) fungal attack according to the standard EN 113 (1996), using Eq. 1:

$$\Delta m = \left[\left(m_{\rm i} - m_{\rm f} \right) / m_{\rm i} \right] \times 100 \tag{1}$$

Statistical analyses

The effect of initial water leaching of the tropical wood species on changes in their rot resistance, which was determined using values of Δm , was analyzed by Duncan tests applying Statistica v. 12 software (StatSoft CR, s.r.o., Prague, Czech Republic).

RESULTS AND DISCUSSION

The rot resistance of 21 tropical wood species to the decaying fungi *C. puteana* and *T. versicolor* was dependent on a few factors, as documented in Table 2. These were as follows: (1) the wood species, (2) application of the initial water leaching, and (3) the

fungal species. Visualization of the fungal mycelia growth on the top surfaces of two selected tropical wood species in their native and water-leached state is shown in Fig. 1.

Table 2. Weight Losses of Native and Initially Water-leached Tropical andReference Woods after 6 Weeks' Action of the Brown-rot Fungus *C. puteana* orthe White-rot Fungus *T. versicolor*

	Weight Loss Δm (%) Caused by Decay Fungus									
Wood	Coniophora puteana				Trametes versicolor					
Species	Native Wood		Water- leached Wood		Duncan Test	Native Wood		Water- leached Wood		Duncan Test
	х	SD	х	SD		х	SD	х	SD	
lpé	0.20	(0.03)	0.61	(0.29)	С	1.39	(1.21)	0.32	(0.26)	_
Okoumé	6.48	(2.98)	5.39	(1.83)	_	6.39	(1.05)	8.23	(1.78)	d
Tineo	5.38	(3.71)	6.00	(2.87)	d	5.35	(1.77)	2.90	(0.36)	_
Dark red meranti	0.22	(0.09)	8.51	(1.70)	а	0.60	(0.27)	1.59	(0.30)	b
Yellow balau	0.40	(0.08)	0.69	(0.46)	d	0.83	(0.23)	0.30	(0.10)	_
Macassar ebony	0.52	(0.30)	4.68	(0.45)	а	2.00	(0.64)	6.84	(1.82)	а
Doussié	1.63	(1.51)	0.53	(0.30)	_	0.73	(0.25)	0.23	(0.11)	_
Cerejeira	0.30	(0.15)	1.05	(0.30)	b	0.80	(0.18)	2.21	(0.94)	b
Bubinga	0.96	(0.13)	0.75	(0.19)	_	1.17	(0.23)	0.66	(0.08)	_
Ovengol	0.61	(1.02)	1.30	(0.04)	d	1.34	(0.15)	1.60	(0.33)	d
Merbau	0.48	(0.32)	5.36	(1.07)	а	0.99	(1.02)	5.33	(1.08)	а
Santos rosewood	1.28	(0.31)	4.67	(0.61)	а	1.70	(0.20)	4.46	(0.23)	а
Zebrano	1.65	(0.62)	4.81	(0.72)	а	0.62	(0.30)	2.33	(0.66)	b
Wengé	0.67	(0.35)	5.84	(1.27)	а	0.99	(0.31)	6.40	(0.91)	а
Padouk	0.95	(1.69)	0.66	(0.25)	_	0.76	(0.07)	0.32	(0.12)	_
Sapelli	3.12	(1.15)	4.72	(2.36)	d	2.55	(0.92)	1.79	(0.17)	_
Iroko	0.20	(0.97)	0.47	(0.52)	d	0.41	(0.16)	1.69	(0.50)	b
Karri	0.69	(0.47)	2.50	(0.36)	а	1.18	(0.21)	2.38	(0.21)	а
Blue gum	0.38	(0.21)	2.30	(0.92)	b	1.82	(0.60)	2.17	(0.43)	d
Maçaranduba	0.08	(0.05)	0.23	(0.05)	b	0.52	(0.40)	0.22	(0.03)	_
Makoré	1.78	(1.84)	1.96	(0.85)	d	1.37	(0.08)	2.70	(1.12)	с
Beech						10.64	(3.65)	11.45	(3.21)	d
Pine	17.01	(3.71)	16.78	(3.25)	_					
x - mean weight loss values were from 4 replicates of tested tropical wood species and from 42 replicates of reference European wood species; SD - standard deviations are in parentheses; a, b, c, d - indexes of the Duncan test characterizing the significance level of water leaching on the decreased rot resistances (<i>i.e.</i> , increased weight loss) of tropical and reference woods (a = very significant decrease of rot resistance > 99.9%, b = significant decrease > 99%, c = less significant decrease > 95%, d = insignificant decrease < 95%, and = even an increase of rot resistance)										

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Fig. 1. Examples of the fungal mycelia growth on the top surfaces of the native and waterleached tropical woods after 6 weeks of incubation. In the Petri dish are 2 micro-specimens of the tested tropical wood species and at the bottom is 1 micro-specimen of reference Scots pine sapwood (for *C. puteana*) or beech sapwood (for *T. versicolor*).

Rot Resistance of Native Tropical Woods

The rot resistance of 18 durable tropical wood species (ipé, dark red meranti, yellow balau, macassar ebony, doussié, cerejeira, bubinga, ovengol, merbau, santos rosewood, zebrano, wengé, padouk, iroko, karri, blue gum, maçaranduba, and makoré) was excellent in their native state, because their weight losses only ranged from 0.08% to 2% (Table 2). These durable wood species exhibited a stronger resistance to the brown-rot fungus *C*. *puteana* ($\Delta m_{\text{C.p.-}}$ average = 0.72%) compared to the white-rot fungus *T*. *versicolor* ($\Delta m_{\text{T.v.-}}$ average = 1.07%). A relatively more intensive attack of durable tropical wood species by *T*. *versicolor* is in the literature attributed to peroxidase and laccase enzymes produced by this fungus. These enzymes are able to catalyze degradation processes not only in lignin, but also in polyphenols, flavonoids, quinones, and some other biocidal extracts (Eriksson *et al.* 1990).

The rot resistance of 3 less durable tropical wood species (okoumé, tineo, and sapelli) was weaker. A slightly lower rot resistance compared to durable species was shown by sapelli (Δm approximately 3%). However, okoumé and tineo had an apparently lower rot resistance (Δm approximately 6%) (Table 2). In summary, these 3 wood species had a comparative resistance to *C. puteana* ($\Delta m_{C.p.-average} = 4.99\%$) and *T. versicolor* ($\Delta m_{T.v.-average} = 4.76\%$) (Table 2). Nevertheless, their resistance to decay processes was approximately from two-times to three-times higher in comparison with the sapwood zones of the reference beech and Scots pine species (Table 2).

Rot Resistance of Water-leached Tropical Woods

The initial water leaching of 21 tropical woods had different effects on their rot resistance (Table 2).

After water leaching, an apparently lower rot resistance to both decaying fungi *C. puteana* and *T. versicolor* were observed in the 1st group of the following eight durable tropical wood species: dark red meranti, macassar ebony, cerejeira, merbau, santos rosewood, zebrano, wengé, and karri. This group includes durable species with a potentially decreased rot resistance in permanent wet exterior conditions. A reduced rot resistance of these tropical species after water leaching, where their weigh losses caused by fungal attacks increased more than 2% (*C. puteana*: $\Delta m_{C.p.-average} = 0.73\%$; $\Delta m_{C.p.-Leached-average} = 4.68\%$; *T. versicolor*: $\Delta m_{T.v.-average} = 1.11\%$; $\Delta m_{T.v.-Leached-average} = 3.94\%$), was confirmed by the Duncan tests at a 99.9% or 99% level of significance (Table 2). The experimental results were in accordance with the works of Wanschura *et al.* (2014) and other researchers (Table 3). They found that some biocidal extracts presented in these tropical wood species are water soluble, so the initial water leaching process decreased their original decay resistance.

Conversely, a relatively unchanged rot resistance after water leaching was observed for the 2nd group of the following 10 durable tropical wood species: ipé, yellow balau, doussié, bubinga, ovengol, padouk, iroko, blue gum, maçaranduba, and maroré. This 2nd group included durable wood species with a potentially stable resistance under wet exterior conditions. Tropical wood species of this group usually had a comparable decay resistance before and after water leaching (*C. puteana*: $\Delta m_{\text{C.p.-average}} = 0.719\%$; $\Delta m_{\text{C.p.-Leached-average}} =$ 0.95%; *T. versicolor*: $\Delta m_{\text{T.v.-average}} = 1.034\%$; $\Delta m_{\text{T.v.-Leached-average}} = 1.021\%$), as confirmed by the Duncan tests (Table 2). Due to the initial water leaching, only blue gum and makoré woods had a lower resistance to decaying fungi with $\Delta m > 2\%$ (Table 2). In summary, no (or only smaller) decreases in the rot resistance of water-leached tropical woods of the 2nd group could be explained by hypothesis 1 that some extracts of durable tropical wood species have a high fungicidal efficiency and simultaneously have strong stability against water leaching.

Rot resistance of the 3rd group of 3 less durable tropical wood species (okoumé, tineo, and sapelli) was not greatly decreased due to the initial water leaching processes (Table 2). This result was explained by hypothesis 2 that water insoluble and soluble extracts of less durable tropical wood species do not have a more apparent fungicidal efficiency (Tables 2 and 3).

The authors wanted to confirm hypotheses 1 and 2 in prepared experiments that analyzed the chemical substances in water and organic-solvent extracts from selected tropical wood species and their efficiency against decaying fungi.

Rot Resistance of Investigated Tropical Woods by Other Researchers

Several of the tropical wood species researched in this work have a known chemical structure (including information about the type and amount of extracts), anatomical structure, and physical-mechanical properties (Porter 2006; Wagenführ 2007). In the following section, the rot resistance of some tropical wood species researched in this work is discussed, both in their native and water-leached states.

Table 3. Water Soluble Extracts Presented in Some of the Researched Tropical Woods

Wood Species	Extracts					
	Water Soluble	References				
Iné	5.1% in water	Wanschura et al. (2014)				
ipe	12.63% in hot water	Jankowska <i>et al.</i> (2018)				
Dark red meranti	1.0% to 2.1% in hot water	Yamamoto and Hong (1989)				
Mallaw kalaw	5.2% in water	Wanschura et al. (2014)				
Yellow balau	1.9% to 7.4% in hot water	Yamamoto and Hong (1989)				
	4.16% in cold water	Kilic and Niemz (2012)				
	2.85% in cold water,	Arslan <i>et al.</i> (2018)				
Devezić	2.40% in hot water	Fan <i>et al.</i> (2012)				
Doussie	7.99% in hot water	Jankowska <i>et al.</i> (2018)				
	10.30% in hot water	Kilic and Niemz (2012)				
	5.99% in hot water	Arslan <i>et al.</i> (2018)				
Coroioiro	10.8% in cold water	da Silva Oliveira et al. (2005)				
Cerejeira	17.4% in hot water	da Silva Oliveira <i>et al</i> . (2005)				
Bubinga	6.86% in hot water	Mahút <i>et al.</i> (1995)				
Ovengol	5.93% in hot water	Jankowska <i>et al.</i> (2018)				
	2.88% in cold water	Kilic and Niemz (2012)				
	6.1% in water	Wanschura <i>et al.</i> (2014)				
Merbau	8.84% in hot water	Kilic and Niemz (2012)				
	12.5% to 16.5% in hot water	Yamamoto and Hong (1989)				
	17.29% in hot water	Jankowska <i>et al.</i> (2018)				
Zohrano	2.42% in cold water	Kilic and Niemz (2012)				
Zebrano	3.41% in hot water	Kilic and Niemz (2012)				
	3.11% in cold water	Kilic and Niemz (2012)				
Wengé	4.33% in hot water	Jankowska <i>et al.</i> (2018)				
	5.89% in hot water	Kilic and Niemz (2012)				
Padouk	5.73% in hot water	Jankowska <i>et al.</i> (2018)				
	5.84% in hot water	Fan <i>et al.</i> (2012)				
	2.59% in cold water	Arslan <i>et al.</i> (2018)				
Sapelli	1.19% in hot water	Arslan <i>et al.</i> (2018)				
Sapelli	5.90% in hot water	Fan <i>et al.</i> (2012)				
	6.07% in hot water	Jankowska <i>et al.</i> (2018)				
	4.45% in cold water	Arslan <i>et al.</i> (2018)				
Iroko	3.93% in hot water	Arslan <i>et al.</i> (2018)				
IIOKO	6.47% in hot water	Jankowska <i>et al.</i> (2018)				
	8.64% in hot water	Fan <i>et al.</i> (2012)				
Maçaranduba	1.4% in hot water	Wanschura et al. (2014)				

Ipé wood has been found to belong to the very durable species group. For example, Lee *et al.* (2013) found that ipé has a high resistance to the white-rot fungus *T. versicolor* ($\Delta m = 0.41\%$ after 12 weeks) as well as to several other brown-rot fungi (*Gloeophyllum trabeum*: $\Delta m = 1.59\%$ after 12 weeks; *Fomitopsis palustris*: $\Delta m = 2.69\%$ after 12 weeks). Additionally, Andres *et al.* (2015) determined a high resistance of ipé to the brown-rot fungus *Serpula lacrymans* ($\Delta m = 3.67\%$ after 16 weeks). Heartwood of ipé contains a remarkable portion of the biocidal extract naphthoquinone "lapachol" (Romagnoli *et al.* 2013), providing it with a high resistance to damaging fungi and termites (Arango *et al.* 2006; Nascimento *et al.* 2013; Futuro *et al.* 2018).

Dark red meranti (*Shorea* spp.) wood grows in different regions of the world. It has many species; therefore, it is often characterized as durable, medium durable, or less durable (Table 1), with a durability ranging from 2 to 4 according to EN 350 (2016). Some

researchers have established that this wood species has a low decay resistance to *T*. *versicolor* (Takahashi and Kishima (1973): $\Delta m = 21.6\%$, and Da Silva Oliveira *et al.* (2005): $\Delta m = 14.5\%$). Conversely, dark red meranti researched in this work had a high resistance to *T. versicolor* (Table 2: $\Delta m = 0.60\%$). Roszaini *et al.* (2016), in accordance with the present experiment (Table 2), also found a high decay resistance of dark red meranti to *T. versicolor* ($\Delta m = 1.65\%$). Based on these different results, the dark red meranti was again confirmed as highly variable in its durability, which depends on the tree growth conditions and other factors.

Yellow balau and merbau wood species were both highly resistant to T. versicolor (Table 2: $\Delta m = 0.83\%$ and 0.99%, respectively), basically in agreement with the experiments of Yamamoto and Hong (1989) ($\Delta m = 1.2\%$ and 3.8%, respectively). A comparable high resistance of these two tropical species to T. versicolor is also documented by Takahashi and Kishima (1973), determining similar weight losses for balau originating from Indonesia ($\Delta m = 2.60\%$) or Singapore ($\Delta m = 1.88\%$) and for merbau ($\Delta m = 2.47\%$). According to Lee et al. (2013), merbau can have a lower resistance to white-rot fungi (e.g., *T. versicolor*: $\Delta m = 5.87\%$ after 12 weeks) compared to brown-rot fungi (*e.g.*, *G. trabeum*: $\Delta m = 1.19\%$). However, the present experiment with *T. versicolor* and *C. puteana* (Table 2: $\Delta m < 1.0\%$) did not correspond with their findings. Yamamoto and Hong (1989) found that under the action of *T. versicolor*, the rot resistance of balau remained unchanged in response to its initial hot-water leaching (Δm remained low, 1.2%). However, rot resistance evidently worsened for water-leached merbau (Δm increased from 3.8% to 22.7%). These findings were in accordance with the present experiment (Table 2). Merbau wood contains extracts of a phenolic nature (Hofmann et al. 2011; Kilic and Niemz 2012), which are relatively easily washed out with water (Yamamoto and Hong 1989). Therefore, in line with previous mycological tests with merbau wood, and with knowledge related to the chemical structure of its extracts, the authors of this article determined that water leaching significantly deteriorated its decay resistance to T. versicolor (Table 2: Δm increased from 0.99% to 5.33%) and *C. puteana* (Table 2: ∆*m* increased from 0.48% to 5.36%).

According to Da Silva Oliveira *et al.* (2005), cerejeira is a tropical species that is highly resistant to *G. trabeum* (Δm approximately 2%), both with and without previous leaching in water. Conversely, this study indicated that its initial water leaching caused a partial but significant drop in its decay resistance to *C. puteana* and *T. versicolor* (Table 2: Δm increased from 0.3% to 1.05% and from 0.8% to 2.21%, respectively).

Bubinga and wengé, in their native state, have a high rot resistance to *C. puteana* and *T. versicolor* (Table 2). Van Acker *et al.* (2003) also reported a high decay resistance of wengé to *C. puteana*, determining its small weight loss (0.2%) by the mini-mycological test on micro-specimens, or only a slightly higher weight loss (0.4%) by the standardized EN 113 (1996) mycological test. However, wengé (as opposed to bubinga) probably contained some water-leachable extracts, as its rot resistance significantly decreased after the initial water leaching (Table 2).

Padouk and iroko belonged to the very durable species group, with durability classes of 1 and 1 to 2 according to EN 350 (2016). Reinprecht *et al.* (2012) found that after 16 weeks of decay attack on 0.6-mm-thick veneers made from padouk and iroko, both of these durable wood species exhibited a strong resistance to the brown-rot fungi *Serpula lacrymans* ($\Delta m = 2.54\%$ and 1.35\%, respectively) and *C. puteana* ($\Delta m = 0.88\%$ and 3.51\%, respectively). The high durability of padouk, according to Déon and Schwartz (1988) and Surowiec *et al.* (2004), can be attributed to pterocarane, deoxysanthalene, sandaline, and

other santal analog extracts. These biocidal extracts of padouk wood were probably waterproof, because its rot resistance had not deteriorated after water leaching (Table 2: $\Delta m < 1\%$).

Generally, the weight loss parameter, which is often used for assessing the rot resistance of wood, is greatly affected by the wood species and its chemical structure, mainly by the presence of biocidal extracts. According to Valette *et al.* (2017), the antifungal efficacy of various extractive compounds present in wood species and other plants is based on the following multiple mechanisms: (1) capture of Fe^{2+} and other metal ions or free radicals, (2) binding of degrading enzymes produced by the fungi, (3) direct attack and destruction of the cell wall and plasma membrane of the fungi, (4) failure of ion homeostasis in the fungi, and (5) chaotropic activity, for example, connected with inhibition of microbial growth and metabolism of more compounds in extracts such as alcohols and phenols.

However, rot processes in the molecular structure of wood species are also influenced by exposure conditions (such as temperature, moisture, and pH value) and their duration. Simultaneously, from evaluation of results achieved through mycological tests performed in a laboratory, high importance should also be assigned to some other factors. First, the dimensions of the wood had a significant effect. For example, thin veneers were usually attacked more easily and quicker by decaying fungi. Second, the actual amount of specific biocidal extracts in wood should be considered. For example, tropical woods were usually attacked faster by white-rot fungi than by brown-rot fungi, because lignin peroxidase, laccases, and other oxidation enzymes produced by white-rot fungi were able to degrade, except for lignin, also phenolic, and some other types of biocidal extracts (Barz and Weltring 1985; Eriksson *et al.* 1990; Couto and Herrera 2006; Fackler *et al.* 2007). Third, the exposition of wood before durability tests is important. For example, some tropical wood species initially exposed to weathering processes often lost water-leachable biocidal extracts, and their original decay resistance was decreased. This was more evident with thinner specimens.

CONCLUSIONS

- 1. The 18 durable tropical wood species in their native state (ipé, dark red meranti, yellow balau, macassar ebony, doussié, cerejeira, bubinga, ovengol, merbau, santos rosewood, zebrano, wengé, padouk, iroko, karri, blue gum, maçaranduba, and makoré) were by means of 6-weeks mycological tests determined as highly rot resistant, with weight losses $\leq 2\%$. Three tropical woods (okoumé, tineo, and sapelli) were characterized as being less rot resistant, with weight losses from 3% to 6%.
- 2. The initial water leaching significantly decreased the high rot resistance of 8 durable tropical species: dark red meranti, macassar ebony, cerejeira, merbau, santos rosewood, zebrano, wengé, and karri. This was probably due to the leaching of their biocidal extracts.
- 3. The initial water leaching had no apparent negative impact on the high rot resistance of 10 durable tropical wood species: ipé, yellow balau, doussié, bubinga, ovengol, padouk, iroko, blue gum, maçaranduba, and makoré. These tropical woods could be applied in more demanding expositions without fungicidal treatment.

- 4. The initial water leaching did not affect a poorer rot resistance of 3 less durable tropical wood species: okoumé, tineo, and sapelli.
- 5. Without water leaching, several tropical wood species had a partly higher resistance to the brown-rot fungus *C. puteana* compared to the white-rot fungus *T. versicolor*. This was in accordance with the knowledge that white-rot fungi are able also to deteriorate the phenolic or other biocidal extracts

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REFERENCES CITED

- Amusant, N., Nigg, M., Thibaut, B., and Beauchêne, J. (2014). "Diversity of decay resistance strategies of durable tropical woods species: *Bocoa prouacencsis* Aublet, *Vouacapoua americana* Aublet, *Inga alba* (Sw.) Wild.," *Int. Biodeter. Biodegr.* 94, 103-108. DOI: 10.1016/j.ibiod.2014.06.012
- Andres, B., Jankowska, A., Kloch, M., Mazurek, A., Oleksiewicz, A., Pałucki, M., and Wójcik, A. (2015). "A study of natural durability of wood in selected tropical wood species from South America and Africa affected by the fungus *Serpula lacrymans* (Wulf., Fr.) Schroet.," *Forestry and Wood Technology* 92, 11-17.
- Arango, R. A., Green, F., Hintz, K., Lebow, P. K., and Miller, R. B. (2006). "Natural durability of tropical and native woods against termite damage by *Reticulitermes flavipes* (Kollar)," *Int. Biodeter.*. *Biodegradr*. 57(3), 146-150. DOI: 10.1016/j.ibiod.2006.01.007
- Arslan, M., Tascioglu, C., Kose, C., Terzi, E., and Akcay, C. (2018). "Yıkanmanın Bazı İthal Ağaç Türlerinde Çürüklük Mantarlarına Karşı Doğal Dayanıklılığa Etkisi," *Duzce University Journal of Science and Technology*, 6(1), 263-274. DOI: 10.29130/dubited.331252
- Baar, J., and Gryc, V. (2012). "The analysis of tropical wood discoloration caused by simulated sunlight," *Eur. J. Wood Wood Prod.* 70(1-3), 263-269. DOI: 10.1007/s00107-011-0551-1
- Barz, W., and Weltring, K.-M. (1985). "Biodegradation of aromatic extractives of wood," in: *Biosynthesis and Biodegradation of Wood Components*, T. Higuchi (ed.), Academic Press, Cambridge, MA, USA, pp. 607-666.
- Couto, S. R., and Herrera, J. L. T. (2006). "Industrial and biotechnological applications of laccases: A review," *Biotechnol. Adv.* 24(5), 500-513. DOI: 10.1016/j.biotechadv.2006.04.003
- Da Silva Oliveira, J. T., Chagas de Souza, L., Della Lucia, R. M., and de Souza Junior, W. P. (2005). "Influence of extracts in decay resistance of six wood species," *Rev. Árvore* 29(5), 819-826. DOI: 10.1590/S0100-67622005000500017

- Déon, G., and Schwartz, R. (1988). "Natural resistance of tropical timbers to biological attacks," B. Soc. Bot. Fr.-Actual. 135(3), 37-48. DOI: 10.1080/01811789.1988.10826900
- EN 84 (1997). "Wood preservatives. Accelerated ageing of treated wood prior to biological testing. Leaching procedure," European Committee for Standardization, Brussels, Belgium.
- EN 113 (1996). "Wood preservatives. Test method for determining the protective effectiveness against wood destroying basidiomycetes. Determination of the toxic values," European Committee for Standardization, Brussels, Belgium.
- EN 335 (2013). "Durability of wood and wood-based products. Use classes: Definitions, application to solid wood and wood-based products," European Committee for Standardization, Brussels, Belgium.
- EN 350 (2016). "Durability of wood and wood-based products. Testing and classification of the durability to biological agents of wood and wood-based materials," European Committee for Standardization, Brussels, Belgium.
- EN 460 (1994). "Durability of wood and wood-based products. Natural durability of solid wood. Guide to the durability requirements for wood to be used in hazard classes," European Committee for Standardization, Brussels, Belgium.
- Eriksson, K.-E. L., Blanchette, R. A., and Ander, P. (1990). *Microbial and Enzymatic Degradation of Wood and Wood Components*, Springer-Verlag Berlin Heidelberg, Berlin, Germany.
- Fackler, K., Gradinger, C., Schmutzer, M., Tavzes, C., Burgert, I., Schwanninger, M., Hinterstoisser, B., Watanabe, T., and Messner, K. (2007). "Biomodification of wood with selective white-rot fungi," *Food Technol. Biotech.* 45(3), 269-276.
- Fan, M., Ndikontar, M. K., Zhou, X., and Ngamveng, J. N. (2012). "Cement-bonded composites made from tropical woods: Compatibility of wood and cement," *Constr. Build. Mater.* 36, 135-140. DOI: 10.1016/j.conbuildmat.2012.04.089
- Futuro, D. O., Ferreira, P. G., Nicoletti, C. D., Borba-Santos, L. P., Da Silva, F. C., Rozental, S., and Ferreira, V. F. (2018). "The antifungal activity of naphtoquinones: An integrative review," *An. Acad. Bras. Cienc.* 90(1), 1187-1214. DOI: 10.1590/0001-3765201820170815
- Hofmann, T., Niemz, P., and Levente, A. (2011). "HPTLC assessment of phenolic extractives in selected extraneous woods," *JPC-J. Planar Chromat.* 24(6), 539-540. DOI: 10.1556/JPC.24.2011.6.16
- International Union for Concervation of Nature (IUCN) (2017). "The IUCN red list of threatened species: Version 2017-2," (http://www.iucnredlist.org), Accessed on 17 Oct 2017.
- Jankowska, A., Boruszewski, P., Drożdżek, M., Rębkowski, B., Kaczmarczyk, A., and Skowrońska, A. (2018). "The role of extractives and wood anatomy in the wettability and free surface energy of hardwoods," *BioResources* 13(2), 3082-3097. DOI: 10.15376/biores.13.2.3082-3097
- Kilic, A., and Niemz, P. (2012). "Extractives in some tropical woods," *Eur. J. Wood Wood Prod.* 70(1-3), 79-83. DOI: 10.1007/s00107-010-0489-8
- Kirker, G. T., Blodgett, A. B., Arango, R. A., Lebow, P. K., and Clausen, C. A. (2013).
 "The role of extractives in naturally durable wood species," *Int. Biodeter. Biodegradr.* 82, 53-58. DOI: 10.1016/j.ibiod.2013.03.007

- Lee, H. T., Lee, J., Choi Y.-S., Hong, J.-H., Min, M., Kim, J.-J., and Kim, G.-H. (2013). "Antifungal properties of methanol extracts from some tropical hardwoods," (http://210.101.116.28/W_files/kiss6/06804249_pv.pdf), Accessed 17 Oct 2017.
- Lopes, D. J. V., Benigno Paes, J., and Dos Santos Bobadilha, G. (2018). "Resistance of *Eucalyptus* and *Corymbia* treated woods against three fungal species," *BioResources* 13(3), 4964-4972. DOI: 10.15376/biores.13.3.4964-4972
- Magel, E., Jay-Allemand, C., and Ziegler, H. (1994). "Formation of heartwood substances in the stemwood of *Robinia pseudoacacia* L. II. Distribution of nonstructural carbohydrates and wood extractives across the trunk," *Trees* 8(4), 165-171. DOI: 10.1007/BF00196843
- Mahút, J., Bučko, J., Čunderlík, I., Hurda, B., Kurjatko, S., Réh, R., and Šupín, M. (1995). Lesser Known and Less Used Tropical Woods: Andoung, Bossé, Bubinga, White Lauan, Red Lauan, Mutenye, Monograph, Technical University in Zvolen, Zvolen, Slovakia.
- Moya, R., and Berrocal, A. (2015). "Evaluation of biodeterioration and the dynamic modulus of elasticity of wood in ten fast-growing tropical species in Costa Rica exposed to field testing," *Wood Res. (Bratislava, Slovakia)* 60(3), 359-374.
- Nascimento, M. S., Santana, A. L. B. D., Maranhão, C. A, Oliveira, L. S., and Bieber, L. (2013). "Phenolic extractives and natural resistance of wood," in: *Biodegradation -Life of Science*, R. Chamy, and F. Rosenkranz (eds.), InTechOpen, London, United Kingdom, pp. 349-370. DOI: 10.5772/56358
- Pánek, M., and Reinprecht, L. (2016). "Effect of vegetable oils on the colour stability of four tropical woods during natural and artificial weathering," *J. Wood Sci.* 62(1), 74-84. DOI: 10.1007/s10086-015-1519-2
- Porter, T. (2006). *Wood: Identification & Use (Revised & Expanded)*, GMC Publications, Lewes, United Kingdom.
- Reinprecht, L., Kmeťová, L., and Iždinský, J. (2012). "Fungal decay and bending properties of beech plywood overlaid with tropical veneers," *J. Trop. For. Sci.* 24(4), 490-497.
- Rodrigues, A. M. S., Theodoro, P. N. E. T., Eparvier, V., Basset, C., Silva, M. R. R., Beauchêne, J., Espíndola, L. S., and Stien, D. (2010). "Search for antifungal compounds from the wood of durable tropical trees," *J. Nat. Prod.* 73(10), 1706-1707. DOI: 10.1021/np1001412
- Romagnoli, M., Segoloni, E., Luna, M., Margaritelli, A., Gatti, M., Santamaria, U., and Vinciguerra, V. (2013). "Wood color in Lapacho (*Tabebuia serratifolia*): Chemical composition and industrial implications," *Wood Sci. Technol.* 47(4), 701-716. DOI: 10.1007/s00226-013-0534-y
- Roszaini, K., Hale, M. D., and Salmiah, U. (2016). "*In-vitro* decay resistance of 12 Malaysian broadleaf hardwood trees as a function of wood density and extractives compounds," *J. Trop. For. Sci.* 28(4), 533-540.
- Sablík, P., Giagli, K., Pařil, P., Baar, J., and Rademacher, P. (2016). "Impact of extractive chemical compounds from durable wood species on fungal decay after impregnation of nondurable wood species," *Eur. J. Wood Wood Prod.* 74(2), 231-236. DOI: 10.1007/s00107-015-0984-z
- Scheffer, T. C., and Cowling, E. B. (1966). "Natural resistance of wood to microbial deterioration," *Annu. Rev. Phytopathol.* 4, 147-168. DOI: 10.1146/annurev.py.04.090166.001051

- Sundararaj, R., Shanbhag, R. R., Nagaveni, H. C., and Vijayalakshmi, G. (2015).
 "Natural durability of timbers under Indian environmental conditions An overview," *Int. Biodeter. Biodegradr.* 103, 196-214. DOI: 10.1016/j.ibiod.2015.04.026
- Surowiec, I., Nowik, W., and Trojanowicz, M. (2004). "Identification of "insoluble" red dyewoods by high performance liquid chromatography-photodiode array detection (HPLC-PDA) fingerprinting," J. Sep. Sci. 27(3), 209-216. DOI: 10.1002/jssc.200301612
- Takahashi, M., and Kishima, T. (1973). "Decay resistance of sixty-five Southeast Asian timber specimens in accelerated laboratory tests," *Japanese Journal of Southeast Asian Studies* 10(4), 525-541. DOI: 10.20495/tak.10.4_525
- Taylor, A. M., Gartner, B. L., Morrell, J. J., and Tsunoda, K. (2006). "Effects of heartwood extractive fractions of *Thuja plicata* and *Chamaecyparis nootkatensis* on wood degradation by termites or fungi," *J. Wood Sci.* 52(2), 147-153. DOI: 10.1007/s10086-005-0743-6
- Valette, N., Perrot, T., Sormani, R., Gelhaye, E., and Morel-Rouhier, M. (2017). "Antifungal activities of wood extractives," *Fungal Biol. Rev.* 31(3), 113-123. DOI: 10.1016/j.fbr.2017.01.002
- Van Acker, J., Stevens, M., Carey, J., Sierra-Alvarez, R., Militz, H., Le Bayon, I., Kleist, G., and Peek, R. D. (2003). "Biological durability of wood in relation to end-use," *Holz Roh- Werkst.* 61(1), 35-45. DOI: 10.1007/s00107-002-0351-8
- Vidholdová, Z., and Reinprecht, L. (2019). "The colour of tropical woods influenced by brown rot," *Forests* 10(4), Article Number 322. DOI: 10.3390/f10040322
- Wagenführ, R. (2007). *Holzatlas*, Fachbuchverlag Leipzig, Carl Hanser Verlag, Munich, Germany.
- Wanschura, R., Windeisen, E., and Richter, K. (2014). "Analysis of extractives of tropical hardwoods and benefits for the surface treatment," in: *Eco-efficient Resource Wood with Special Focus on Hardwoods, Proceedings of IAWS Plenary Meeting*, R. Németh, A. Teischinger, and U. Schmitt (eds.), Sopron, Hungary, and Vienna, Austria, pp. 71-72.
- Wong, A. H. H., Kim, Y. S., Singh, A. P., and Ling, W. C. (2005). "Natural durability of tropical wood species with emphasis on Malaysian hardwoods — Variations and prospects," in: *Annual Meeting of the International Research Group on Wood Protection*, Document No. IRG/WP 05-10568, Bangalore, India, pp. 0-33.
- Yalcin, M., Tascioglu, C., Plarre, R., Akcay, C., and Busweiler, S. (2018). "Investigation of natural durability of some native and exotic wood species against *Hylotrupes bajulus* (Cerambycidae) and *Anobium punctatum* (Anobiidae)," *Kastamonu University Journal of Forestry Faculty* 18(1), 83-91. DOI: 10.17475/kastorman.311971
- Yamamoto, K., and Hong, L. T. (1989). "Location of extractives and decay resistance in some Malaysian hardwood species," *J. Trop. For Sci.* 2(1), 61-70.

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