

Dyeing Process and Mechanism of Eucalyptus Veneer with *Pterocarpus macrocarpus* Kurz Heartwood Pigment as Natural dye

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To make full use of the processing residues of *Pterocarpus macrocarpus* Kurz and reduce the environmental pollution caused by synthetic dyes, natural dye was extracted from *Dalbergia bariensis* Pierre heartwood. The purpose of the work was to prepare natural dyes of *Pterocarpus macrocarpus* and identify the key color-producing components to better explore the mechanisms of combination between dyes and eucalyptus veneers. The main components of *Pterocarpus macrocarpus* heartwood were analyzed by ultra-high performance liquid chromatography with quadrupole-electrostatic field Orbitrap high resolution-mass spectrometry (UPLC-Q-EXACTIVE Orbitrap-MS). The best dyeing process and color fastness were measured. Research technology combining Fourier transform infrared spectroscopy (FTIR) and field emission scanning electron microscopy (FESEM) was used to explore the binding mechanism between eucalyptus veneers and dyestuffs. The UPLC-Q-EXACTIVE Orbitrap-MS results showed 16 flavonoids. The optimal dyeing process parameters of eucalyptus veneer were a 90 °C dyeing temperature, 12 h dyeing time, 4 wt% pigment, and 2 wt% NaCl. The FTIR and FESM results revealed that the dyeing was mainly achieved by physical adsorption and intermolecular hydrogen bonding.

Keywords: *Pterocarpus macrocarpus* Kurz; Wood dyeing; Natural dyes; Imitation mahogany; *Eucalyptus veneer*

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INTRODUCTION

Rosewood is favored by most customers because of its high density, hardness, and unique wood style (Cao *et al.* 2003). With the improvement of people's living standards, the demand for rosewood products has increased. To meet the market demand, a substitute for rosewood is needed. In the context of the rapid development of modern forestry, eucalyptus has been widely planted because of its fast growth and high wood quality (Liu and Li 2010). However, the single color of eucalyptus greatly limits its value. Artificial dyeing of eucalyptus can improve its surface color and form more vivid patterns. Therefore, how to change the color of eucalyptus to increase its value has gradually attracted people's attention.

In the current research, most of the dyes used in wood dyeing are synthetic ones, which are relatively polluting during processing and pose a carcinogenic risk to the human body. The synthetic dyes are mostly azo dyes, and they can decompose with the formation of aromatic amine compounds that have carcinogenic effects on animals under reducing

conditions (Ye *et al.* 2018). It has been found that humans (or animals) are stained with dyes after a close contact with azo dyestuffs for a long time. Especially when the dyeing fastness is poor, the dye molecules can migrate to human (or animal) skin and enter the body surface. Once they meet azo reductase secreted in the body, they are reduced to carcinogenic aromatic amine molecules. Among them, the aromatic amines of small molecules can penetrate the membrane of human (or animal) somatic cells and further metabolize to produce nitrogen positive ions in the human (or animal) body. Nitrogen positive ions attack the nuclear DNA of the cell as a strong electrophile and render normal cells cancerous (Puvanewari *et al.* 2006). Some researchers have investigated the dyeing mechanism of acid scarlet 3R dyes on Chinese fir but ignored that acid scarlet 3R is an azo dyestuff. It is a highly conjugated molecule containing a benzene ring, which has a greater impact on environmental water (Wang *et al.* 2018). This type of water (Class V) is mainly used for agricultural watering and general landscape watering. In recent years, the European Standards Committee has published the latest test standard for banned azo dyes as EN 14362-1 (2012), which proposed 30 mg/kg as the determination limit for banned azo colorants, and required that it should be "prohibited" rather than "restricted" (Ahlström *et al.* 2005). Therefore, in the process of synthesis and use, synthetic dyes will flow to animals and plants along the food chain, causing a series of problems in ecology and the environment during synthesis and use, forcing researchers to pay attention again and look for new dyestuffs.

The development and utilization of natural dyes have begun to receive attention again with the introduction of the concept of environmentally friendly dyes (Umbreen *et al.* 2008). Natural dyes contain natural pigments that are neither carcinogenic nor harmful to the environment. Their colors are soft, soothing, warm, and attractive. Natural dyes are earth-friendly and contribute to maintaining ecological balance (Moiz *et al.* 2010). Metal salts are able to combine with dyes to form dye aggregates, which make the fabric appear dark. Natural dyes made from *Pterocarpus soyauxii* was used to dye bamboo fiber fabrics, and the good dyeing effect and color fastness were obtained (Saha Tchinda *et al.* 2014). The color difference value of cotton fabric dyed with padouk and movingui natural dyes extract was found to be better than that of bamboo fiber and wood viscose fiber, which may be easier to combine with dye molecules due to lower crystallinity. Hematoxylin-dyed silk has certain antibacterial properties (Yang *et al.* 2018). The color difference value of wool fabric dyed with natural dyes from green tea was maintained at 10.50, and its color fastness was also good (Moiz *et al.* 2010). Some studies have shown that natural dyes have a certain relationship with the types of compounds (Zhang *et al.* 2018). Although some research on natural dyes has been carried out, little is known about the chromogenic components of natural dyes and how they bind to wood surfaces.

Pterocarpus macrocarpus Kurz is mainly produced in Myanmar, Laos, and Thailand. In Pingxiang City, Guangxi Province, *P. macrocarpus* is used as wood raw material to manufacture mahogany furniture. Nevertheless, a large quantity of shavings from this tree are produced during the processing of wood. Most of it is treated as waste, resulting in a large loss (Li *et al.* 2016). *P. macrocarpus* contains a large number of inclusions, including phenolic compounds, terpenoids, and flavonoid pigments, so it is prepared as a natural dye for dyeing eucalyptus veneers.

The purpose of this study was to identify the key color-producing components in the *P. macrocarpus* dye to better explore the mechanisms of combination between dyes and eucalyptus veneers. This method not only addresses the environmental problems caused by incineration, but also enables its effective use. This research provides a

theoretical basis for developing new natural dyes and increasing the value of eucalyptus in the future.

EXPERIMENTAL

Materials

Ten-year-old fast-growing *Eucalyptus grandis x urophylla* was cut in Guangxi in the southwest of China. The sapwood was rotary cut and air-dried to make its moisture content (MC) less than 85%. The crack-free, knotless wood veneers with smooth fibers and uniform color distribution were selected and prepared with dimensions of $30 \times 2.0 \times 30 \text{ mm}^3$ (Tangential \times Radial \times Longitudinal).

Preparation of Natural Dyes

In 2018, *Pterocarpus macrocarpus* was harvested by researchers in Wacheng, Myanmar. The heartwood was processed into shavings. The shavings were washed with distilled water, and they were also air-dried to a MC less than 85%. Afterwards, they were ground with a grinder. Wood powder of approximately 80 to 100 mesh was used as raw material and it was extracted using a Soxhlet apparatus with anhydrous ethanol. The ratio of wood powder mass to solvent mass is based on the preliminary experimental results of this research group. Related information and references are as follows (Yang *et al.* 2019). The concentration of the extraction solution was the same as the weight ratio of wood powder in an organic solvent (0.05 g/mL), and the extraction temperature was the boiling point of anhydrous ethanol (79 °C). 12.5 grams of dye can be extracted per 100 grams of *P. macrocarpus* shavings. The solution was extracted completely when the liquid in the extraction tube was almost colorless. Subsequently, the extract was evaporated under the reduced pressure to obtain a concentrated solution, which was then freeze-dried under vacuum to obtain a dye. Finally, the dye was stored in a Petri dish in the refrigerator until it was used.

Dyeing of Wood Veneers

Because the volatility of absolute ethanol was reduced during the dyeing process, a specific condensation dyeing device (round bottom flask with condensing tube) from Leigu Co., Ltd. (Shanghai, China) was selected (Zhu *et al.* 2018). The dyeing process was completed by water bath heating. The factors, such as staining time, temperature, mass fraction of NaCl solution, and pigment, were investigated (Minitab Software, Minitab, LLC., Minitab, PA, USA). The factors and levels refer to previous experimental results (Sun *et al.* 2012). The factors and levels of the experimental setup are listed in Table 1.

Three pieces of eucalyptus veneers were vertically inserted into the groove of the dyeing apparatus and some appropriate space was left to ensure the evenness during the whole dyeing process. After being dyed, the eucalyptus veneer is placed in the shade for 48 h and dried naturally. The natural dyeing of eucalyptus veneers was ensured without adding mordant in the dyeing solution. The color difference of eucalyptus veneers before and after the dyeing process was tested.

Table 1. Factors and Levels of Experimental Setup

Level	Temp (°C)	Dyeing Time (h)	Mass Fraction of Pigment (%)	Mass Fraction of NaCl (%)
1	60	6	1.0	1.0
2	70	8	2.0	1.5
3	80	10	3.0	2.0
4	90	12	4.0	2.5

Color Values Measurement

The color change values of the samples dyed with dye solutions before and after dyeing were measured using colorimeter (ADCI series automatic colorimeter; Chentaik Co., Ltd., Beijing, China) under D65 illuminant and 10° standard observer at three pieces of eucalyptus veneers, and the average data are reported. The eucalyptus veneers was recorded using the CIE- L^* , a^* , b^* uniform color space (CIE-Lab) within a three-dimensional color space, where L^* indicates lightness, a^* indicates hue on a green (–) to red (+) axis, and b^* indicates hue on a blue (–) to yellow (+) axis (Xu *et al.* 2001). The values of each chromaticity parameter were collected, and the total color difference (ΔE), which denotes the surface color change, was calculated from Eqs. 1 to 4,

$$\Delta L^* = L_a^* - L_b^* \quad (1)$$

$$\Delta a^* = a_a^* - a_b^* \quad (2)$$

$$\Delta b^* = b_a^* - b_b^* \quad (3)$$

$$\Delta E = [(\Delta L^*)^2 + (\Delta a^*)^2 + (\Delta b^*)^2]^{1/2} \quad (4)$$

where the ΔL^* , Δa^* , and Δb^* represent the difference values of L^* , a^* , and b^* of the wood veneers before (L_b^* , a_b^* , and b_b^*) and after (L_a^* , a_a^* , and b_a^*) dyeing, respectively (Zhu *et al.* 2018).

Chemical Composition of the Extraction Analyses

The chemical composition of dyestuff was analyzed using ultra-high performance liquid chromatography with quadrupole-electrostatic field Orbitrap high resolution-mass spectrometry (UPLC-Q-EXACTIVE-MS). The column was a 50 mm × 2.1 mm, 1.7 μm (inside diameter), ACQUITY UPLCBEHC18 column (Water Technologies Co., Ltd., Milford, USA). The mobile phases A and B consisted of 0.1% formic acid and 99.9% methanol, respectively. The elution gradient (26 min) was as follows. After maintaining the level of eluent B at 5% for 2 min, it was linearly increased to 95% B within 18 min; then 95% B was maintained for 2 min. Finally, the gradient was switched to 5% B again over 4 min, and the column was re-equilibrated for 3 min. The flow rate of the mobile phase was kept at 0.3 mL/min, and the injection volume was set at 1 μL. The elution procedure was set according to the sample gradient shown in Table 2. The mass spectrometer method was performed in positive and negative mode in full scan MS mode with a mass range of 100 to 500 amu. The ion accumulation time was set to 30 ms, with an event time of 300 ms with three repetitions. The triple-quadrupole mass spectrometer was an Agilent 1290 instrument that was equipped with an ESI source using jet stream technology (Agilent Technologies Co., Ltd., Santa Clara, CA, USA). The ion source parameters were set as follows: 300 °C of sheath gas temperature, 12 L/min of sheath gas flow, 320 °C of gas temperature, 8 L/min of gas flow, 30 psi of nebulizer, 3000 V of capillary voltage, 0 V of nozzle voltage.

Table 2. Gradient Elution Conditions of Mobile Phase for Samples

Injection Time (min)	Flow Velocity (mL·min ⁻¹)	Mobile Phase A (%)	Mobile Phase B (%)
0	0.3	95	5
2.0	0.3	95	5
20.0	0.3	5	95
22.0	0.3	5	95
22.1	0.3	95	5
26.0	0.3	95	5

FTIR and FESEM Analyses

The surface chemical structure of eucalyptus veneers before and after dye was characterized using Fourier transform infrared spectroscopy (FTIR). All samples were mixed with KBr at a ratio of 1:100 and ground into ultrafine powder in the mortar. Then they were placed on the diamond crystal of the FTIR spectrometer (SENSOR II; Bruker, Karlsruhe, Germany). The spectra were collected in transmittance mode with 32 scans ranging from 400 to 4000 cm⁻¹ at a resolution of 1 cm⁻¹.

The samples were pasted on the sample table with a double-sided conductive adhesive and treated with gold spraying (Ion Sputter Coater 150; SuPro Instruments Co., Ltd., Shenzhen, China). The micromorphology of the eucalyptus veneers before and after dyeing was observed using a field emission scanning electron microscope (Hitachi s-3400n FESEM; Hitachi Ltd., Hitachi, Japan).

Color Fastness to Washing

The color difference value before and after washing of the dyed veneers was used as an index to measure the color fastness to washing (Ferguson and Taylor 2008). Each group of dyed veneers whose color had been measured was placed into a beaker with 100 mL of distilled water and heated in a constant temperature water bath at 60 °C for 2 h. Then the wood chips were removed and air-dried at room temperature. After measuring the surface color, the color difference before and after the water washing treatment was calculated.

RESULTS AND DISCUSSION

Main Chemical Composition of the Extraction Analysis

Positive and negative ion chromatogram spectra of ethanol extraction are presented in Figs. 1 and 2. It can be clearly seen that main peaks were found at retention time between 7 min and 15 min. For further clarification, relative peak areas were used to exhibit the relative content of main compositions of ethanol extraction, which can be obtained by calculating the ratio of each peak area.

The UPLC-Q-EXACTIVE Orbitrap-MS conditions were optimized to enable samples to obtain the best instrumental performance. Both positive and negative ion modes were employed to screen as many potential compounds in *P. macrocarpus* as possible, but the positive ion mode provided higher signal intensity and the ability to detect more peak signals, which may be due to the easy ionization of some compounds in the positive ion mode (Duan *et al.* 2017).

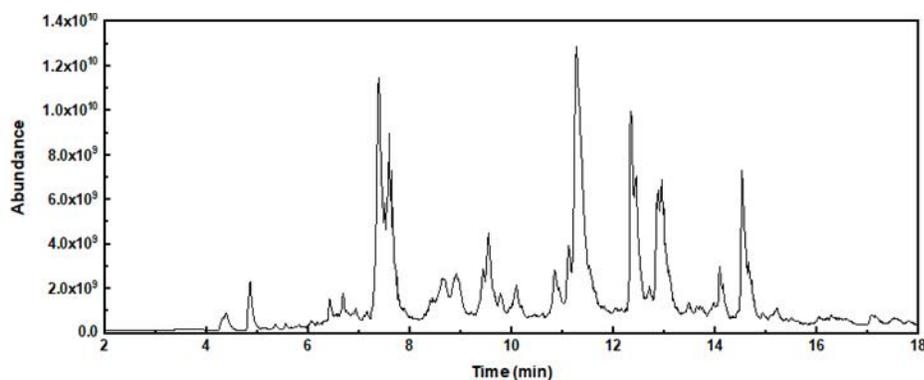


Fig. 1. Total ion chromatogram of mass spectrometer in positive ion mode

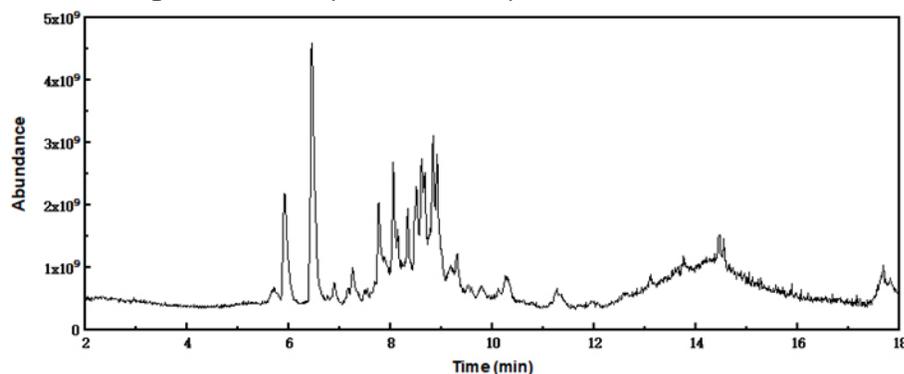


Fig. 2. Total ion chromatogram of mass spectrometer in negative ion mode

Table 3. Characterization of Chemical Constituents in Pigment from *Pterocarpus macrocarpus* Kurz Heartwood by UPLC-Q-Extractive Orbitrap-MS

No.	Retention Time (min)	Formula	Experimental Molecular Mass	Theoretical Molecular Mass	Ratios of Fragment Ions (m/z)	Component	Reference
1	4.677	C ₂₁ H ₂₀ O ₁₁	448.10027	[M+H] ⁺ 448.10056	431.096, 413.086, 396.075, 329.064, 311.054, 256.631	Orientin	Xu <i>et al.</i> (2017)
2	5.045	C ₂₁ H ₂₀ O ₁₀	432.10546	[M+H] ⁺ 432.10565	415.101, 397.091, 379.080, 313.070, 269.080,	Vitexin	Fu <i>et al.</i> (2008)
3	5.097	C ₁₆ H ₁₂ O ₆	300.06301	[M+H] ⁺ 300.06339	269.043, 241.048, 213.054,	Diosmetin	Sammani <i>et al.</i> (2017)
4	5.844	C ₁₆ H ₁₂ O ₇	316.05815	[M+H] ⁻ 316.05830	271.096, 253.085, 225.090, 179.033, 147.043,	Rhamnetin	Marzouk <i>et al.</i> (1999)

5	6.050	C ₁₅ H ₁₀ O ₅	270.05263	[M+H] ⁺ 270.05282	253.049, 183.080, 165.069, 137.059,	Genistein	Tian <i>et al.</i> (2011)
6	6.145	C ₁₆ H ₁₂ O ₆	300.06301	[M+H] ⁻ 300.06339	286.046, 269.044, 241.049, 213.054, 151.038, 137.023	Hispidulin	Ganzera <i>et al.</i> (2005)
7	6.388	C ₁₇ H ₁₄ O ₇	330.07353	[M+H] ⁺ 331.08176	303.085, 271.059, 243.064, 215.070, 153.054, 107.049	Malvidin	Heier <i>et al.</i> (2002)
8	6.400	C ₁₄ H ₁₂ O ₃	228.07846	[M+H] ⁺ 228.07864	211.075, 193.080, 135.043, 119.043,	Resveratrol	Wang <i>et al.</i> (2002)
9	6.463	C ₂₁ H ₂₀ O ₁₀	254.05769	[M+H] ⁺ 254.05791	217.185, 189.163, 147.043, 123.044, 107.049	Chrysin	Chen <i>et al.</i> (2010)
10	6.641	C ₁₅ H ₁₂ O ₄	256.07326	[M+H] ⁺ 256.07356	239.069, 211.074, 148.047,	4',7- Dihydroxyflav anone	Chae <i>et al.</i> (2016)
11	6.857	C ₁₅ H ₁₀ O ₅	270.05263	[M+H] ⁺ 270.05282	243.065, 183.080, 147.043, 119.049	Apigenin	Abdullah <i>et al.</i> (2017)
12	6.869	C ₁₆ H ₁₂ O ₅	284.06831	[M-H] ⁻ 284.06847	253.049, 225.054, 177.054, 153.054, 137.023, 123.044	Glycitein	Romani <i>et al.</i> (2010)
13	7.282	C ₁₅ H ₂₂ O	218.16675	[M-H] ⁻ 218.16707	201.163, 159.116, 135.116, 107.085,	Nootkatone	Xie <i>et al.</i> (2009)
14	8.865	C ₁₆ H ₁₂ O ₄	228.0784	[M+H] ⁺ 228.07 864	237.053, 213.089, 197.059, 118.041, 82.136	Formononetin	Xie <i>et al.</i> (2014)
15	9.236	C ₁₆ H ₁₂ O ₅	284.06832	[M+H] ⁺ 284.06 847	257.080, 175.038, 151.038, 123.044	(-)-Maackiain	Gao <i>et al.</i> (2011)
16	12.322	C ₉ H ₆ O ₃	162.03147	[M+H] ⁺ 162.03 169	135.044, 79.054	4-Hydroxy- coumarin	Jung and Park (2009)

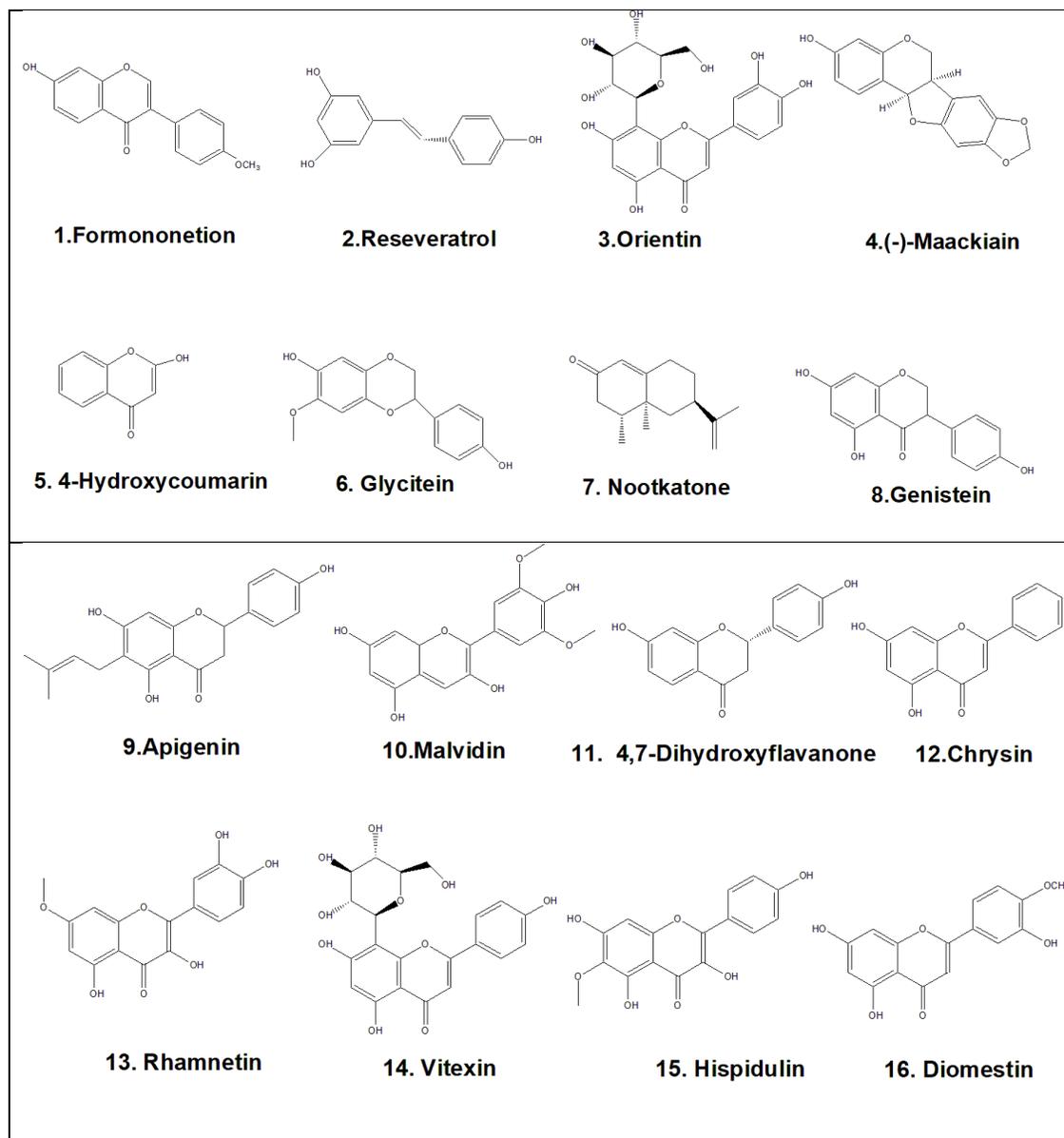


Fig. 3. The molecular structure of each compound

Among them, 16 compounds were tentatively identified by comparison with (thermo state) reference standards. After that, the compounds were analyzed and determined based on the accurate mass of quasimolecular, fragmentation ions, and MS spectra with the literature. The chemical compositions were identified by retrieval of mass spectral and Wiley libraries.

As shown in Table 3, there were 16 main chemical components. These 16 compounds were formononeti, resveratrol, orientin, (-)-maackiain, 4-hydroxycoumarin, glycitein, nootkatone, genistein, apigenin, malvidin, 4',7-dihydroxyflavanone, chrysin, rhamnetin, vitexin, hispidulin, and diosmetin.

The results presented in Figs. 3 and 4 indicated that the extracts of *P. macrocarpus* contained flavonoids and phenols compounds. Flavone has chromofunctional groups, such as conjugated carbonyl, non-conjugated carbonyl, hydroxyl, and carbon-carbon triple bonds, which probably led to the surface discoloration of wood veneers after dyeing.

However, the relationship between flavonoid and the specific chromaticity index (L , a^* , and b^*) variation needs further study.

Influence of Main Factors on Dyeing Effect

The eucalyptus veneers were dyed according to the orthogonal design table, and the color difference before and after dyeing was measured (Table 4). The order of the factors affecting the staining was: dye mass fraction > temperature > dyeing time > NaCl mass fraction. The analysis of variance showed that dye mass fraction and temperature had significant effects on the test results ($P < 0.05$).

Table 4. Comparison Results of Color Differences Before and After Dyeing

No.	Dyeing Temperature (°C)	Dyeing Time (h)	Dye Mass Fraction (%)	NaCl Mass Fraction (%)	Null Columns	Color Difference Before and After Dyeing
1	60	6	1	1.0	1	17.92
2	60	8	2	1.5	2	30.34
3	60	10	3	2.0	3	39.66
4	60	12	4	2.5	4	36.06
5	70	6	2	2.0	4	34.82
6	70	8	1	2.5	3	19.72
7	70	10	4	1.0	2	42.79
8	70	12	3	1.5	1	36.06
9	80	6	3	2.5	2	36.22
10	80	8	4	2.0	1	41.56
11	80	10	1	1.5	4	30.23
12	80	12	2	1.0	3	33.35
13	90	6	4	1.5	3	43.32
14	90	8	3	1.0	4	47.13
15	90	10	2	2.5	1	43.87
16	90	12	1	2.0	2	41.60
Average 1 (k_1)	31.00	33.07	27.37	35.30		
Average 2 (k_2)	33.35	34.69	35.59	34.99		
Average 3 (k_3)	35.34	39.14	39.77	39.41		
Average 4 (k_4)	43.98	36.77	40.93	33.97		
Range (R)	12.98	6.07	13.56	5.44		

The best process parameters were selected from a higher level of factors. They were 90 °C of dyeing temperature, 10 h of dyeing time, 4 wt% of dyeing liquid, and 1.5 wt% NaCl. In Fig. 4, the dyed samples were compared with the real picture of *P. macrocarpus*. The color difference value was calculated as 40.02. The color difference values of samples 3, 7, and 10 were slightly different from those of *P. macrocarpus*. After observation and comparison (Fig. 4), No. 10 was the closest to *P. macrocarpus* in color.

Table 5. Analyses of Variance

Factor	Sum of Squares	Freedom	F	P	Significant Figure
Temperature	384.69	3	10.25	0.044	Significance P < 0.05
Time	82.85	3	2.21	0.266	
Dye Mass Fraction (%)	452.72	3	12.06	0.035	Significance P < 0.05
NaCl Mass Fraction (%)	69.00	3	1.84	0.315	
Error	37.52	3			

S = 3.53665 R² = 96.35%

**Fig. 4.** Dyeing effect figures of test samples (samples 1 to 16) and comparison of sample 13 and *Pterocarpus macrocarpus* Kurz**Effect of Process Parameters on the Dyeing Effect of Eucalyptus Veneer***Influence of temperature on dyeing effect*

As shown in Fig. 5a, temperature had a greater effect on the dyeing effect of eucalyptus veneer than dyeing time. With increasing the temperature, the color difference of eucalyptus veneers gradually increased. The color difference value increased from 31.0 to 44.0. According to the diffusion principle, the higher the temperature, the larger the diffusion coefficient, indicating that the molecular diffusion speed becomes faster (Morita *et al.* 1986). Therefore, the dyeing solution of flavonoids and phenolic compounds diffused faster in the wood ducts and pits, and the color of the specimen became darker. When the

temperature rose up to 90 °C, the color difference of the veneers reached the highest value, that is, the darkest color. The optimum dyeing temperature was 90 °C.

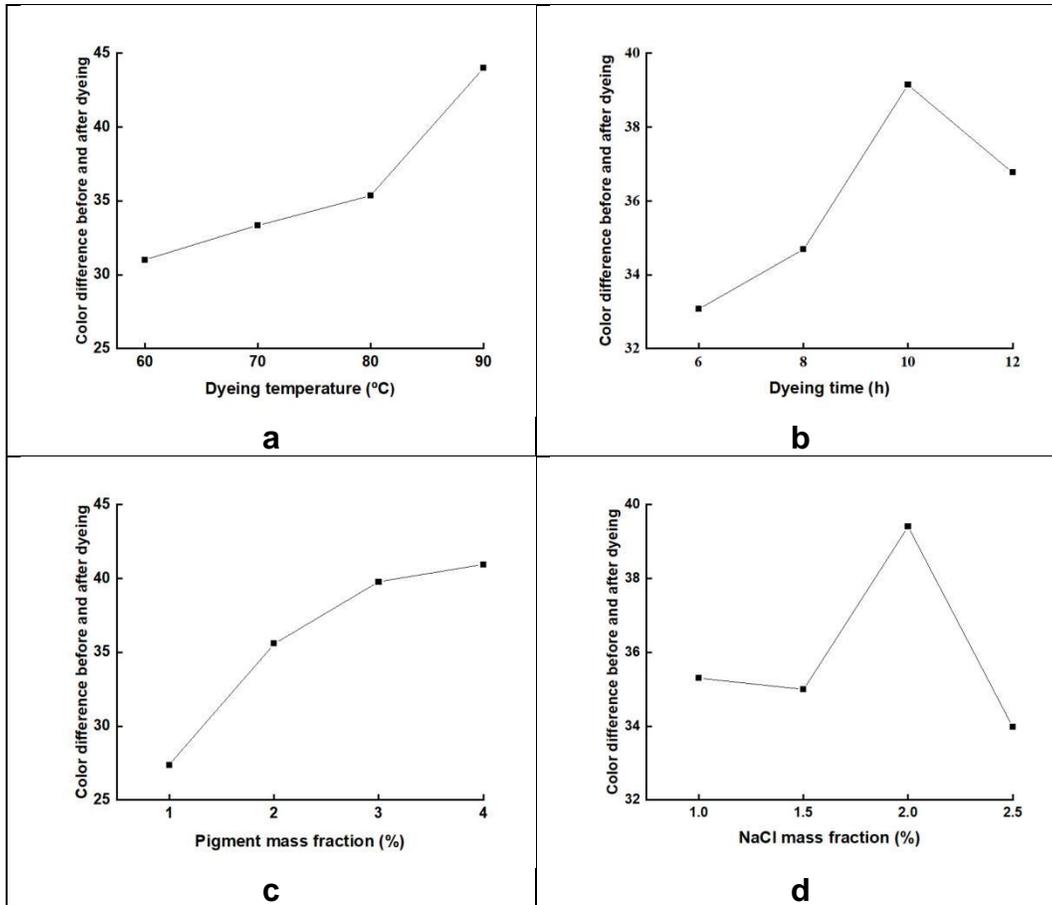


Fig. 5. Staining effect curve of different influencing factors

Effect of dyeing time on dyeing effect

As shown in Fig. 5b, the color difference value of eucalyptus veneers was almost unchanged when the dyeing time was 6 to 8 h, and it increased after 8 h. With the extension of the dyeing time, the pigment in the dye gradually penetrated the internal structure of the specimen, but the combination still needed a long time. The color difference of eucalyptus veneers reached the maximum at 10 h, which was determined as the best dyeing time.

Effect of mass fraction of dyeing solution on dyeing effect

As shown in Fig. 5c, the color difference of eucalyptus veneers increased with increased mass fraction of dyeing solution. With increased mass fraction, the contact areas between pigment of dyeing solution and the wood fibers increased, which helped the wood to absorb the dye liquor and improved the dyeing effect. The best mass fraction of dyeing solution was 4%.

Effect of mass fraction of NaCl solution on dyeing effect

As shown in Fig. 5d, the color difference of eucalyptus veneers increases with the mass fraction of NaCl added. The specimen color became deeper. In the process of wood

dyeing, NaCl can provide significantly positively charged sodium ions (Burkinshaw and Salihi 2017), which can reduce the negative charge repulsion between the surface of the wood fibers and the flavonoids and phenolic compounds in dyes. This method can effectively reduce the inhibition effect of dye on the surface of wood fibers, allowing the dye molecules to approach closer to them during treatment with aqueous solution to improve the dyeing rate. The best mass fraction of NaCl solution was 2.0%.

Color Fastness Test

Table 6 shows the color difference results of stained eucalyptus veneers before and after washing.

Table 6. Comparison Results of Color Differences Before and After Washing

No.	Dyeing Temperature (°C)	Dyeing Time (h)	Dye Mass Fraction (%)	NaCl Mass Fraction (%)	Color Difference Before and After Washing
1	60	8	1	1.0	1.04
2	60	10	2	1.5	5.50
3	60	12	3	2.0	7.99
4	60	14	4	2.5	0.59
5	70	8	2	2.0	5.20
6	70	10	1	2.5	5.62
7	70	12	4	1.0	3.52
8	70	14	3	1.5	2.88
9	80	8	3	2.5	2.34
10	80	10	4	2.0	4.52
11	80	12	1	1.5	3.53
12	80	14	2	1.0	3.66
13	90	8	4	1.5	4.16
14	90	10	3	1.0	3.47
15	90	12	2	2.5	3.24
16	90	14	1	2.0	3.11
Average 1 (k_1)	3.780	3.185	3.325	2.923	
Average 2 (k_2)	4.305	4.777	4.400	4.018	
Average 3 (k_3)	3.512	4.570	4.170	5.205	
Average 4 (k_4)	3.495	2.560	3.197	2.948	
Range (R)	0.810	2.217	1.203	2.282	

The relatively small variation in the color difference value indicated better color fastness (Zhao *et al.* 2014). In this test, the average color difference of the stained veneers before and after washing was 3.77. This small value of the color difference indicated that the color fastness of the stained veneers was relatively good. The order of the factors affecting the color fastness was as follows: NaCl mass fraction > dyeing time > dye mass fraction > temperature. The optimal combination of process parameters based on the highest color fastness to washing was: 70 °C of dyeing temperature, 8 h of dyeing time, 2 wt% of dyeing solution, and 2 wt% of NaCl solution.

The flavonoids in the *P. macrocarpus* dye and wood fibers were mainly combined by intermolecular hydrogen bonding and adsorption (Clerck *et al.* 2007). During the combination process, NaCl can reduce the negative charge on the surface of the *Eucalyptus*

veneers to reduce the repulsive force of the negative charge to the dye anions and improve the binding force (Burkinshaw and Salihu 2017). Therefore, the main factor affecting the color fastness of eucalyptus veneers was the NaCl mass fraction. Table 6 lists the various factors affecting the chromaticity value.

FTIR Analysis of the Untreated and Samples Dyed with Absolute Ethanol Extraction

Figure 6 presents the FTIR spectra of the samples before and after dyeing with ethanol extraction. The main peaks at 3330 cm^{-1} and 2928 cm^{-1} were ascribed to hydroxyl groups and C-H stretching vibration, respectively (Devashankar 2017). In addition, the peaks at 1736 cm^{-1} corresponded to the cellulose, hemicellulose, and lignin in wood (Wang *et al.* 2016).

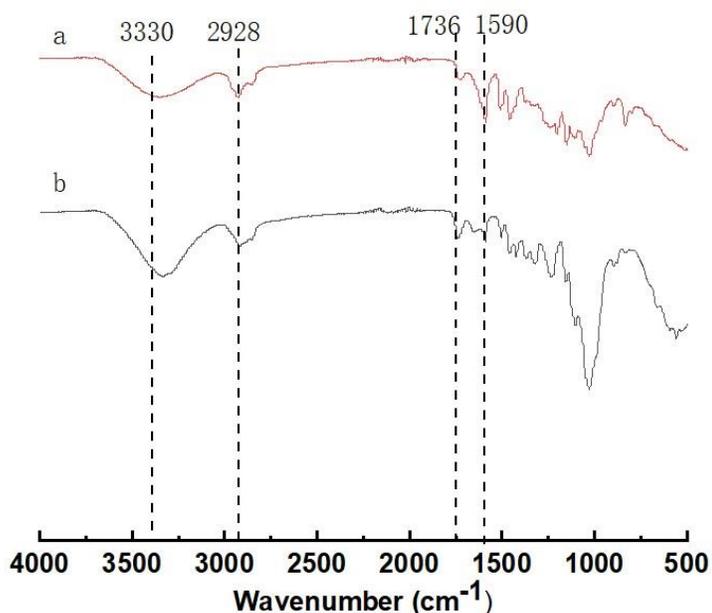


Fig. 6. FTIR spectra of untreated (b) and dyed eucalyptus wood (a)

After dyeing the *Eucalyptus* veneers, the intensity of characteristic peaks at 3330 cm^{-1} and 2928 cm^{-1} increased with the addition of the dye solution. This may be because the flavonoids in the dye solution were adsorbed on the wood surface. The compounds contain hydroxyl groups, which increased the amount of hydroxyl groups on the wood surface, resulting in an increase in the intensity of the (-OH) characteristic peak.

In contrast, the dyed sample had a carbonyl C=O stretch vibration peak located at 1590 cm^{-1} , which is attributed to the fact that flavonoids in the dye are adsorbed to the wood surface by intermolecular hydrogen bonding during the preparation of the dyed sample (Huang *et al.* 2014).

It can be seen that the intensity of the peaks located between 1721 cm^{-1} and 1615 cm^{-1} became stronger, indicating that the flavonoids were well adsorbed on the wood surface. These results demonstrated that only physical adsorption occurred between the natural dyestuff and wood tissues (Wang *et al.* 2017).

FESEM Analysis

In the transverse section of the untreated sample, circular tube holes were observed to be aligned neatly with axial parenchyma cells at magnification (Fig. 7A). The surface of wood fiber and the vessel wall of the untreated sample were smooth (Fig. 7B); the pits on the vessel wall (Fig. 7C) and xylem ray cells (Fig. 7D) were clearly visible. After the dyeing process, as shown in Fig. 7E, it was observed that the dyeing solution partially covered the duct in the cross-section of the wood, and the axial thin-walled tissue was completely covered with good flatness (Fig. 7F). In the tangential section of the dyed sample, both the wood ray (Fig. 7G) and the vessel wall (Fig. 7H) were covered by the dyestuff. Moreover, some pits were even completely covered (Fig. 7H). The results indicated that the flavonoids and phenolic compounds in the dyes formed physical adsorption with wood fibers, which can be stabilized on the wood surface (Peters *et al.* 2008; Guimaraes *et al.* 2011).

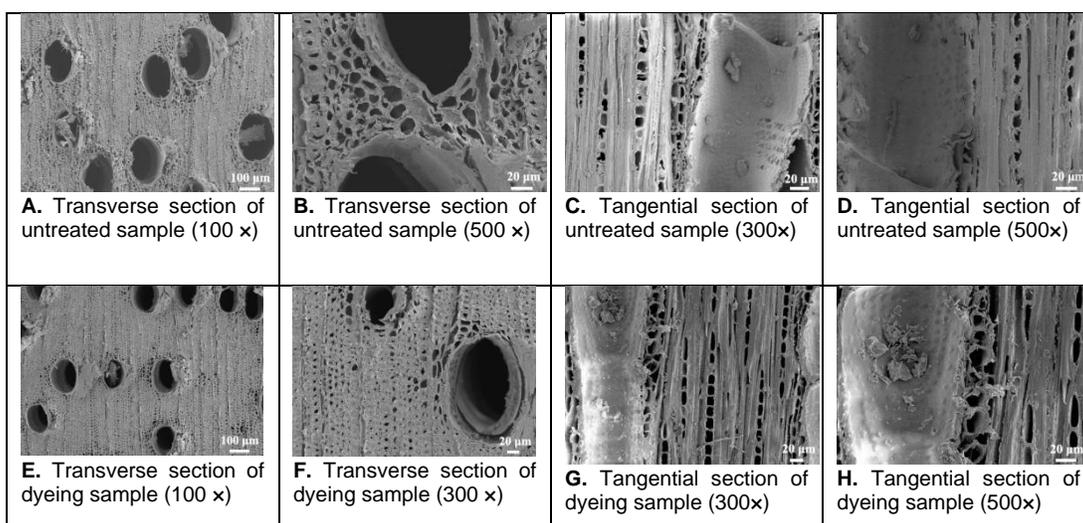


Fig. 7. FESEM micrographs of untreated and dyed *eucalyptus* wood

After the dyeing mechanism is determined, the next study can explore which components of the dye are more likely to bind to the wood surface. By changing the functional groups on the wood surface, the dye can be more firmly bound to the wood surface. In addition, it can show different colors because the dye contains different chromophoric groups and auxochromic groups. When the composition of the heartwood dyestuff from *Pterocarpus macrocarpus* Kurz is ascertained, these functional groups can be modified to change the color. In this way, the dye can change from a single color to multiple colors, thereby exploiting the color system of natural products. It can lay a theoretical foundation for improving the color fastness of eucalyptus stained wood and developing more colors.

CONCLUSIONS

1. The UPLC-Q-EXACTIVE-MS method was used to isolate and identify 16 kinds of flavonoids and phenolic compounds from the heartwood pigments of *Pterocarpus macrocarpus* Kurz.

2. The pigment was extracted from the heartwood of *Pterocarpus macrocarpus* Kurz to make a natural dye, and then the eucalyptus veneers were dyed. The order of factors influencing the dyeing effect was as follows: temperature > mass fraction of dyeing solution > dyeing time > mass fraction of NaCl solution. Through the analysis of variance of orthogonal test, the optimal dyeing process parameters were obtained as follows: 90 °C of temperature, 4 wt% of dyeing solution, 2 wt% NaCl solution, and 12 h of dyeing time. In addition, the color fastness of washed samples was measured, and the order of factors was as follows: mass fraction of NaCl > dyeing time > mass fraction of dyeing solution > temperature. The main factor affecting the color fastness of the dyed sample was temperature. The optimal dyeing process for washing with water was that the temperature was 90 °C, the dyeing time was 10 h, the mass fraction of pigment was 4%, and the mass fraction of NaCl solution was 2.0%.
3. The dyeing mechanisms were obtained by detecting the pigment composition of the heartwood of *Pterocarpus macrocarpus* Kurz combined with infrared and electron microscopy analysis. The natural dye was stably bound to the tracheid, axial tracheid wall, and wood-ray parenchyma cells, as well as the axial thin-walled tissue and chordal section of wood through physical adsorption and intermolecular hydrogen bonding.

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