# Vegetative Growth, Yield, and Fruit Quality of Guava (*Psidium guajava* L.) cv. 'Maamoura' as Affected by Some Biostimulants

Walid F. A. Mosa,<sup>a,\*</sup> Lidia Sas-Paszt,<sup>b</sup> Krzysztof Górnik,<sup>b</sup> Hayssam M. Ali,<sup>c,d</sup> and Mohamed Z. M. Salem <sup>e,\*</sup>

The present study was performed during two successive seasons 2019 and 2020 to investigate the effect of the soil application of fulvic acid (FA). seaweed extract (SE), and their different combinations on vegetative growth, yield, and fruit guality of six-years-old guava (Psidium guajava L.) cv. 'Maamoura'. The trees were planted  $4 \times 4 \text{ m}^2$  apart in clay soil under a flood irrigation system. They were treated three times starting from mid-March with one-month intervals with the following treatments: Control (water only), 3 and 4 g/L FA, 3 and 4 g/L SE, and their different combinations; 3 g/L FA + 3 g/L SE, 3 g/L FA + 4 g/L SE, 4 g/L FA + 3 g/L SE, and 4 g/L FA + 4 g/L SE. The results clearly showed that the application of FA or SE solely or in combinations increased shoot length and diameter, as well as leaf chlorophyll compared with the control. The treatments also increased fruit set percentage, fruit yield, and fruit physical and chemical characteristics such as fruit weight, size, TSS%, total reduced and non-reduced sugars, as well as leaf mineral content, while they decreased the fruit acidity compared with the control in the two seasons.

Keywords: Guava; Seaweed extract; Fulvic acid; Yield; Fruit quality; Biostimulant

Contact information: a: Plant Production Department (Horticulture- Pomology), Faculty of Agriculture, Saba Basha, Alexandria University, Alexandria, Egypt; b: Research Institute of Horticulture, Skierniewice, Poland; c: Botany and Microbiology Department, College of Science, King Saud University, P.O. Box 2455, Riyadh 11451, Saudi Arabia; d: Timber Trees Research Department, Sabahia Horticulture Research Station, Horticulture Research Institute, Agriculture Research Center, Alexandria 21526, Egypt; e: Forestry and Wood Technology Department, Faculty of Agriculture (El-Shatby), Alexandria University, Alexandria 21545, Egypt; \*Corresponding Authors: walidbreeder@yahoo.com; zidan\_forest@yahoo.com

## INTRODUCTION

Guava (*Psidium guajava* L.) belongs to the family of Myrtaceae, which is an important subtropical fruit crop. This fruit is rich in ascorbic acid and pectin and has a moderate content of calcium, phosphorus, and other vitamins (Lall *et al.* 2017).

The application of chemical fertilizers is important in increasing the yield of crops and improving the food security effectively (Ni *et al.* 2011; Duan *et al.* 2016; Suhag 2016). However, the excessive usage and the insistence on the use of chemical fertilizers has created some environmental problems, such as soil deterioration, water pollution, destruction of the soil ecology, and reduced fertility of the soil; thus it causes harmful influences on human health (Liu *et al.* 2013; Stuart *et al.* 2014; Youssef and Eissa 2014; Sierra *et al.* 2015; Smith, and Siciliano 2015; Suhag 2016; Uphoff and Dazzo 2016). So, the implementation of the organic substances instead of chemical fertilizers is necessary to raise and maintain soil fertility and health as well as food quality (Suhag 2016; Hui *et al.* 2017; Ning *et al.* 2017; Wanga *et al.* 2018).

Fulvic acid (FA) plays a crucial role in promoting the photosynthesis process. It

minimizes the opening of stomata and the transpiration rate, thus minimizing the loss of water under well-watered and also drought conditions (Lu and Beamish 2001; Li et al. 2005; Anjum et al. 2011; Huang et al. 2020). Besides, the use of FA promotes the growth of roots (Canellas et al. 2002) thereby increasing the uptake of nutrients (Razavi and BahramParvar 2007; Yang et al. 2013; Priya et al. 2014; El-Helaly 2018; Justi et al. 2019; Wang et al. 2019). It was found by Zimmerli et al. (2008) that FA has the ability to remain in the soil under high salt concentration as well as a large range from pH. Wassel et al. (2014) stated that FA has the ability to promote the vegetative growth in plants because it can raise the plants' hormones, such as indole acetic acid, gibberellic acid, and cytokines, as well as antioxidants and vitamins. El-Kenway (2017) performed a study during 2015 and 2016 seasons on 'Thompson seedless' grapevines to investigate the influence of the spray of FA at 500 ppm three times, at the beginning of the growth, after berry with one week set, and at the start of the ripening. The obtained results showed that the application of FA improved the length of shoots, leaf area, and leaf content from total chlorophyll, as well as minerals from nitrogen, phosphorous, and potassium. Moreover, it also raised the obtained yield and fruit physical and chemical characteristics in terms of weight of clusters and berry, berry cohesion, soluble solids percentage, while it reduced the percentages of juice acidity, weight loss of clusters, deterioration, and breaking down of berries compared with control in both experimental seasons. In addition, it was reported that FA enhanced the chlorophyll leaf content, roots dry matter, carbohydrates, and carotenoids (El-Helaly 2018).

Seaweed extract (SE) has a good effect on the time of flowering because of balanced carbohydrate and nitrogen content (Neumann and Zur Nieden 2001) and it contains a large variation of plant growth regulators such as auxins and cytokinin (Zhang and Ervin 2004), as well as organic compounds and polysaccharides (Sivasankari et al. 2006; Rioux et al. 2009). The application of SE to the soil increased the organic carbon, and the uptake of nitrogen, phosphorus, potassium calcium, sulfur, magnesium, zinc, manganese, and iron (Mancuso et al. 2006; Rathore et al. 2009; Zodape et al. 2011) and water content (Kocira et al. 2018). It was found that the foliar spraying of seaweed extracts (SE) was beneficial for four apple cultivars, 'Gala Must', 'Golden Delicious', 'Jonagold Decosta', and 'Elstar'. The spraying was done at the end of bloom, and subsequently four times at intervals of 4 weeks, the last time being about 4 weeks before harvest. Both preparations encouraged shoot and leaf growth, improving the quality of flower formation, extending the time of blooming, as well as raising the percentage of fruit set percentage and the fruit size (Basak 2008). SE could be applied in low quantities as soil or foliar spraying, and it can improve the resistance to insect and pathogen attack and abiotic stress such as drought, frost, salinity, and high or low temperature (Khan et al. 2009; Craigie 2011; Battacharyya et al. 2015; Elansary et al. 2016). Because of the high content of SE from nutrients, hormones, amino acids, vitamins, and antioxidants, it can be defined as a stimulant, which can increase the cell division in plants (Prasad et al. 2010; Colavita et al. 2011). de Sousa et al. (2019) reported that spraying SE on 'Gala' apple cultivars at 0.1, 0.2, 0.3, 0.4, and 0.6% increased the percentage of fruit set, number, weight, and length of fruits compared with control, and the prior treatment was 0.3%. The literature suggests that SE plays an important role in raising the tolerance of stresses, the absorption of minerals, growth, and yield. In addition, it has been shown to reduce seed dormancy and enhance root systems and flowering (Ali et al. 2019).

Therefore, this study was conducted to demonstrate the possibility of depending on FA and SE as natural sources for the nutrition of fruit trees and safe alternatives to alleviate

the undesirable effects of chemical fertilizers in increasing water, soil, and environment pollution.

## EXPERIMENTAL

## Materials

This study was conducted at a private orchard located at Abou El Matamir region, Beheira, Governorate, Egypt over two successive seasons 2019 and 2020. The study investigated the effect of the soil application of FA (X-HUMATE Fulvic Acid - 100% Water Soluble Powder- Humate (Tianjin) International Ltd., Tianjin, China), SE (seaweed extract composition: Alginic acid 15-18%, Organic matter 45-55%, Potassium K2O 16%, N 2.5-3%, P2O5 4.5-5% and Water solubility 99.1% - Hebei Hontai Biotech Co., Ltd., Shijiazhuang, Hebei, China), and their combinations on vegetative growth, yield, and fruit quality of six-year-old guava (*Psidium guajava* L.) cv. 'Maamoura'.

The trees were about 200 cm in height and were planted 4 m × 4 m apart in clay soil under a flood irrigation system. The physicochemical characteristics of the experimental soil according to Sparks *et al.* (2016) are presented in Table 1. A total of 80 trees, similar in vigor and size, were randomly chosen, arranged in a randomized complete block design (RCBD), and subjected to the same applied agricultural practices in the field during the two seasons. The following treatments of a Control (water only), FA at 3 and 4 g/L, and SE at 3 and 4 g/L, as well as their combinations – 3 g/L FA + 3 g/L SE, 3 g/L FA + 4 g/L SE, 4 g/L FA + 3 g/L SE, and 4 g/L FA + 4 g/L SE – were applied to the trees three times by solubilizing SE or FA. This was done in separate containers and by adding the material to the trees by hand. The study began in mid-March with application intervals of one month. In total, 2 L of solution for each tree/replicate were employed.

Depth (cm)	Texture	рН	Electrical Conductivity (EC) (ds/cm)	N (%)	Р (%)	K (%)	Fe (mg/L)	Zn (mg/L)	Mn (mg/L)
0 to 60	Clay	7.6	0.2	24.50	26.82	34.25	0.35	0.13	0.02

Table 1. Physicochemical Properties for the Soil of the Experiment

## Vegetative growth parameters

Starting from the vegetative season, four shoots from each side of each replicate/tree were selected and labeled, whereas, at the end of each growing season, the shoot length and diameter were measured in centimeters. The average leaf area (cm<sup>2</sup>) was determined using the following equation as reported by Demirsoy (2009),

$$LA = 0.70 (L x W) - 1.06 \tag{1}$$

where LA is a leaf area ( $cm^2$ ), *L* is the maximum length of leaf (cm), and *W* is the maximum width of the leaf (cm).

Total chlorophyll ( $\mu$  Mol/m<sup>2</sup>) in the fresh leaves was determined as SPAD units by using a Minolta chlorophyll meter (SPAD - 502; Konica Minolta, Osaka, Japan).

## Fruit set percentage, fruit yield, and fruit quality

From each side of each tree/replicate, four branches were chosen and labeled

carefully in March 2019 and 2020. The number of flowers on each branch was calculated, and then the percentage of fruit set was estimated according to Eq. 2:

Fruit set (%) = 
$$\frac{Number of set fruitlets}{Number of total flowers} \times 100$$
 (2)

The fruit yield was obtained in October, which is the typical time for harvesting. The yield of each treatment was estimated as fruit weight in kg per tree and ton per hectare.

The fruit quality was evaluated by sampling 10 fruits per tree/replicate. A total of 80 fruits for each treatment were picked randomly at harvest time in both seasons and then transported to the laboratory of Plant Production Department, Faculty of Agriculture, Saba Basha, Alexandria University, Alexandria, Egypt to determine the fruits' physical and chemical characteristics.

### **Characterization Methods**

Physical and chemical characteristics of fruits

Fruit weight (g) was estimated by calculating the average weight of 10 fruits from each tree/replicate. Average fruit length, fruit diameter (D), were measured by using a Digital Vernier Caliper (Suzhou Sunrix Precision Tools Co., Ltd., Suzhou, Jiangsu, China).

The fruit volume (cm<sup>3</sup>) was calculated by dipping the fruit in water and weighing the removed water. The pulp weight (g), seed weight (g), juice weight (g), and fruit firmness (Lb/ inch<sup>2</sup>) were estimated using a Magness and Taylor pressure tester with a <sup>7</sup>/<sub>18</sub>-inch plunger using a Magness-Taylor pressure tester (mod. FT 02 (0-2 Lb., Via Reale, 63 - 48011 Alfonsine, Italy). Total soluble solids (TSS) were measured using a hand refractometer (ATAGO CO., LTD., Tokyo, Japan), from the fresh-cut guava fruit. The result was expressed as a percentage (%). The total and reducing sugars were estimated calorimetrically using the Nelson arsenate–molybdate colorimetric method (Nielsen 2010). Non-reducing sugars were accounted as the value between total sugars and reducing sugars. The titratable acidity (%) in fruit juice was found using an AOAC method (AOAC 2005) where it was expressed as the amount of citric acid in g/100 mg fruit juice. The TSS/acid ratio was calculated by dividing the value of TSS over the value of titratable acidity. The ascorbic acid content of the juice (Vitamin C mg/100 mg juice) was estimated by titration with 2,6 dichloro phenol-indo-phenol (Nielsen 2017) and calculated as mg/100 mL of juice.

#### Leaves chemical composition

After the harvest time in November, samples of 30 leaves taken from the middle of vegetative shoots (Arrobas *et al.* 2018) were randomly selected from each replicate to determine their chemical composition of nitrogen (N), phosphorus (P), and potassium (K). The leaf samples were washed with tap water and then with distilled water, and dried at 70 °C until a constant weight was obtained; finally, the dried leaf samples were ground and acid digested using H<sub>2</sub>SO<sub>4</sub> and H<sub>2</sub>O<sub>2</sub> until the digested solution became clear. The digested solution was used for the determination of nitrogen using the micro-Kjeldahl method (Wang *et al.* 2016), phosphorus by vanadomolybdate method (Weiwei *et al.* 2017), and potassium using a flame photometer (SKZ International Co., Ltd., Jinan Shandong, China) (Arrobas *et al.* 2018).

## Statistical analysis

The results were statistically analysed using a one-way analysis of variance (ANOVA) according to Ott and Longnecker (2015), and the least significant difference

(LSD) at 0.05% was utilized for comparing between the means of treatments and measured with CoHort Software (Pacific Grove, CA, USA).

## **RESULTS AND DISCUSSION**

### Vegetative Growth Parameters

Data in Table 2 show that shoot length, shoot diameter, leaf area, and leaf total chlorophyll content were significantly increased by the soil application of SE or FA individually or in combination compared to the control in both two seasons. Moreover, the obtained results showed that the combination between FA and SE was more effective than the usage of each one of them individually. The highest increments were obtained by the soil application of 4 g/L FA combined with 3 g or 4 g/L SE in both experimental seasons.

These results agree with the previous findings of Piccolo and Mbagwu (1999). They reported that due to the structure of FA, which contains groups of COOH, OH, and phenolic compounds, it can improve the soil texture, fertility, and stability. Moreover, FA can increase the chemical elements uptake, particularly those that engaged in the process of photosynthesis like iron, zinc, and manganese (Mackowiak *et al.* 2001; Nardi *et al.* 2002; Eyheraguibel *et al.* 2008; Yang *et al.* 2013; Wang *et al.* 2019). Besides, FA also plays an important role in increasing length and number of root hairs in plants (Canellas *et al.* 2002; Schmidt *et al.* 2007), the rate of photosynthesis, and minimizing the opening of stomata, transpiration rate, and water loss so it can stimulate the growth of plants under drought and water lack conditions (Li *et al.* 2005; Anjum *et al.* 2011; Huang *et al.* 2020).

Treatment		Length m)		)iameter m)	Leaf Area (cm²)		Total Chlorophyll SPAD (μ Mol m <sup>-2</sup> )			
	2019	2020	2019	2020	2019	2020	2019	2020		
Control	22.73e	24.73e	2.19f	2.30g	35.44f	38.50f	41.11e	43.20f		
Control	±1.51	±0.3	±0.04	±0.04	±0.93	±1.6	±96	±0.03		
	24.7d	27.71cd	2.78e	3.16e	44.03d	46.87de	45.05d	46.52e		
FA (3 g/L)	±1.00	±1.03	±0.02	±0.01	±0.59	±0.52	±0.69	±1.02		
FA (4 g/L)	27.73c	29.56c	2.90d	3.29d	46.21c	51.96c	49.99b	51.73c		
FA (4 9/L)	±1.06	±1.16	±0.07	±0.04	±0.57	±1.48	±1.58	±0.75		
SE (3 g/L)	24.63d	27.63d	2.78e	2.77f	42.3e	45.28e	43.73d	45.41e		
3E (3 g/L)	±0.94	±1.01	±0.06	±0.08	±0.98	±1.08	±0.55	±0.25		
SE (4 g/L)	27.66c	27.94cd	2.79e	3.22de	45.32cd	48.79d	47.07c	48.81d		
3L (4 9/L)	±1.09	±1.57	±0.05	±0.04	±1.28	±1.34	±0.5	±1.33		
FA (3 g/L +	28.66bc	32.00b	3.58c	3.61c	46.61c	54.28b	50.39b	52.01c		
SE 3 g/L)	±0.96	±0.16	±0.06	±0.04	0.96	±0.98	±1.01	±0.67		
FA (3 g/L +	29.66b	32.08b	3.73b	3.76b	46.82c	56.70a	50.39b	53.26 <sup>bc</sup>		
SE 4 g/L)	±1.12	±1.01	±0.04	±0.6	±0.94	±1.10	±1.59	±1.55		
FA (4 g/L +	32.11a	33.04b	3.81ab	4.16a	48.89b	57.25a	52.89a	53.70b		
SE 3 g/L)	±0.97	±1.21	±0.04	±0.01	±104	±0.96	±0.65	±0.59		
FA (4 g/L +	33.61a	35.57a	3.82a	4.16a	53.78a	57.88a	54.29a	56.57a		
SE 4 g/L)	±0.96	±0.94	±0.02	±0.04	±1.16	±1.44	±0.97	±0.83		
LSD (0.05)	1.87	1.85	0.09	0.08	1.71	2.15	1.73	1.60		
Means not sh significance	Means not sharing the same letter(s) within each column are significantly different at 0.05 level of									

**Table 2.** Influence of Soil Application of SE and FA on Shoot Length, Shoot

 Diameter, Leaf Area, and Leaf Total Chlorophyll in Guava cv. 'Maamoura'

In the same trend, it was reported that because SE contains a high content from micro and macronutrients, such as iron, copper, sulfur, manganese, nitrogen, phosphorous, and gibberellic acid, indole acidic acid, cytokines, and amino acids. So, it could be classified as a biostimulant for the growth of the plant and has a great ability to enhance the plant cell division (Abdel-Mawgoud *et al.* 2010; Prasad *et al.* 2010; Hayyawi *et al.* 2020), as well as magnesium, which is very important for the synthesis of chlorophyll (Almaroai and Eissa 2020). Besides, it was reported that FA can increase the plant growth because its effect, which is similar to that of auxin, gerbilline, and cytokinin (Samavat and Samavat 2014). Additionally, the foliar application of FA was also more effective in improving leaf area and shoot length in apple (Taha *et al.* 2016; Khan *et al.* 2019). The application of SE enhanced leaf total chlorophyll, photosynthesis, transpiration, and the conductance of stomata (Fan *et al.* 2013; Al-Ghamdi and Elansary 2018; Rouphael *et al.* 2018; Kulkarni *et al.* 2019).

## Fruit Set Percentage and Fruit Yield

Results in Table 3 make it clear that fruit set percentage, fruit yield in kg per tree, and fruit yield in ton per hectare were significantly enhanced by the soil application of SE or FA at 3 or 4 g/L individually or in combinations. This may be due to the role of SE and FA in improving the vegetative growth parameters and leaf total chlorophyll as well as improving the availability of nutrients. The results showed also that mixing FA with SE was more effective in their effect than the individual application in the two seasons. The combination of 4 g/L FA with 4 g/L SE was the superior treatment as compared to the control and the other applied treatments in both experimental seasons. Additionally, fruit set and yield were significantly increased by the combinations of 3 g/L FA + 4 g/L SE and 4 g/L FA + 3 g/L SE, in the two seasons compared with control. These results were previously explained in several studies (Stirk et al. 2003; Craigie 2011; Gupta et al. 2011; Khan et al. 2011; Stirk and Van Staden, 2014; Battacharyya et al. 2015; Aremu et al. 2016; Patel et al. 2018; Renaut et al. 2019; Yalçın et al. 2019; Al-Juthery et al. 2020). These groups reported that, as SEs are rich in polysaccharides, auxins, gibberellins, cytokinins, indole-3-acetic acid (IAA), vitamins, oils, fats, acids, amino acids, polyphenols antioxidants, pigments, antimicrobial factors, Fe, Cu, Zn, Co, Mo, Mn, and Ni, they can improve the yield and yield components.

The foliar application of fulvic acid was more effective in improving the fruit set percentage and yield in kg/tree in apple (Basak 2008; Khan *et al.* 2019). In the same trend, it was reported previously that the application of SEs increased the flower set percentage and fruit yield in "Koroneiki" olive cultivar (Chouliaras *et al.* 2009), in pear (Calvo *et al.* 2014) and in "Fagri Kalan" mango cultivar (El-Sharony *et al.* 2015). Besides, SE enhanced the plant resistance to biotic (Machado *et al.* 2014; Ben Salah *et al.* 2018) and abiotic stress (Bradáčová *et al.* 2016; Cabo *et al.* 2019; Khompatara *et al.* 2019), stimulated the growth of crops, and increased yield (Renaut *et al.* 2019; El Boukhari *et al.* 2020). In addition, treating orange with SEs increased the maturity index and yield, while it decreased the fruit drop (Arioli *et al.* 2015; Gomathi, *et al.* 2017). Spraying SE on apple (*Malus domestica* L.) increased fruit set and fruit yield (Soppelsa *et al.* 2018; Valencia *et al.* 2018; de Sousa *et al.* 2019). The application of SEs at 0, 2, 4, and 8 gL<sup>-1</sup> on 'Albion' strawberry cultivar *via* fertigation system increased the number of flowers per plant, and the yield, compared with the control, was also increased (Al-Shatri *et al.* 2020). **Table 3.** Influence of Soil Application of SE and FA on Fruit Set Percentages, Fruit

 Yield in kg per Tree, and Fruit Yield in Ton per Hectare in Guava cv. 'Maamoura'

Treetment	Fruit	Set (%)	Fruit Yield	d (kg/Tree)	Yield (ton	)/Hectare			
Treatment	2019	2020	2019	2020	2019	2020			
Control	54.9d	58.95f	40.52f	46.19f	24.31f	27.72f			
Control	±0.54	±1.45	±2.03	±2.65	±1.22	±1.59			
	67.2bc	66.39e	61.28d	66.96de	36.77d	40.18de			
FA (3 g/L)	±1.43	±0.68	±1.32	±1.5	±0.79	±0.90			
	68.72b	68.69cd	65.31c	72.64bc	39.19c	43.59bc			
FA (4 g/L)	±1.24	±1.41	±1.04	±1.90	±0.63	±1.14			
	66.37c	65.25e	58.48e	65.4e	35.09e	39.24e			
SE (3 g/L)	±0.97	±1.59	±1.80	±1.13	±1.08	±0.67			
	67.78bc	67.49de	64.27c	70.2cd	38.56c	42.12cd			
SE (4 g/L)	±2.00	±1.02	±1.24	±3.08	±0.74	±1.85			
FA (3 g/L + SE 3	69.03b	70.65bc	71.05b	74.25b	42.63b	44.55b			
g/L)	±1.78	±0.11	±1.23	±0.86	±0.74	±0.52			
FA (3 g/L + SE 4	71.67a	71.09b	72.34b	75.93b	43.40b	45.56b			
g/L)	±1.76	±1.10	±2.11	±2.50	±1.27	±1.50			
FA (4 g/L + SE 3	72.09a	71.49b	73.52b	81.65a	44.11b	48.99a			
g/L)	±1.73	±1.82	±1.98	±3.18	±1.19	±1.90			
FA (4 g/L + SE 4	72.83a	74.33a	76.29a	83.25a	45.77a	49.95a			
g/L)	±0.85	±1.57	±0.82	±0.64	±0.49	±0.39			
LSD (0.05)	2.31	2.27	2.60	3.83	1.56	2.30			
Means not sharing the of significance	Means not sharing the same letter(s) within each column are significantly different at 0.05 level								

## **Fruit Physical Characteristics**

The data shown in Table 4 demonstrates that fruit weight, volume, length, and diameter were significantly increased by the soil application of FA or SE at 3 or 4 g/L alone or in combination with the other in the two seasons compared to the control. Moreover, the same fruit physical characteristics were increased with the combinations of FA at 3 or 4 g/L with SE at 3 or 4 rather than the usage of FA, SE alone, or control treatment in both experimental seasons. It was noticed that the effect of 4 g/L FA + 4 g SE combination was superior in comparison with 4 g/L FA + 3 g SE, 3 g/L FA + 4 g SE, and 3 g/L FA + 3 g SE combinations as compared to control in both experimental seasons. Similar results were obtained by El-Boray et al. (2015). They reported that spraying 'Superior seedless' grape cultivar with FA at 9 mL/L per vine significantly increased the average cluster weight, size, length, and width. Additionally, the foliar application of FA was more effective in improving fruit weight, fruit length, and diameter in apricot (Haggag et al. 2016). Al-Musawi (2018) stated that the spraying of sour orange with algae extracts increased fruit weight, length, width, and size, width and weight of peel, and moisture content in fruit and peel. Al-Shatri *et al.* (2020) reported that the application of SEs at 0, 2, 4, and 8  $gL^{-1}$  on 'Albion' strawberry cultivar via fertigation system increased fruit weight and volume, as compared with control. The obtained results also agree with the prior findings of Harhash et al (2021a). They reported that the foliar application of 'Flame seedless' grape trees with FA at 1000, 1500, and 2000 ppm, and SE at 2000, 3000, and 4000 ppm, before flowering, during the full bloom, and three weeks later improved weight, length, width, size, and number of clusters as well as the weight of 100 berries as compared to control. The current results are in agreement with the findings of Harhash et al. (2021b). They found that spraying pomegranate cultivar cv. 'Wonderful' at the beginning of flowering, full bloom,

and one month later three times with FA at 0.2%, 0.3%, and 0.4%, improved weight, size, length, number, and width of fruit.

Treatment	Fruit Weight (g)		Fruit Volume (cm <sup>3</sup> )		Fruit Length (cm)		Fruit Diameter (cm)	
	2019	2020	2019	2020	2019	2020	2019	2020
Control	136.96e	143.48f	149.68e	154.88d	7.17g	7.18g	4.37e	4.38f
	±1.58	±1.60	±0.15	±4.49	±0.06	±0.04	±0.12	±0.03
	155.43cd	156.82de	166.2d	171.62c	8.35e	8.46e	5.20c	5.3d
FA (3 g/L)	±1.79	±2.35	±2.59	±1.85	±0.02	±0.03	±0.07	±0.06
	161.48b	162.93c	172.23c	174.97bc	8.5cd	8.55cde	5.48b	5.56bc
FA (4 g/L)	±2.65	±0.36	±4.30	±1.51	±0.06	±0.06	±0.03	±0.01
	152.90d	155.04e	164.48d	170.47c	8.1f	8.12f	5.02d	5.07e
SE (3 g/L)	±2.18	±1.94	±0.80	±1.90	±0.02	±0.02	±0.08	±0.10
	156.33c	158.37d	166.42d	171.86c	8.49d	8.53de	5.47b	5.50c
SE (4 g/L)	±1.38	±1.80	±1.12	±4.72	±0.05	±0.06	±0.12	±0.09
FA (3 g/L + SE	162.58b	164.34bc	173.6c	177.38 <sup>ab</sup>	8.59 <sup>bc</sup>	8.62bcd	5.53b	5.58bc
3 g/L)	±0.97	±2.14	±2.94	±2.57	±0.04	±0.05	±0.08	±0.05
FA (3 g/L + SE	164.22b	165.06bc	175.85 <sup>bc</sup>	178.01ab	8.63b	8.64bc	5.51b	5.58bc
4 g/L)	±1.55	±2.12	±0.46	±0.46	±0.10	±0.01	±0.11	±0.13
FA (4 g/L + SE	164.23b	166.63ab	178.25 <sup>ab</sup>	178.55ab	8.65b	8.68b	5.54b	5.63b
3 g/L)	±0.93	±1.61	±3.04	±1.81	±0.05	±0.04	±0.02	±0.04
FA (4 g/L + SE	167.46a	168.37a	180.19a	181.83a	8.8a	8.9a	5.81a	6.14a
4 g/L)	±1.02	±2.30	±1.63	±1.83	±0.02	±0.13	±0.03	±0.02
LSD (0.05)	2.83	2.41	4.08	4.56	0.09	0.11	0.13	0.11
Means not shari significance	ng the same	e letter(s) w	ithin each	column are	significa	ntly differer	nt at 0.05	level of

**Table 4.** Influence of Soil Application of SE and FA on Fruit Weight, Volume,

 Length, and Diameter of Guava cv. 'Maamoura'

Table 5 shows that the weight of juice and pulp and fruit firmness were statistically increased by the soil application of FA or SE at 3 or 4 g/L solely or in combinations in the two seasons. The effect of mixing of FA with SE have a higher effect on the same forementioned fruit characteristics than the single application of each one of, where the highest increments were obtained from 3 g/L FA + 4 g/L SE, 4 g/L FA + 3 g/L SE, and 3 g/L FA + 3 g/L SE g/L. The impact of 4 g/L FA + 4g/L SE combination was superior as compared to the rest of the applied treatments and control in both experimental seasons. In contrast, the control treatment significantly increased the seed weight compared with the usage of FA or SE at 3 and 4 g/L or their combinations in both experimental seasons. These results are in parallel with the results obtained by Haggag et al. (2016) on apricot and Taha et al. (2016) on apple, where they reported that spraying FA enhanced the fruit firmness. Additionally, Ravi et al. (2018) stated that spraying strawberry with SE improved bunch weight, hands, and fingers of banana cv. 'Grand Naine'. Spraying grape cv. 'Flame seedless' with FA at 1000, 1500, and 2000 ppm, and SE at 2000, 3000, and 4000 ppm, before flowering, during the full bloom, and three weeks later improved juice percentage of 100 berries and fruit firmness compared with the control (Harhash et al. 2021a). Similarly, Harhash et al. (2021b) found that the foliar application of FA at 0.2%, 0.3%, and 0.4% on eight-year-old pomegranate cultivar cv. 'Wonderful' improved fruit firmness and juice weight.

**Table 5.** Influence of Soil Application of SE and FA on Weight of Juice, Pulp, and Seed and Fruit Firmness of Guava cv. 'Maamoura'

Treatment	Juice Weight (g)		Pulp Weight (g)		Seed Weight (g)		Fruit Firmness (lb/inch <sup>2</sup> )	
	2019	2020	2019	2020	2019	2020	2019	2020
Control	86.10e	86.42e	115.29e	121.79f	21.68a	21.69a	5.12f	5.15f
	±1.47	±1.59	±2.22	±2.75	±0.67	±1.19	±0.05	±0.06
$E \wedge (2 \alpha / 1)$	89.15cd	90.55d	136.16c	136.73de	19.27c	20.09bc	5.88d	6.00de
FA (3 g/L)	±1.05	±1.90	±1.66	2.08	0.13	0.30	0.02	±0.04
	91.34bc	92.29bc	144.41b	143.55c	18.68d	19.41cd	5.92d	6.05d
FA (4 g/L)	2.18	±1.60	±2.60	±1.01	±0.49	±0.86	±0.03	±0.11
	88.55de	89.43d	133.07d	134.05e	19.83b	20.99ab	5.72e	5.84e
SE (3 g/L)	±1.03	±1.00	±2.06	±1.68	0.17	0.61	±0.07	±0.10
	90.15 <sup>bcd</sup>	90.61cd	137.50c	138.50d	18.83cd	19.86c	5.92d	6.00de
SE (4 g/L)	±1.66	±0.55	±1.35	1.47	±0.07	±0.45	±0.02	±0.09
FA (3 g/L + SE 3	91.44bc	93.06b	144.73b	146.25bc	17.85e	19.38cd	6.08c	6.07d
g/L)	±1.09	±0.52	±0.58	±1.98	±0.43	±0.65	±0.02	±0.06
FA (3 g/L + SE 4	92.24ab	93.27b	145.55b	146.55b	17.25f	18.51de	6.29b	6.36c
g/L)	±1.59	±0.54	±1.34	±2.24	±0.13	±0.12	±0.05	±0.10
FA (4 g/L + SE 3	94.49a	95.05a	146.97b	147.22b	17.33f	18.31e	6.40b	6.57b
g/L)	±1.01	±0.48	±1.54	±0.89	±0.15	±0.12	±0.05	±0.15
FA (4 g/L + SE 4	94.61a	95.51a	150.13a	150.07a	17.07f	18.09e	7.37a	7.6a
g/L)	±4.03	±2.00	±0.91	±2.4	±0.09	±0.21	±0.15	±0.2
LSD (0.05)	2.56	1.72	2.95	2.79	0.50	1.05	0.11	0.20
Means not sharing the same letter(s) within each column are significantly different at 0.05 level of significance								

## **Fruit Chemical Characteristics**

According to the results in Table 6, it was noticed that the soil application of FA and SE at 3 or 4 g/L and their combinations improved TSS and vitamin C, while they decreased the juice acidity percentage in the two experimental seasons, and the influence of FA and SE increased by mixing them together. In addition, 4g/L FA + 4g/L SE combination gave a significant increment in the percentage of TSS and fruit content from vitamin C, while it reduced the juice acidity percentage comparing with the combinations of 3 g/L FA + 3 SE g/L, 3 g/L FA + 4 g/L SE, and 4 g/L FA + 3 g/L SE in both seasons. On the opposite side, control treatment raised the percentage of juice acidity comparing with the rest applied treatments in the two seasons. The optimal treatment that achieved the best results was 4 g/L FA + 4 g/L SE in the two seasons. The present obtained results are similar with those found by Suh *et al.* (2014). They reported that the application of FA on tomato raised soluble solids concentration and SSC/acidity ratio, while it minimized total acidity percent. In the same orientation, it was noticed that treating orange with SEs increased fruit content from vitamin C and TSS (Arioli et al. 2015; Gomathi et al. 2017). Moreover, the application of FA on grape cultivars such as 'Superior seedless' (El-Boray et al. 2015) with at 9 mL/L per vine, 'Thompson seedless' (El-Kenawy 2017) with 500 ppm, 'King Ruby' (Mostafa et al. 2017) with 9 mL/L/vine and 'Flame seedless' (Harhash et al. 2021a) with 2000 ppm improved TSS, TSS/acid ratio, while it reduced the percentage of titratable acidity. Spraying of FA increased total soluble solids, and vitamin C, while it reduced the percentage of fruit juice acidity comparing with control in apricot (Haggag et al. 2016) and in apple (Khan et al. 2019). The application of SEs at 0, 2, 4, 8 g.L-1 on 'Albion' strawberry cultivar via fertigation system increased TSS, TSS/TA ratio, while decreased the fruit titratability acidity comparing with control were increased (Al-Shatri *et al.* 2020). Harhash *et al.* (2021b) found that spraying eight years old pomegranate cv. Wonderful with FA at 0.2%, 0.3%, and 0.4%, at the beginning of flowering, full bloom, and one month later three times increased total soluble solids, total reduced and non-reducing sugars, and TSS/acid ratio, while it minimized the percentages of fruit acidity as compared to the control.

Treatment	TSS	S (%)	Acidi	ty (%)	Vitamin C (mg/100 mL)		
	2019	2020	2019	2020	2019	2020	
Control	8.54 h	9.55g	0.5a	0.51a	175.93e	176.91f	
Control	±0.2	±0.12	±0.05	±0.03	±1.81	±2.11	
$E \wedge (2 \alpha / 1)$	10.33f	11.18e	0.42b	0.44b	184.88d	182.82de	
FA (3 g/L)	±0.05	±0.04	±0.01	±0.01	±2.39	±2.56	
FA (4 g/L)	11.92d	12.10c	0.37c	0.38c	185.5d	185.19de	
FA (4 9/L)	±0.15	±0.14	0.006	±0.02	±1.91	±2.19	
	9.6g	10.22f	0.49a	0.50a	177.55e	181.84e	
SE (3 g/L)	0.25	±0.06	±0.04	±0.04	±2.83	±1.07	
	11.25e	11.57d	0.38c	0.39c	185.42d	185.09de	
SE (4 g/L)	±0.10	±0.18	±0.006	±0.01	±1.00	2.37	
FA (3 g/L + SE 3	12.6c	12.74b	0.36c	0.37c	187.39d	185.89d	
g/L)	±0.31	±0.08	±0.006	±0.01	±1.91	±1.14	
FA (3 g/L + SE 4	12.67c	12.99b	0.29d	0.32d	195.83c	196.58c	
g/L)	0.13	±0.19	±0.01	±0.01	±1.09	±2.15	
FA (4 g/L + SE 3	13.61b	14.12a	0.29d	0.31d	200.09b	203.13b	
g/L)	±0.36	±0.03	±0.02	±0.01	±2.94	±3.20	
FA (4 g/L + SE 4	14.06a	14.2a	0.24e	0.26e	207.42a	208.11a	
g/L)	±0.25	±0.37	±0.01	±0.006	±2.00	±2.52	
LSD (0.05)	0.36	0.28	0.04	0.04	3.37	3.35	
Means not sharing the same letter(s) within each column are significantly different at 0.05 level of significance							

**Table 6.** Influence of Soil Application of SE and FA on TSS and Acidity

 Percentages and Vitamin C in Guava cv. 'Maamoura'

From the results in Table 7, the addition of FA and SE to the soil greatly improved the fruit content from total, reduced, and non-reduced sugars compared to the control in the two seasons. This perhaps was because of their influence in raising the leaf total chlorophyll, TSS, and leaf surface area in the two seasons as well as their role in reducing the juice acidity percentages. Combining FA with SE was more effective in improving the fruit content from total, reduced and non-reduced sugars more than the individual addition of FA or SE in the two seasons. It was seen from the results that 4 g/L FA + 4 g/L SE and 4 g/L FA + 3 g/L SE statistically increased these fruit chemical characteristics. Next in order of effectiveness were 3 g/L FA + 4 g/L SE and 3 g/L FA + 3 g/L SE as compared to control. These results are in parallel with the findings of (El-Miniawy et al. 2014; Kapur et al. 2018; Mattner et al. 2018). They stated that treating strawberry with SE increased crown carbohydrate. Similarly, spraying FA improved total carbohydrates in grape cvs. 'Thompson seedless' (El-Kenawy 2017) and 'King Ruby' (Mostafa et al. 2017). Kapur et al. (2018) found that the foliar application of SEs on strawberry improved the fruit chemical content from sucrose and fructose. In the same trend, it was found by Harhash et al. (2021a) that spraying grape cv. 'Flame seedless' with FA at 1000, 1500, and 2000 ppm, and with SE at 2000, 3000, and 4000 ppm before flowering, during the full bloom, and three weeks later improved the percentages of total sugars and total soluble solids as compared to the control in the two seasons. Moreover, in another study, it was reported that treatment of pomegranate cv. 'Wonderful' with FA improved the fruit content from total, reduced and non-reduced sugars compared with the control (Harhash *et al.* 2021b).

Treatment	Total Sug	gar (%)	Reduced	Sugar (%)	Non reduced Sugar (%)		
	2019	2020	2019	2020	2019	2020	
Control	7.11h	7.19g	5.07g	5.13f	2.03e	2.06f	
Control	±0.03	±0.04	±0.06	±0.03	±0.07	±0.06	
$E \wedge (2 \alpha / 1)$	7.91f	7.97f	5.56de	5.53de	2.35d	2.45e	
FA (3 g/L)	0.07	±0.13	±0.10	±0.13	±0.03	±0.100	
$EA(4\alpha/L)$	8.57e	8.46e	5.39ef	5.43e	3.18b	3.03c	
FA (4 g/L)	±0.02	±0.09	±0.08	±0.07	±0.09	±0.09	
	7.63g	7.93f	5.25f	5.40e	2.37d	2.53de	
SE (3 g/L)	±0.02	±0.06	±0.08	±0.07	±0.06	±0.12	
	8.01f	8.09f	5.55de	5.61d	2.46d	2.48e	
SE (4 g/L)	±0.11	±0.12	±0.09	±0.08	±0.06	±0.06	
FA (3 g/L + SE 3	8.83d	8.86d	5.69d	6.22b	3.14b	2.64d	
g/L)	±0.07	±0.08	±0.11	±0.04	±0.06	±0.11	
FA (3 g/L + SE 4	8.97c	9.41c	6.26c	6.01c	2.71c	3.40b	
g/L)	±0.14	±0.15	±0.07	±0.09	±0.18	±0.06	
FA (4 g/L + SE 3	9.32b	9.68b	6.55b	6.55a	2.77c	3.13c	
g/L)	±0.07	±0.15	±0.10	±0.10	±0.07	±0.10	
FA (4 g/L + SE 4	10.25a	9.85a	6.73a	6.22b	3.53a	3.63a	
g/L)	±0.06	±0.1	±0.11	±0.10	±0.06	±0.10	
LSD (0.05)	0.14	0.16	0.17	0.15	0.16	0.14	

**Table 7.** Influence of Soil Application of SE and FA on the Percentages of the Total, Reduced, and Non-reduced Sugars in Guava cv. 'Maamoura'

Means not sharing the same letter(s) within each column are significantly different at 0.05 level of significance

#### Leaf Nitrogen (N), Phosphorous (P) and Potassium (K) Composition

Leaf mineral composition from N, P, and K macronutrients was significantly increased by the soil application of SE or FA at 3 or 4 g/L even when each was added alone or in combination compared with the control during the two seasons (Table 8). The combinations between FA and seaweed gave the most pronounced effect rather the single application. Moreover, the addition of 4 g/L FA + 4 g/L SE statistically increased NPK leaf mineral content more than 4 g/L FA + 3 g/L SE, 3 g/L FA + 4 g/L SE, and 3 g/L FA + 3 g/L SE in the two experimental seasons. Similar results were earlier obtained by Chen et al. (2004). Based on their findings, they stated that FA plays an important role in transporting the mineral nutrients directly to the sites of metabolism in the plant cells. In addition, because FA can increase significantly chelating, absorption, and distribution of elements in roots and shoots, it can encourage their growth (Bocanegra et al. 2006; Razavi, and BahramParvar 2007; Yildirim 2007; Yang et al. 2013; Canellas et al. 2015; Lotfi et al. 2015; Wang et al. 2019). Additionally, SE is rich in macro-nutrients including C, Mg, P, K, and S and micronutrients such as B, Co, F, Mn, Mo, Se, Si, Zn (Parthiban et al. 2013; Circuncisão *et al.* 2018), and it can increase the uptake of plants to soil nutrients (Renaut et al., 2019; EL Boukhari et al. 2020). Moreover, the results of the present experiment are in parallel with the findings of El-Miniawy et al. (2014), Kapur et al. (2018), and Mattner et al. 2018). They mentioned that the spray of strawberry with SE increased leaf mineral content of phosphorus and potassium. The foliar application of FA raised the leaf mineral content of nitrogen, phosphorous, potassium, zinc, manganese, and iron in comparison to the control in apricot (Haggag *et al.* 2016) and in apple (Taha *et al.* 2016; Khan *et al.* 2019). The foliar spraying of "Wonderful" pomegranate cultivar with FA at 0.2%, 0.3%, and 0.4% raised the leaf mineral content from N, P, and K in the two seasons, as compared to control (Harhash *et al.* 2021b).

Trootmont	Ν	(%)	P (	%)	K	(%)		
Treatment	2019	2020	2019	2020	2019	2020		
Control	1.99e	2.07d	0.31e	0.31d	2.49g	2.56f		
Control	±0.04	±0.02	±0.01	±0.01	±0.04	±0.03		
FA (3 g/L)	2.11d	2.20c	0.36d	0.34d	2.72f	2.81e		
FA (3 g/L)	±0.03	±0.04	±0.01	±0.01	±0.07	±0.05		
$E\Lambda (4 \alpha/L)$	2.28c	2.33b	0.39cd	0.41c	3.01e	3.05d		
FA (4 g/L)	±0.06	±0.03	±0.02	±0.02	±0.05	±0.04		
	2.06de	2.11cd	0.32e	0.34d	2.71f	2.78e		
SE (3 g/L)	±0.02	±0.04	0.01	0.006	±0.05	±0.09		
	2.23c	2.31b	0.37cd	0.4c	2.96e	3.04d		
SE (4 g/L)	±0.06	±0.04	±0.01	±0.04	±0.08	±0.08		
FA (3 g/L + SE 3	2.62b	2.72a	0.39cd	0.42c	3.47d	3.52c		
g/L)	±0.16	±0.13	0.01	±0.02	0.05	±0.03		
FA (3 g/L + SE 4	2.66b	2.72a	0.40c	0.43bc	3.88c	3.98b		
g/L)	±0.04	±0.06	0.02	±0.01	±0.03	±0.06		
FA (4 g/L + SE 3	2.76a	2.79a	0.44b	0.45b	3.99b	4.09a		
g/L)	±0.07	±0.09	0.02	±0.02	±0.05	±0.01		
FA (4 g/L + SE 4	2.85a	2.8a	0.5a	0.51a	4.14a	4.18a		
g/L)	±0.10	±0.02	±0.03	±0.03	±0.01	±0.02		
LSD (0.05)	0.10	0.09	0.03	0.03	0.10	0.09		
Means not sharing the same letter(s) within each column are significantly different at 0.05 level of significance								

 Table 8. Influence of Soil Application of SE and FA on Leaf Composition of N, P, and K in Guava cv. 'Maamoura'

# CONCLUSIONS

- 1. The soil application of fulvic acid (FA) and seaweed extract (SE), as well as their combinations, had significant effects in improving shoot length, shoot diameter, leaf area, and leaf total chlorophyll content when comparing results within two seasons.
- 2. The soil treating of the trees with FA and SE increased fruit set percentage, yield in kg per tree and in ton per hectare, as well as and fruit physical and chemical characteristics such as fruit weight, volume, firmness, and juice, TSS%, total, reduced and non-reduced sugars, and vitamin C, while they reduced that fruit acidity percentage comparing with control in both experimental seasons.
- 3. FA or SE can be used as safe and effective alternatives to the chemical fertilizers in the nutrition of fruit trees to produce safe food, keep the soil fertility, and reduce the environmental contamination.

## ACKNOWLEDGMENTS

This research was funded by the Researchers Supporting Project (RSP-2021/123) King Saud University, Riyadh, Saudi Arabia.

# **REFERENCES CITED**

- Abdel-Mawgoud, A. M., Lépine, F., and Déziel, E. (2010). "Rhamnolipids: Diversity of structures, microbial origins and roles," *Applied Microbiology and Biotechnology* 86(5), 1323-1336. DOI: 10.1007/s 00253-010-2498-2
- Ali, O., Ramsubhag, A., and Jayaraman, J. (2019). "Biostimulatory activities of *Ascophyllum nodosum* extract in tomato and sweet pepper crops in a tropical environment,"*Plos One* 14(5), e0216710. DOI: 10.1371/journal.pone.0216710
- Al-Ghamdi, A. A., and Elansary, H. O. (2018). "Synergetic effects of 5-aminolevulinic acid and Ascophyllum nodosum seaweed extracts on Asparagus phenolics and stress related genes under saline irrigation," Plant Physiology and Biochemistry 129, 273-284. DOI: 10.1016/j.plaphy.2018.06.008.
- Al-Juthery, H. W., Drebee, H. A., Al-Khafaji, B. M., and Hadi, R. F. (2020). "Plant biostimulants, seaweeds extract as a model (article review)," in: *IOP Conference Series: Earth and Environmental Science*, 553(1), 012015. DOI: 10.1088/1755-1315/553/1/012015
- Almaroai, Y. A., and Eissa, M. A. (2020). "Role of marine algae extracts in water stress resistance of onion under semiarid conditions," *Journal of Soil Science and Plant Nutrition* 20(3), 1092-1101. DOI: 10.1007/s42729-020-00195-0
- Al-Musawi, M. A. H. M. (2018). "Effect of foliar application with algae extracts on fruit quality of sour orange, *Citrus aurantium*, L.," *Journal of Environmental Science and Pollution Research* 4(1), 250-252. DOI: 10.30799/jespr.122.18040104
- Al-Shatri, A. H. N., Pakyürek, M., and Yavic, A. (2020). "Effect of seaweed application on the vegetative growth of strawberry cv. Albion grown under Iraq ecological conditions," *Applied Ecology and Environmental Research* 18(1), 1211-1225. DOI: 10.15666/aeer/1801\_12111225
- Anjum, S. A., Xie, X. Y., Wang, L.C., Saleem, M. F., Man, C., and Lei, W. (2011).
  "Morphological, physiological and biochemical responses of plants to drought stress," *African Journal of Agricultural Research* 6(9), 2026-2032.
  DOI: 10.5897/AJAR10.027
- Aremu, A. O., Plačková, L., Gruz, J., Bíba, O., Novák, O., Stirk, W. A., Dolezal, K., and Van Staden, J. (2016). "Seaweed-derived biostimulant (Kelpak®) influences endogenous cytokinins and bioactive compounds in hydroponically grown *Eucomis autumnalis*," *Journal Plant Growth Regulation* 35(1), 151-162. DOI: 10.1007/s00344-015-9515-8
- Arioli, T., Mattner, S.W., and Winberg, P.C. (2015). "Applications of seaweed extracts in Australian agriculture: Past, present and future," *Journal of Applied Phycology* 27, 2007-2015. DOI: 10.1007/s10811-015-0574-9
- Arrobas, M., Afonso, S., and Rodrigues, M. Â. (2018). "Diagnosing the nutritional condition of chestnut groves by soil and leaf analyses," *Scientia Horticulturae* 228, 113-121. DOI: 10.1016/j.scienta.2017.10.027

- Association of Official Analytical Chemist (AOAC) (2005). *Official Methods of Analysis*, 18<sup>th</sup> Edition, AOAC International, Gaithersburg, MD, USA.
- Basak, A. (2008). "Effect of preharvest treatment with seaweed products, Kelpak® and Goëmar BM 86®, on fruit quality in apple," *International Journal of Fruit Science* 8(1-2), 1-14. DOI: 10.1080/15538360802365251
- Battacharyya, D., Babgohari, M. Z., Rathor, P., and Prithiviraj, B. (2015). "Seaweed extracts as biostimulants in horticulture," *Scientia Horticulturae* 196, 39-48. DOI: 10.1016/j.scienta.2015.09.012
- Ben Salah, I., Aghrouss, S., Douira, A., Aissam, S., El Alaoui-Talibi, Z., Filali-Maltouf, A., and El Modafar, C. (2018). "Seaweed polysaccharides as bio-elicitors of natural defenses in olive trees against verticillium wilt of olive," *Journal of Plant Interactions* 13(1), 248-255. DOI: 10.1080/17429145.2018.1471528
- Bocanegra, M. P., Lobartini, J. C., and Orioli, G. A. (2006). "Plant uptake of iron chelated by humic acids of different molecular weights," *Communications in Soil Science and Plant Analysis* 37(1-2), 239-248. DOI: 10.1080/00103620500408779
- Bradáčová, K., Weber, N. F., Morad-Talab, N., Asim, M., Imran, M., Weinmann, M., and Neumann, G. (2016). "Micronutrients (Zn/Mn), seaweed extracts, and plant growthpromoting bacteria as cold-stress protectants in maize," *Chemical and Biological Technologies in Agriculture* 3(1), 1-10. DOI: 10.1186/s40538-016-0069-1
- Cabo, S., Morais, M. C., Aires, A., Carvalho, R., Pascual-Seva, N., Silva, A. P., and Gonçalves, B. (2019). "Kaolin and seaweed-based extracts can be used as middle and long-term strategy to mitigate negative effects of climate change in physiological performance of hazelnut tree," *Journal of Agronomy and Crop Science* 206(1), 28-42. DOI: 10.1111/jac.12369
- Calvo, P., Nelson, L., and Kloepper, J. W. (2014). "Agricultural uses of plant biostimulants," *Plant and Soil* 383(1), 3-41. DOI: 10.1007/s11104-014-2131-8
- Canellas, L. P., Olivares, F. L., Okorokova-Façanha, A. L., and Façanha, A. R. (2002). "Humic acids isolated from earthworm compost enhance root elongation, lateral root emergence, and plasma membrane H+-ATPase activity in maize roots," *Plant Physiology* 130(4), 1951-1957. DOI: 10.1104/pp.007088
- Canellas, L. P., Olivares, F. L., Aguiar, N. O., Jones, D. L., Nebbioso, A., Mazzei, P., and Piccolo, A. (2015). "Humic and fulvic acids as biostimulants in horticulture," *Scientia Horticulturae* 196, 15-27. DOI: 10.1016/j.scienta.2015.09.013
- Circuncisão, A. R., Catarino, M. D., Cardoso, S. M., and Silva, A. (2018). "Minerals from macroalgae origin: Health benefits and risks for consumers," *Marine Drugs* 16(11), 1-30. DOI: 10.3390/md16110400
- Chen, Y., Clapp C. E., and Magen, H. (2004). "Mechanisms of plant growth stimulation by humic substances: The role of organo-iron complexes," *Soil Science and Plant Nutrition* 50(7), 1089-1095. DOI: 10.1080/00380768.2004.10408579
- Chouliaras, V., Tasioula, M., Chatzissavvidis, C., Therios, I., and Tsabolatidou, E. (2009). "The effects of a seaweed extract in addition to nitrogen and boron fertilization on productivity, fruit maturation, leaf nutritional status and oil quality of the olive (*Olea europaea* L.) cultivar Koroneiki," *Journal of the Science of Food and Agriculture* 89(6), 984-988. DOI: 10.1002/jsfa.3543
- Colavita, G. M., Spera, N., Blackhall, V. and Sepulveda, G. M. (2011). "Effect of seaweed extract on pear fruit quality and yield," *Acta Horticulturae* 909, 601-607 DOI: 10.17660/ActaHortic.2011.909.72

- Craigie, J. S. (2011). "Seaweed extract stimuli in plant science and agriculture," *Journal* of Applied Phycology 23(3), 371-393. DOI: 10.1007/s10811-010-9560-4
- Demirsoy, H. (2009). "Leaf area estimation in some species of fruit tree by using models as a non-destructive method," *Fruits* 64(1), 45-51. DOI: 10.1051/fruits/2008049
- de Sousa, A. M., Ayub, R. A., Viencz T., and Botelho, R. V. (2019). "Fruit set and yield of apple trees cv. Gala treated with seaweed extract of *Ascophyllum nodosum* and *thidiazuron*," *Revista Brasileira de Fruticultura* 41(1), (e-072). DOI: 10.1590/0100-29452019072
- Duan, Y., Xu, M., Gao, S., Liu, H., Huang, S., and Wang, B. (2016). "Long-term incorporation of manure with chemical fertilizers reduced total nitrogen loss in rain-fed cropping systems," *Scientific Reports* 6(1), 1-10. DOI: 10.1038/srep33611
- Elansary, H. O., Norrie, J., Ali, H. M., Salem, M. Z. M., Mahmoud, E. A., and Yessoufou, K. (2016). "Enhancement of *Calibrachoa* growth, secondary metabolites and bioactivity using seaweed extracts," *BMC Complementary and Alternative Medicine* 16, article no. 341. DOI: 10.1186/s12906-016-1332-5
- El-Boray, M. S., Mostafa, M. F., Shaltout, A. D., and Hassan, K. H. (2015). "Influence of fulvic acid plus some microelements and microorganisms on yield and quality characteristics of superior seedless grapevines," *Journal of Plant Production* 6(3), 287-305. DOI: 10.21608/jpp.2015.49320
- El-Boukhari, M. E., Barakate, M., Bouhia, Y., and Lyamlouli, K. (2020). "Trends in seaweed extract based biostimulants: Manufacturing process and beneficial effect on soil-plant systems," *Plants* 9(359), 1-23. DOI: 10.3390/plants9030359
- El-Helaly, M. A. (2018). "Effect of foliar application of humic and fulvic acids on yield and its components of some carrot (*Daucus carota* L.) cultivars," *Journal of Horticultural Science & Ornamental Plants* 10(3), 159-166. DOI: 10.5829/idosi.jhsop.2018.159.166
- El-kenawy, M. A. (2017). "Effect of chitosan, salicylic acid and fulvic acid on vegetative growth, yield and fruit quality of Thompson seedless grapevines," *Egyptian Journal of Horticulture* 44(1), 45-59. DOI: 10.21608/EJOH.2017.1104.1007
- El-Miniawy, S. M., Ragab, M. E., Youssef, S. M., and Metwally, A. A. (2014).
  "Influence of foliar spraying of seaweed extract on growth, yield and quality of strawberry plants," *Journal of Applied Sciences Research* 10(2), 88-94.
- El-Sharony, T. F., El-Gioushy, S. F., ans Amin, O. A. (2015). "Effect of foliar application with algae and plant extracts on growth, yield and fruit quality of fruitful mango trees cv. *Fagri Kalan*," *Journal of Horticulture* 2(4),1-6. DOI:10.4172/2376-0354.1000162
- Eyheraguibel, B., Silvestre, J., and Morard, P. (2008). "Effects of humic substances derived from organic waste enhancement on the growth and mineral nutrition of maize," *Bioresource Technology* 99(10), 4206-4212. DOI: 10.1016/j.biortech.2007.08.082
- Fan, D., Hodges, D. M., Critchley, A. T., and Prithiviraj, B. (2013). "A commercial extract of brown macroalga (*Ascophyllum nodosum*) affects yield and the nutritional quality of spinach *in vitro*," *Communications in Soil Science and Plant Analysis* 44(12), 1873-1884. DOI: 10.1080/00103624.2013.790404
- Gomathi, R., Kohila, S., and Ramachandiran, K. (2017). "Evaluating the effect of seaweed formulations on the quality and yield of sugarcane," *Madras Agricultural Journal* 104 (4-6), 161-165, DOI:10.29321/MAJ 2019.000213

- Gupta, V., Kumar, M., Brahmbhatt, H., Reddy, C. R. K., Seth, A., and Jha, B. (2011). "Simultaneous determination of different endogenetic plant growth regulators in common green seaweeds using dispersive liquid–liquid microextraction method," *Plant Physiology and Biochemistry* 49(11), 1259-1263. DOI: 10.1016/j.plaphy.2011.08.004
- Haggag, L. F., Fawzi, M., Shahin, M. and EL-Hady, E. S. (2016). "Effect of yeast, humic acid, fulvic acid, citric acid, potassium citrate and some chelated micro-elements on yield, fruit quality and leaf minerals content of "Canino" apricot trees," *International Journal Chemistry Technology Research* 9(4), 07-15.
- Harhash M. M., Abd EL-Megeed N. A., Abaidalah A. S., and Mosa W. F. A. (2021a).
  "Effect of the foliar spraying of fulvic acid, folic acid, and seaweed extract on vegetative growth, yield and fruit quality of grape cv. Flame seedless," *Plant Archives* 21(1), 482-492. DOI: 10.51470/PLANTARCHIVES.2021.v21.no1.068
- Harhash M. M., Saad R. M., and Mosa W. F. A. (2021b). "Response of "Wonderful" pomegranate cultivar to the foliar application of some biostimulants," *Plant Archives* (in press).
- Hayyawi, N. J. H., Al-Issawi, M. H., Alrajhi, A. A., Al-Shmgani, H., and Rihan, H. (2020). "Molybdenum induces growth, yield, and defense system mechanisms of the mung bean (*Vigna radiata* L.) under water stress conditions," *International Journal of Agronomy* 2020, article no. 8887329. DOI: 10.1155/2020/8887329
- Huang, S., Xiong, B., Sun, G., He, S., Liao, L., Wang, J., Wang, B., and Wang, Z. (2020). "Effects of fulvic acid on photosynthetic characteristics of citrus seedlings under drought stress," *IOP Conference Series: Earth and Environmental Science* 474(2020). DOI: 10.1088/1755-1315/474/3/032007
- Hui, L. I., Feng, W. T., He, X. H., Ping, Z. H. U., Gao, H. J., Nan, S. U. N., and Xu, M. G. (2017). "Chemical fertilizers could be completely replaced by manure to maintain high maize yield and soil organic carbon (SOC) when SOC reaches a threshold in the Northeast China Plain," *Journal of Integrative Agriculture* 16(4), 937-946. DOI: 10.1016/S2095-3119(16)61559-9
- Justi, M., Morais, E. G., and Silva, C. A. (2019). "Fulvic acid in foliar spray is more effective than humic acid *via* soil in improving coffee seedlings growth," *Archives of Agronomy and Soil Science* 65(14), 3-17. DOI: 10.1080/03650340.2019.1584396
- Kapur, B., Sarıdaş, M. A., Çeliktopuz, E., Kafkas, E., and Kargı, S. P. (2018). "Health and taste related compounds in strawberries under various irrigation regimes and biostimulant application," *Food Chemistry* 263, 67-73. DOI: 10.1016/j.foodchem.2018.04.108
- Khan, O. A., Sofi, J. A., Kirmani, N. A., Hassan, G. I., Bhat, S. A., Chesti, M. H., and Ahmad, S. M. (2019). "Effect of N, P and K Nano-fertilizers in comparison to humic and fulvic acid on yield and economics of red delicious (*Malus domestica* Borukh.)," *Journal of Pharmacognosy and Phytochemistry* 8(2), 978-981
- Khan, W., Rayirath, U. P., Subramanian, S., Jithesh, M. N., Rayorath, P., Hodges, D. M., Critchley, A. T., Craigie, J. S., Norrie, J., and Prithiviraj, B. (2009). "Seaweed extracts as biostimulants of plant growth and development," *Journal of Plant Growth Regulation* 28(4), 386-399. DOI: 10.1007/s00344-009-9103-x

- Khan, Z. H., Khan, M. M. A., Aftab, T., Idrees, M., and Naeem, M. (2011). "Influence of alginate oligosaccharides on growth, yield and alkaloid production of opium poppy (*Papaver somniferum* L.)," *Frontiers of Agriculture in China* 5(1), 122-127. DOI: 10.1007/s11703 -010-1056-0
- Khompatara, K., Pettongkhao, S., Kuyyogsuy, A., Deenamo, N., and Churngchow, N. (2019). "Enhanced resistance to leaf fall disease caused by *Phytophthora palmivora* in rubber tree seedling by *Sargassum polycystum* extract," *Plants* 8(6), 1-14. DOI: 10.3390/plants8060168
- Kocira, A., Świeca, M., Kocira, S., Złotek, U., and Jakubczyk, A. (2018). "Enhancement of yield, nutritional and nutraceutical properties of two common bean cultivars following the application of seaweed extract (*Ecklonia maxima*)," *Saudi Journal of Biological Sciences* 25(3), 563-571. DOI: 10.1016/j.sjbs.2016.01.039
- Kulkarni, M. G., Rengasamy, K. R. R., Pendota, S. C., Gruz, J., Plačková, L., Novák, O., Doležal, K., and Van Staden, J. (2019). "Bioactive molecules derived from smoke and seaweed *Ecklonia maxima* showing phytohormone-like activity in *Spinacia oleracea* L.," *New Biotechnology* 48, 83-89. DOI: 10.1016/j.nbt.2018.08.004
- Lall, D., Prasad, V. M., Singh, V. K., and Kiishor, S. (2017). "Effect of foliar application of Biovita (biofertilizer) on fruit set, yield and quality of guava (*Psidium guajava* L.)," *Research in Environment and Life Science* 10(5), 432-434.
- Li, M.-S., Li, S., Zhang, S.-Y., and Chi, B.-L. (2005). "Physiological effect of new FA antitranspirant application on winter wheat at ear filling stage," *Chinese Agriculture Science* 38(4), 703-708.
- Liu, X., Zhang, Y., Han, W., Tang, A., Shen, J., Cui, Z., Vitousek, P., Erisman, J.W., Goulding, K., Christie, P. and Zhang, F. (2013). "Enhanced nitrogen deposition over China," *Nature* 494(7438), 459-462. DOI: 10.1038/nature11917
- Lotfi, R., Pessarakli, M., Gharavi-Kouchebagh, P., and Khoshvaghti, H. (2015). "Physiological responses of *Brassica napus* to fulvic acid under water stress: Chlorophyll a fluorescence and antioxidant enzyme activity," *The Crop Journal* 3(5), 434-439. DOI: 10.1016/j.cj.2015.05.006
- Lu, J. W., and Beamish, P. W. (2001). "The internationalization and performance of SMEs," *Strategic Management Journal* 22(6-7), 565-586. DOI: 10.1002/smj.184
- Machado, L. P., Matsumoto, S. T., Jamal, C. M., da Silva, M. B., da Cruz Centeno, D., Neto, P. C., de Carvalho, L.R., and Yokoya, N. S. (2014). "Chemical analysis and toxicity of seaweed extracts with inhibitory activity against tropical fruit anthracnose fungi," *Journal of the Science of Food and Agriculture* 94(9), 1739-1744. DOI: 10.1002/jsfa.6483
- Mackowiak, C., Grossl, P., and Bugbee, B. (2001). "Beneficial effects of humic acid on micronutrient availability to wheat," *Soil Science Society of America Journal* 65(6), 1744-1750. DOI: 10.2136/sssaj2001.1744
- Mancuso, S., Azzarello, E., Mugnai, S., and Briand, X. (2006). "Marine bioactive substances (IPA extract) improve foliar ion uptake and water stress tolerance in potted *Vitis vinifera* plants," *Advances in Horticultural Science* 20(2), 156-161.
- Mattner, S. W., Milinkovic, M., and Arioli, T. (2018). "Increased growth response of strawberry roots to a commercial extract from *Durvillaea potatorum* and *Ascophyllum nodosum*," J. Applied Phycology 30, 2943-2951. DOI: 10.1007/s10811-017-1387-9
- Mostafa, M. F. M., EL-Boray, M. S., El-Baz, E. L., and Omar, A. S. (2017). "Effect of fulvic acid and some nutrient elements on king ruby grapevines growth, yield and chemical properties of berries," *Journal of Plant Production* 8(2), 321-328. DOI:

10.21608/jpp.2017.39630

- Nardi, S., Pizzeghello, D., Muscolo, A., and Vianello, A. (2002). "Physiological effects of humic substances on higher plants," *Soil Biology and Biochemistry* 34(11), 1527-1536. DOI: 10.1016/S0038-0717(02)00174-8
- Neumann, D., and Zur Nieden, U. (2001). "Silicon and heavy metal tolerance of higher plants," *Phytochemistry* 56(7), 685-692. DOI: 10.1016/S0031-9422(00)00472-6
- Ni, B., Liu, M., Lu, S., Xie, L., and Wang, Y. (2011). "Environmentally friendly slowrelease nitrogen fertilizer," *Journal of Agricultural and Food Chemistry* 59(18), 10169-10175. DOI: 10.1021/jf202131z
- Nielsen, S. S. (2010). "Phenol-sulfuric acid method for total carbohydrates," in: *Food Analysis Laboratory Manual*, S. S. Nielsen (ed.), Springer, Boston, MA, USA, pp. 47-53. DOI: 10.1007/978-1-4419-1463-7
- Nielsen, S. S. (2017). "Vitamin C determination by indophenol method," in: *Food Analysis Laboratory Manual*, S S. Nielsen (ed.), Springer, Boston, MA, USA, 143-146. DOI: 10.1007/978-3-319-44127-6\_32
- Ning, C. C., Gao, P. D., Wang, B. Q., Lin, W. P., Jiang, N. H., and Cai, K. Z. (2017). "Impacts of chemical fertilizer reduction and organic amendments supplementation on soil nutrient, enzyme activity and heavy metal content," *Journal of Integrative Agriculture* 16(8), 1819-1831. DOI: 10.1016/S2095-3119(16)61476-4
- Ott, R. L., and Longnecker, M. T. (2015). An Introduction to Statistical Methods and Data Analysis, Cengage Learning, Boston, MA, USA.
- Parthiban, C., Saranya, C., Girija, K., Hemalatha, A., Suresh, M., and Anantharaman, P. (2013). "Biochemical composition of some selected seaweeds from Tuticorin coast," *Advances in Applied Science Research* 4(3), 362-366.
- Patel, R. V., Pandya, K. Y., Jasrai, R. T., and Brahmbhatt, N. (2018). "Significance of green and brown seaweed liquid fertilizer on seed germination of *Solanum melongena*, *Solanum lycopersicum* and *Capsicum annum* by paper towel and pot method," *International Journal of Recent Scientific Research* 9, 24065-24072.
- Piccolo, A., and Mbagwu, J. S. C. (1999). "Role of hydrophobic components of soil organic matter in soil aggregate stability," *Soil Science Society of America Journal* 63(6), 1801-1810. DOI: 10.2136/sssaj1999.6361801x
- Prasad, K., Das, A. K., Oza, M. D., Brahmbhatt, H., Siddhanta, A. K., Meena, R., Eswaran, K., Rajyaguru, M. R., and Ghosh, P. K. (2010). "Detection and quantification of some plant growth regulators in a seaweed-based foliar spray employing a mass spectrometric technique sans chromatographic separation," *Journal* of Agricultural and Food Chemistry 58(8), 4594-4601. DOI: 10.1021/jf904500e
- Priya, B. N. V., Mahavishnan K., Gurumurthy D. S., Bindumadhava H., Upadhyay A. P., and Sharma N.K. (2014). "Fulvic acid (FA.) for enhanced nutrient uptake and growth: Insights from biochemical and genomic studies," *Journal of Crop Improvement*, 28(6), 740-757. DOI:10.1080/15427528.2014.923084
- Rathore, S. S., Chaudhary, D. R., Boricha, G. N., Ghosh, A., Bhatt, B. P., Zodape, S. T., and Patolia, J. S. (2009). "Effect of seaweed extract on the growth, yield and nutrient uptake of soybean (*Glycine max*) under rainfed conditions," *South African Journal of Botany* 75(2), 351-355. DOI: 10.1016/j. sajb.2008.10.009
- Ravi, I., Kamaraju, K., Kumar, S., and Nori, S. S. (2018). "Foliar application of seaweed bio formulation enhances growth and yield of banana cv. Grand Naine (AAA)," *Indian Journal of Natural Sciences* 8(47), 13482-13488.
- Razavi, S. M. A., and BahramParvar, M. (2007). "Some physical and mechanical

Mosa *et al.* (2021). "Biostimulants & growth," *BioResources* 16(4), 7379-7399.

properties of kiwifruit," *International Journal of Food Engineering* 3(6), 1-14. DOI: 10.2202/1556-3758.1276

- Renaut, S., Masse, J., Norrie, J. P., Blal, B., and Hijri, M. (2019). "A commercial seaweed extract structured microbial communities associated with tomato and pepper roots and significantly increased crop yield," *Microbial Biotechnology* 12(6),1346-1358. DOI: 10.1111/1751-7915.13473
- Rioux, L. E., Turgeon, S. L., and Beaulieu, M. (2009). "Effect of season on the composition of bioactive polysaccharides from the brown seaweed *Saccharina longicruris*," *Phytochemistry* 70(8), 1069-1075. DOI: 10.1016/j.phytochem.2009.04.020
- Rouphael, Y., Giordano, M., Cardarelli, M., Cozzolino, E., Mori, M., Kyriacou, M. C., Bonini, P., and Colla, G. (2018). "Plant-and seaweed-based extracts increase yield but differentially modulate nutritional quality of greenhouse spinach through biostimulant action," *Agronomy* 8(126), 1-15. DOI: 10.3390/agronomy8070126
- Samavat, S., and Samavat, S. (2014). "The effects of fulvic acid and sugar cane molasses on yield and qualities of tomato," *International Research Journal of Applied Basic Sciences* 8(3), 266-268.
- Schmidt, W., Santi, S., Pinton, R., and Varanini, Z. (2007). "Water-extractable humic substances alter root development and epidermal cell pattern in *Arabidopsis*," *Plant and Soil* 300(1), 259-267. DOI: 10.1007/s11104-007-9411-5
- Sierra, J., Causeret, F., Diman, J. L., Publicol, M., Desfontaines, L., Cavalier, A., and Chopin, P. (2015). "Observed and predicted changes in soil carbon stocks under export and diversified agriculture in the Caribbean. The case study of Guadeloupe," *Agriculture, Ecosystems and Environment* 213, 252-264. DOI: 10.1016/j.agee.2015.08.015
- Sivasankari, S., Venkatesalu, V., Anantharaj, M., and Chandrasekaran, M. (2006). "Effect of seaweed extracts on the growth and biochemical constituents of *Vigna sinensis*," *Bioresource Technology* 97(14), 1745-1751. DOI: 10.1016/j.biortech.2005.06.016
- Smith, L. E., and Siciliano, G. (2015). "A comprehensive review of constraints to improved management of fertilizers in China and mitigation of diffuse water pollution from agriculture," *Agriculture, Ecosystems and Environment* 209, 15-25. DOI: 10.1016/j.agee.2015.02.016
- Soppelsa, S., Kelderer, M., Casera, C., Bassi, M., Robatscher, P., and Andreotti, C. (2018). "Use of biostimulants for organic apple production: effects on tree growth, yield, and fruit quality at harvest and during storage," *Frontiers in Plant Science* 9(1342), 1-17. DOI: 10.3389/fpls.2018.01342
- Sparks, D. L., Page, A. L., Helmke, P. A., and Loeppert, R. H. (2016). *Methods of Soil Analysis, Part 3: Chemical Methods*, John Wiley & Sons, Hoboken, NJ, USA.
- Stirk, W. A., and Van Staden, J. (2014). "Plant growth regulators in seaweeds: Occurrence, regulation and functions," *Advances in Botanical Research* 71, 125-159. DOI: 10.1016/B978-0-12-408062-1.00005-6
- Stirk, W., Novák, O., Strnad, M., and Van Staden, J. (2003). "Cytokinins in macroalgae," *Plant Growth Regulation* 41(1), 13-24. DOI: 10.1023/A:1027376507197
- Stuart, D., Schewe, R. L., and McDermott, M. (2014). "Reducing nitrogen fertilizer application as a climate change mitigation strategy: Understanding farmer decisionmaking and potential barriers to change in the US," *Land Use Policy* 36, 210-218. DOI: 10.1016/j.landusepol.2013.08.011
- Suh, H. Y., Yoo, K. S., and Suh, S. G. (2014). "Effect of foliar application of fulvic acid

on plant growth and fruit quality of tomato (*Lycopersicon esculentum* L.)," *Horticulture, Environment, Biotechnology* 55(6), 455-461. DOI: 10.1007/s13580-014-0004-y

Suhag, M. (2016). "Potential of biofertilizers to replace chemical fertilizers," *International Advanced Research Journal in Science, Engineering and Technology* 3(5), 163-167. DOI 10.17148/IARJSET.2016.3534.

Taha, A. A., Omar, M. M., and Ghazy, M. A. (2016). "Effect of humic and fulvic acids on growth and yield of lettuce plant," *Journal of Soil Sciences and Agricultural Engineering* 7(8), 517-522. DOI: 10.21608/jssae.2016.39782

- Uphoff, N., and Dazzo, F. B. (2016). "Making rice production more environmentallyfriendly," *Environments* 3(12), 1-7. DOI: 10.3390/environments3020012
- Valencia, R. T., Acosta, L. S., Hernández, M. F., Rangel, P. P., Robles, M. Á. G., Rocío del Cruz, A., and Vázquez, C.V. (2018). "Effect of seaweed aqueous extracts and compost on vegetative growth, yield, and nutraceutical quality of cucumber (*Cucumis sativus* L.) fruit," *Agronomy*, 8(264), 1-3. DOI: 10.3390/agronomy8110264
- Wang, H., Pampati, N., McCormick, W. M., and Bhattacharyya, L. (2016). "Protein nitrogen determination by Kjeldahl digestion and ion chromatography," *Journal of Pharmaceutical Sciences* 105(6), 1851-1857. DOI: 10.1016/j.xphs.2016.03.039
- Wang, Y., Yang, R., Zheng, J., Shen, Z., and Xu, X. (2019). "Exogenous foliar application of fulvic acid alleviate cadmium toxicity in lettuce (*Lactuca sativa* L.)," *Ecotoxicology and Environmental Safety* 167, 10-19. DOI: 10.1016/j.ecoenv.2018.08.064.
- Wang, Y., Zhu, Y., Zhang, S., and Wang, Y. (2018). "What could promote farmers to replace chemical fertilizers with organic fertilizers?" *Journal of Cleaner Production* 199, 882-890. DOI: 10.1016/j.jclepro.2018.07.222
- Wassel, A. M. M., Gobara A. A., Rizk E. A., and El-Wany A. R. M. (2014). "Reducing mineral N fertilizer partially in 'Thompson Seedless' vineyards by using fulvic acid and effective microorganisms," *World Rural Observ.* 6(4), 36-42.
- Weiwei, C., Jinrong, L., Fang, X., and Jing, L. (2017). "Improvement to the determination of activated phosphorus in water and wastewater by yellow vanadomolybdate method," *Industrial Water Treatment* 37(2), 95-97. DOI: 10.11894/1005-829x.2017.37(2).095
- Yalçın, S., Şükran Okudan, E., Karakaş, Ö., Önem, A. N., and Başkan, K. S. (2019). "Identification and quantification of some phytohormones in seaweeds using UPLC-MS/MS," *Journal of Liquid Chromatography & Related Technologies*, 42(15-16), 1-10. DOI: 10.1080/10826076.2019.1625374
- Yang, S., Zhang, Z., Cong, L., Wang, X., and Shi, S. (2013). "Effect of fulvic acid on the phosphorus availability in acid soil," *Journal of Soil Science and Plant Nutrition* 13(3), 526-533. DOI: 10.4067/S0718-95162013005000041
- Yildirim, E. (2007). "Foliar and soil fertilization of humic acid affect productivity and quality of tomato," *Acta Agriculturae Scandinavica Section B-Soil and Plant Science* 57(2), 182-186. DOI: 10.1080/09064710600813107
- Youssef, M. M. A., and Eissa, M. F. M. (2014). "Biofertilizers and their role in management of plant parasitic nematodes (A review)," *Journal of Biotechnology and Pharmaceutical Research* 5(1), 1-6.
- Zhang, X., and Ervin, E. (2004). "Cytokinin-containing seaweed and humic acid extracts associated with creeping bentgrass leaf cytokinins and drought resistance," *Crop Science* 44(5), 1737-1745. DOI: 10.2135/cropsci2004.1737

- Zimmerli, L., Hou, B. H., Tsai, C. H., Jakab, G., Mauch-Mani, B., and Somerville, S. (2008). "The xenobiotic β-aminobutyric acid enhances Arabidopsis thermotolerance," *The Plant Journal* 53(1), 144-156. DOI: 10.1111/j.1365-313X.2007.03343
- Zodape, S. T., Gupta, A., Bhandari, S. C., Rawat, U. S., Chaudhary, D. R., Eswaran, K., and Chikara, J. (2011). "Foliar application of seaweed sap as biostimulant for enhancement of yield and quality of tomato (*Lycopersicon esculentum* Mill.)," *Journal of Scientific & Industrial Research* 70(3), 215-219.

Article submitted: July 31, 2021; Peer review completed: September 5, 2021; Revised version received and accepted: September 14, 2021; Published: September 17, 2021. DOI: 10.15376/biores.16.4.7379-7399