Effects of the Membrane-covered Technology and Superphosphate on the Compost Quality and Nitrogencontaining Gas Emissions during Aerobic Composting

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A lab-scale experiment was conducted to assess effects of the membranecovered technology and superphosphate on the compost quality and emissions of nitrogen-containing gases during aerobic composting. The results showed that the membrane-covered technology increased the temperature of the compost pile and accelerated degradation of organic matter. The membrane-covered (T1) sample attained a germination index (GI) of 50% and 80%, approximately 2 and 9 d earlier, respectively, relative to the control (CK) sample. However, the application of superphosphate might have an adverse effect on the GI value. The NH₃ and N₂O emissions during the aerobic composting were decreased by 25.8% and 13.1%, respectively, for the T1 sample compared to the CK sample, while these were reduced correspondingly by 21.3% and 27.2% for the superphosphate addition (T2) sample. Compared with the superphosphate addition, the membrane-covered approach reduced the NH₃ emission but not the N₂O emission. Thus, the membrane-covered aerobic composting is a potential technology for the adequate utilization of organic waste as a resource.

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INTRODUCTION

Aerobic composting is a convenient and popular method for the exploitation of organic wastes from agriculture as a resource. Products from this composting are suitable alternatives for chemical fertilizers, elevating the organic matter contents of soils, enhancing of soil's fertility (Hubbe *et al.* 2010), and improving the quality of crops.

Nitrogen (N), the main component of proteins, significantly contributes to the growth of plants, including the production of fruits. In fact, it is the nutrient with the greatest impact on crop yield. However, during aerobic composting, N is mainly lost in the form of ammonia (NH₃) and nitrous oxide (N₂O), that is not only the loss of nutrients in composting products, and these products are secondary pollutants of the environment. The nitrogen cycle is illustrated in Fig. 1.

In the process of aerobic composting, organic nitrogen decomposes and produces ammonium nitrogen by ammoniation. Ammonium nitrogen escapes easily from the composting system in the form of NH₃ under high temperature (65 to 70 °C) and pH (8.4 to 9.0) environmental conditions. N₂O is mainly produced by nitrification and

denitrification on the surface and inside the compost pile, respectively. Under the condition that the raw materials are determined, researchers mostly choose to change the nitrogen chemical process by adding exogenous substances (Jiang et al. 2016; Chen et al. 2016) to reduce NH_3 and N_2O emissions during aerobic composting. Therefore, additives, such as superphosphate, zeolite, woody peat, and biochar, are commonly added to reduce the loss of N during the composting process (Ding et al. 2019). According to Yuan et al. (2018), phosphogypsum and dicyandiamide can reduce methane (CH₄) and N₂O emissions by 75.6% and 86.4%, respectively; however, the NH_3 emission can increase by 22.0%. Jiang et al. (2014) used an aerated static box to demonstrate that superphosphate facilitates the composting process. It significantly reduced the emission of NH₃ emission during the thermophilic phase, and thereby it elevated the N and phosphorus (P) contents in the compost pile. Conversely, bentonite increased the emission of NH₃, and thus reduced the N content of the compost pile. Zhang et al. (2017) indicated that although the utilization of superphosphate elevated the emission of N₂O, it reduced emissions of CH₄, NH₃, and H₂S by approximately 35.5%, 37.9%, and 65.5%, respectively. Overall, its application reduced the total emission of greenhouse gases by approximately 34.7%. Among all additives, superphosphate has been widely used because of its low cost and the possibility of appropriately increasing the P content in composting products.



Fig. 1. Nitrogen cycle during aerobic composting

Membrane-covered aerobic composting is a technology that has gained popularity in recent years. In contrast to the traditional trough and windrow composting methods in which oxygen is supplied by mechanical turning, in the membrane-covered system, the oxygen required by aerobic microorganisms is pumped in the form of air from a pipeline beneath the compost. The molecular membrane cover is the most important component of the system. Its main structure is expanded polytetrafluoroethylene (e-PTFE), which is characterized by evenly distributed micropores. These pores can prevent the diffusion of volatile organic compounds (VOCs), bioaerosols, and dust, but permit the diffusion of small gas molecules into the atmosphere. Conversely, the water film formed through the condensation of water vapor on the inner side of the membrane contains dissolved gases, such as NH₃, which can return to the compost pile by dripping (Ma *et al.* 2017). Concurrently, the molecular membrane creates a relatively stable composting environment, which is resistant to external adverse factors such as rain, wind, and ultraviolet light. Therefore, the membrane-covered composting system is suitable for open-air operation, and this can significantly reduce the costs associated with the construction of composting sites.

Owing to the high efficiency, low operation cost, and low gas emission, the membrane-covered aerobic composting technology is increasingly being studied and utilized worldwide. González *et al.* (2016) evaluated the composting process using an industrial-scale system and showed that the membrane-covered technology increased the temperature of the compost pile and prolonged the duration of the thermophilic phase. According to Ma *et al.* (2021), the molecular membrane produced by the Qingdao Zhiteng Technology Company decreased N₂O and CO₂ emissions by 68.4% and 1.56%, respectively, and thus, it reduces the global warming potential by 53.4%. Further, Li *et al.* (2020) demonstrated that the membrane-covered composting system decreased NH₃ and H₂S emissions by 58.6% and 38.1%, respectively, while the rate of loss of total N from the compost pile reduced by 17.3%. The ammonium nitrogen and nitrate nitrogen contents of the pile also increased correspondingly by 37.7% and 11.8%.

According to previous studies, both the superphosphate and membrane cover application can be expected to reduce the emission of gases during aerobic composting. However, the effects of both methods on the N-containing gases emitted and the compost quality have rarely been reported. In addition, because of the intensive development of agriculture in China, the replacement of chemical fertilizers with organic fertilizers in the agriculture industry is desired for the protection of the rural environmental. Therefore, the utilization of organic wastes from agriculture as fertilizers is increasing in China. In the present study, stalks of the tomato plant and mushroom residues were utilized as feedstock to evaluate the effects of the membrane-covered and superphosphate approaches on parameters associated with aerobic composting including the emission of NH₃ and N₂O. The present study highlights the potential of the membrane-covered aerobic composting technology for the utilization of agricultural wastes as fertilizers.

EXPERIMENTAL

Materials

Fresh stalks of the tomato plant and mushroom residues were collected from a vegetable park on Guli Street in Jiangsu Province, Nanjing, China. The stalks were sliced to produce straws ranging between 1 to 3 cm in length, and these were mixed with mushroom residues at a ratio of 4:1 (dry weight). The moisture content of the compost pile was adjusted to approximately 60% using water during the mixing, and the initial carbon to nitrogen (C/N) ratio was approximately 23:1. In the composting, a microbial community comprising the *Bacillus licheniformis*, *Bacillus subtilis*, *Streptomyces rochei*, and *Aspergillus oryzae*, which was produced by the Jiangsu Academy of Agricultural Sciences (Jiangsu Province, Nanjing, China) was employed, and this yielded an effective viable bacteria concentration of more than 50 million/g. Data for the physicochemical properties of the compost are presented in Table 1.

Parameters	TC (%)	TN (%)	C/N	MC (%)
Tomato stalks	42.3 ± 1.21	1.65 ± 0.07	25.7 ± 0.64	25.7 ± 1.34
Mushroom residues	32.4 ± 1.45	1.77 ± 0.04	17.9 ± 0.69	54.4 ± 0.87
Mixture	39.6 ± 0.45	1.72 ± 0.04	23.1 ± 0.25	62.8 ± 2.96

Table 1. Physicochemical Properties of the Composting Samples

TC: Total carbon; TN: Total nitrogen; C/N: the ratio of total carbon to total nitrogen; MC: Moisture content

Experimental Design

The aerobic composting device utilized mainly comprised a 500 L plastic tank with an aeration pipe at the bottom, three solid sampling ports at the top, side, and bottom, and a gas collection and sampling device at the top. The adopted molecular membrane was about 3 mm thick and mainly had three layers, the inner and outer layers are polyester fiber, which provided good corrosion and scratch resistance for the membrane, and the middle layer was e-PTFE. The permeability index was 0.028 m³/min·m at 125 Pa. Experiments were conducted using the following groups of samples: the T1 group involving the molecular membrane covering at the top of the compost pile, the T2 group containing 5% superphosphate (dry weight) of initial raw materials, and the control group (CK) representing the compost without the membrane covering or superphosphate application. Air was pumped into each compost pile from the bottom at a rate of 0.1 m³/min m³ during the experiments.

Sample Collection and Analysis

Samples were collected from the upper, middle, and lower layers of the compost pile on day 0, 3, 9, 15, 21, 27, and 33 at 2 p.m. during the experiments. These samples were thoroughly mixed and divided into two subsamples. The first subsample was used to determine the pH and EC instantly, while the other subsample was stored at -20 °C in a refrigerator for the subsequent analysis of the germination index (GI), TC, and TN. The temperature was automatically recorded once in 24 h using a digital thermometer, while the moisture content was measured by drying to a constant weight at 105 °C (Tian et al. 2012). The compost samples were then mixed with deionized water at a ratio of 1:10 (w/w), and the mixture was shaken for 30 min at the speed of 150 r/min. The mixture was allowed to settle for 4 h, and then filtered to obtain the supernatant that was employed for measurement of the pH and EC by a detector (FE28, Mettler-Toledo, Shanghai, China) (Wang et al. 2015). Ten Chinese cabbage seeds were distributed on filter paper in Petri dishes and moistened with 5 mL of the compost water extract. Incubation was performed at 25 ± 2 °C for 48 h, and the GI was calculated based on the seed germination rate and root length. For the CK samples, distilled water instead of the compost water extract was used. The GI was calculated as follows:

 ${\rm GI} = \frac{{\rm seed \ germination \ of \ the \ treatment \ (\%) \times root \ length \ of \ the \ treatment \ (mm)}}{{\rm seed \ germination \ of \ the \ control \ (\%) \times root \ length \ of \ the \ control \ (mm)}} \ x \ 100$

The error associated with the ruler used to measure the root length was less than 1 mm. Gas samples were collected using an interval of two days during the duration of the experiment, the concentrations of NH_3 and N_2O were determined correspondingly using a detection tube (GASTEC, Kanagawa, Japan) and a gas chromatograph (Agilent Technologies, Santa Clara, CA, USA).

Data and Statistical Analyses

Excel 2016 (Microsoft Corp, Redmond, WA, USA) and SPSS 22 (IBM, Armonk, NY, USA) were utilized for all statistical analyses. All parameter measurements (temperature, pH, EC, GI, NH₃ and N₂O) were performed in triplicates for the same sample. One-way analysis of variance (ANOVA) was used to analyze the data. When an ANOVA result was significant, the means were compared with Fisher's protected least significant difference test at $p \leq 0.05$. Prior to the ANOVA, the normality of distribution and homogeneity of variance were checked with Shapiro-Wilk and Levene's test, respectively. Plots were generated using OriginPro 9.1 software (OriginLab, Northampton, MA, USA).

RESULTS AND DISCUSSION

Changes in Physicochemical Parameters of Composting Samples

Changes in temperature

Temperature is an important parameter in the aerobic composting process, and thus, it is employed to characterize the process into the warming, thermophilic, and cooling phases. To eliminate the impacts of insect eggs, pathogens, and other harmful organisms in the compost material, the thermophilic phase is generally required to attain a temperature of at least 50 °C and last between 4 to 5 days (Li *et al.* 2018).



Fig. 2. Plots showing changes in temperature during the composting of three groups

The changes in temperature during the composting of different groups displayed in Fig. 2 are typical of aerobic composting. The temperatures of the T1, T2, and CK groups increased rapidly during the first two days, and reached maximum temperatures of 70.2, 67.7, and 63.8 °C, respectively, on 4th, 3rd, and 4th day, respectively. The increase in

temperature in the compost was attributed to heat generated by microorganisms using C and N sources. Figure 1 also reveals that the thermophilic phase of the T1 group lasted significantly longer than those of the T2 and CK groups, and this highlights the superiority of the membrane-covered approach for elevating and preserving the composting temperature, which is consistent with the conclusion of Ma *et al.* (2017). During the initial 15 d, the temperatures of the T2 group also significantly surpassed those of the CK sample, and this difference was attributed to the utilization by microorganisms of the high P and trace elements from the superphosphate.

pH Variations

According to previous studies, the release of NH₃ during the composting process is strongly correlated with the pH of the compost (Li *et al.* 2008; Zhu *et al.* 2017). This is mainly because the accumulation of ammonium nitrogen and the increase in the concentration of NH₃ in a compost elevates the associated pH value. Concurrently, the high pH value enhances the release of NH₃ to the environment.



Fig. 3. Plots showing changes in the (a) pH and (b) EC value during the composting of three types of samples

As shown in Fig. 3a, the pH values for the types of samples initially increased and subsequently decreased. During the experiments, the T1, T2, and CK samples attained maximum pH values of 8.7, 8.4, and 6.9, respectively. The production of NH₃ and the degradation of acids mainly accounted for the increasing pH during the early stage for all sample types (Jia *et al.* 2016). Most of the NH₄⁺-N was converted by nitrification, while short chain organic acids produced from the degradation of organic matter likely decreased the pH value of the samples during a later stage (Zhou *et al.* 2019). The higher pH value of the T1 sample relative to that of the CK sample might have been because the water film on the inner side of the membrane dissolved part of NH₃, which dropped back into the compost pile. The NH₃ became alkaline after dissolving in water, which might have led to the increased pH value. Among the three types of samples, the T2 displayed the lowest pH value because the superphosphate was acidic, and its pH value was about 3.0. Nevertheless, the final pH value of the T1, T2, and CK samples were 8.11, 8.03, and 6.48, respectively, and these were consistent with the pH values required for organic fertilizers.

EC Changes

The electrical conductivity (EC) value reflects the salinity of soluble components from the compost, and a high salinity can damage plants through osmotic pressure (Huang et al. 2004). The changes in the EC value during the experiments for the three types of samples are exhibited in Fig. 3b. The EC value for the T1 and CK samples increased during the initial 9 d and decreased subsequently. The release of soluble mineral salts, such as ammonium ions and phosphate, caused by the rapid mineralization of organic matter might be the reason the EC value increased in the initial stage. Then, due to the production of NH₃ and the precipitation of mineral salts, the EC value decreased gradually and stabilized in the following time. The maximum EC values for the T1 and CK samples were 4.12 and 4.04 mS cm⁻¹, respectively, while the final values were correspondingly 3.22 and 3.38 mS cm⁻¹. The significantly higher final EC value for the T2 sample relative to those for the T1 and CK samples was mainly linked to the calcium dihydrogen phosphate, free phosphoric acid, and sulfuric acid in the superphosphate. Calcium dihydrogen phosphate contained inorganic salts, and the presence of free phosphoric acid and sulfuric acid accelerated the dissolution of inorganic salts, which led to an increased EC value. In fact, the final EC value of 5.67 mS cm⁻¹ for the T2 sample exceeded the safety threshold for plants of 4.0 mS cm⁻¹ (Li et al. 2007). This may have been the reason for the low GI value in T2. According to the Fig. 3b, the EC value showed a downward trend at the end of the composting period, which might be caused by the biodegradation of water-soluble substances, such as organic acids, during this period (Hosseini and Aziz 2013).

GI Characterization

The GI is a vital parameter associated with the aerobic composting process. It can be employed to evaluate the stability and risk of products from composting (Zucconi 1981). If the GI of a composting product attains 50%, the product is characterized as phytotoxicity-free, whereas if the GI exceeds 80%, the product is considered mature (Saidi *et al.* 2009). The changes in the GI of the three types of samples are shown in Fig. 4.



Fig. 4. Plot showing changes in the GI during composting of three types of samples

The GI of the T1 and CK samples decreased slightly during the initial stage, and then they increased gradually until the end of the respective composting experiments. The decrease in the GI during the initial stage was likely because of rapid degradation of organic matter, which probably generated substances that were unfavorable for the germination of the seeds. The final GI of the T1 and CK samples were correspondingly 93.1% and 83.2%, and both values satisfied the maturity requirement. However, compared with the CK sample, the T1 sample attained a GI of 50% and 80% approximately 2 and 9 d earlier, respectively.

These results indicate that the membrane-covered technology can enhance the speed of the composting process, and thus shorten the composting cycle. The increased speed was linked to the temperature enhancement of the compost using the membrane-covered technology, which accelerated the degradation of organic matter. Although the GI of the T2 sample increased continuously, these values were lower compared to those of the T1 and CK sample during an identical period. The final GI of 74.7% for the T2 sample was probably because of the free acids and high salinity from the superphosphate (Jiang *et al.* 2014; Yang *et al.* 2015). According to the trends in Fig. 3, a longer composting time is probably required to enhance the GI of the T2 sample. However, Yang *et al.* (2015) reported that although the superphosphate can decrease the degradation of organic matter in compost, no adverse effect on the maturity and GI of the compost products was observed. These observations were probably because the study involved a longer composting time.

Nitrogen-containing Gases Emissions from the Aerobic Composting

NH3 emission characterization

The emission of NH₃ accounts for more than 75% of the total N loss during aerobic composting (Jiang *et al.* 2013). The NH₃ emission rates and cumulative emissions for the three groups are shown in Fig. 5. Evidently, the production and emission of NH₃ are strongly correlated with the composting temperature (p < 0.001). The three groups exhibited sharp rises, followed by gradual declines. The NH₃ emissions for all groups in the present study occurred mainly in the initial 18 d of composting. This was associated with the rapid temperature increase, the high-temperature fermentation, and the rapid mineralization of organic N in the compost during this stage of the experiments. Then due to the depletion of organic matter and the reduction of ammonification bacteria activity, NH₃ emission decreased in the subsequent time.

Fukumoto *et al.* (2011) indicated that the NH₃ emitted during aerobic composting mainly associated with the thermophilic phase, followed by a gradual decline in the next stage, which was consistent with the results obtained in the present study. The total NH₃ emitted from the T1 was 25.8% less than that from the CK group, and this indicated that the membrane-covered approach reduced NH₃ emission during anaerobic composting. This decrease was attributed to the condensed water vapor on the internal surface of the membrane, which allows dissolved NH₃ to return to the compost because of the force of gravity (Ma *et al.* 2017, 2021). The total NH₃ emitted from the T2 was 21.3% less than that of the CK group, and this was mainly because the superphosphate utilized decreased the pH value of the compost pile, and this acidic environment then reduced the emission of NH₃.



Fig. 5. Plots displaying the emission rate and the cumulative emission of NH_3 during composting of three groups

N₂O Emission Characterization

The chemical compound N₂O is an atmospheric pollutant that damages the ozonosphere and enhances the greenhouse effect, and its warming potential is 298 times higher than that of CO₂. The production of N₂O in compost is mainly associated with the nitrification of NH₄⁺-N and the incomplete denitrification of NO_x⁻-N (Huang *et al.* 2004). The N₂O emission rates and cumulative emissions for the three groups are shown in Fig. 6.

The trends of a rapid rise followed a gradual decrease displayed by the three groups were broadly consistent with those for the emission of NH₃. The N₂O emissions for all groups in the present study mainly occurred in the initial 12 d of composting. This observation was probably because of the rapid oxidation of organic N and NH₄⁺ promoting the emission of N₂O, which is consistent with the conclusion of Stange and Neue (2009). However, some studies have pointed out that neither nitrifier nor denitrifier can adapt to the high-temperature environment of compost, so N₂O could not be produced in the thermophilic phases. According to some previous studies (Bian *et al.* 2017; Ma *et al.* 2021), the maximum N₂O emission rates were observed during the intermediate stage of composting when the temperature had decreased. The NO₂⁻ accumulation was considered the main reason for this phenomenon. The total N₂O emissions of the T1 and T2 groups were 13.1% and 27.2%, respectively, less than that of the CK group. This indicated that the membrane-covered approach can reduce the emission of N₂O, but the performance was inferior to that of the 5% superphosphate addition.



Fig. 6. Plot showing the emission rates and cumulative emissions of N₂O during composting of the three groups

CONCLUSIONS

- 1. The present study demonstrated that the membrane-covered technology improved the temperature and the germination index (GI) value of compost. Conversely, the application of 5% superphosphate in the process might have an adverse effect on the GI value.
- 2. Membrane-covered technology accelerated the composting process. The product was characterized as phytotoxicity-free and mature approximately 2 and 9 d earlier, respectively, compared with the control (CK) group samples.
- 3. Both the membrane-covered and 5% superphosphate application reduced emissions of NH₃ and N₂O during the composting process. However, the membrane-covered technology exhibited superiority for the reduction of NH₃ emission, while the superphosphate application was better for the reduction of N₂O emission. Compared with the CK group samples, the membrane-covered technology reduced NH₃ and N₂O correspondingly by 25.8% and 13.1%, while the 5% superphosphate application decreased the NH₃ and N₂O emissions by 21.3% and 27.2%, respectively.

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