

# Synthesis of Silica from Rice Husk as Coating Material on Magnetic Nanoparticle for Efficient Adsorption of Phenol from Water Samples

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Silica (SiO<sub>2</sub>) from rice husk was coated with magnetic nanoparticles (MNPs) as an adsorbent to adsorb phenol from river water. The structure of SiO<sub>2</sub> and SiO<sub>2</sub>-MNPs were characterized by Fourier transform infrared spectroscopy (FTIR) and X-ray diffraction (XRD). Field emission scanning electron microscopy (FESEM) showed a rod shape, with a rough surface area in the range of 2 to 3 μm. Transmission electron microscopy (TEM) and energy dispersive x-ray spectroscopy (EDX) were used to examine the resulting spherical shape of the synthesized SiO<sub>2</sub>-MNP. The results showed a range of 1.3 to 1.5 nm for SiO<sub>2</sub> and 4.2 to 6.4 nm for SiO<sub>2</sub>-MNP. Vibrating-sample magnetometer (VSM) showed an Fe value of 45.1% in SiO<sub>2</sub>-MNP (VSM); for MNP, SiO<sub>2</sub>, SiO<sub>2</sub>-MNP 104.12, 4.72, and 8.01 emu/g, respectively. Response surface methodology (RSM) was used to study the parameters and responses to obtain an optimized condition in SiO<sub>2</sub>-MNPs usage. The optimized parameters were extraction time selected at 5 min, pH 8, 8 mL acetonitrile as solvent and 15 min as sonication time. The application of SiO<sub>2</sub>-MNPs was applied to real water samples, with recovery of 84% of phenol. Thus, the synthesized adsorbent, SiO<sub>2</sub>-MNPs, was developed successfully for phenol removal from water samples.

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## INTRODUCTION

Phenol is a common environmental pollutant because it is used in many industries (Norseyrihan *et al.* 2016; Hazrina *et al.* 2018). The waste produced by industries becomes a challenge for pollutants removal and disposal process, thus requiring further removal steps, which involve high costs. Contamination of water sources with endocrine-disrupting organic micropollutants, such as phenolic compounds, affects aquatic life and human health (Ahmed *et al.* 2010). The phenolic group has the highest pollutant toxicity, causing severe, chronic harm even at low concentrations (Mecha *et al.* 2016; Frasetya *et al.* 2019). Therefore, the separation or extraction of wastewater containing phenol is essential.

Sustainable and green chemistry could be used to convert agro-industrial solid wastes into valuable products. This is aligned with global plans to reduce waste and

environmental contamination. For example, the by-products of sugarcane bagasse, rice husk, and straw have a high silicon content. Rice husk and ash are used frequently as renewable sources to produce value-added materials, such as silica magnetic nanoparticles ( $\text{SiO}_2\text{-MNP}$ ).

Rice husk contains materials as the main constituent, and the ash comprises 87% to 98%, including a small percentage of metallic elements. It is a by-product of rice milling and is used for poultry, brick industry fuel, planting medium, and compost. According to Ariffin *et al.* (2019), the silicone content of husk is 18.8% to 22.3%, and the silicone content of the straw is 9.30% (Mecha *et al.* 2016).

Due to their high surface area, increased magnetic capabilities, and low cost, magnetic nanoparticles (MNPs) synthesized from ferrous compounds ( $\text{Fe}_3\text{O}_4$ ) are well suited for use as a magnetic adsorbent (Zhang and Shi. 2012; Ul-Islam *et al.* 2017). However, it has drawbacks that limit its use for phenol extraction, including the ease with which it oxidizes when exposed to air, agglomerates in aqueous media, is unstable in an acidic environment, and is very hydrophilic (Jones and Clarke 2005). Finding a solution for these issues, such as materials to functionalize the surface of MNPs, is imperative to overcoming these limitations.

The effectiveness of the phenol extraction method in water samples has been demonstrated in numerous investigations. There are many complicated techniques, including liquid-liquid extraction (LLE), solid phase extraction (SPE), micro-solid phase extraction (MSPE), and supercritical fluid extraction (SFE) (Labat *et al.* 2000; Lee *et al.* 2005; Lee *et al.* 2006; Dan *et al.* 2022). However, these approaches have several drawbacks, including their time requirement, intricate procedures, heavy reliance on organic solvents, and low sensitivity.

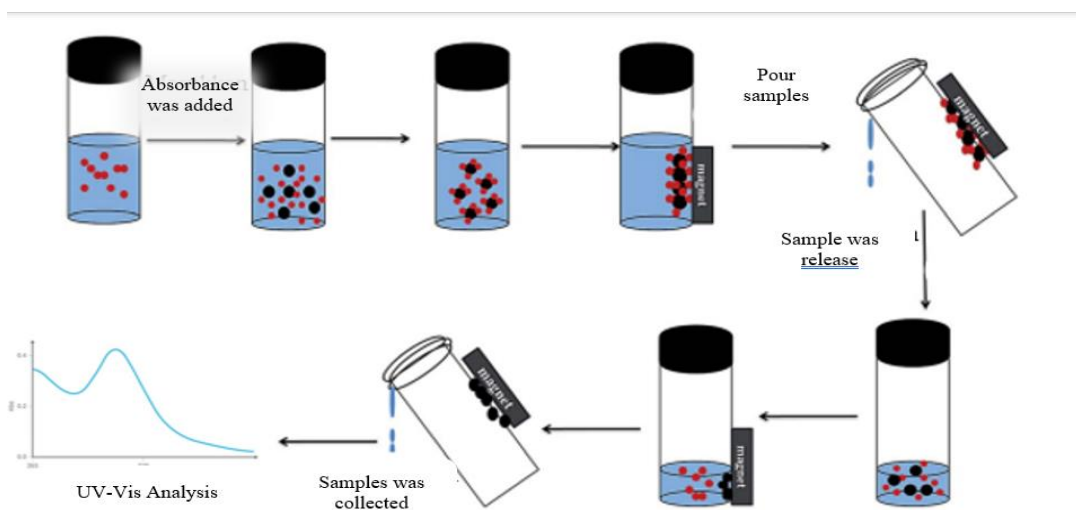


Fig. 1. MSPE method

The methodology used in this study is known for removing selected analytes from aqueous media based on magnetic adsorbents (MNPs) (Begum *et al.* 2021). The adsorbent is not tightly wrapped in the cartridge. It can be placed directly in a large volume of water sample solution or suspension, and the extraction time is shortened. Other benefits of MSPE include the use of low-volume, non-toxic organic solvents, good repeatability, quick sample processing, and compatibility with most analytical instruments (Noushad *et al.*

2012; Rafiee *et al.* 2012; Boudesocque *et al.* 2019; Ram Talib *et al.* 2019). Figure 1 summarizes the MSPE approach.

In addition, the response surface methodology (RSM) was used to determine the concentration of phenol in water using the optimum condition of parameters studied. RSM is a mathematical and statistical method that is widely used in industry to determine the optimal conditions in the experiment. It is helpful for maximizing or minimizing various variables because it evaluates the effect of multiple factors and their respective interactions on one or more variables moving silently. It also reduces the impact of noise to improve the convergence of the optimization process (Boudesocque *et al.* 2019).

This study established the viability of using rice husk silica as a coating material for MNPs ( $\text{SiO}_2$ -MNP) to extract phenols from river water. When compared to other paddy plants debris, the rice husk has the highest content of silica. Fourier transform infrared spectroscopy (FTIR), X-ray powder diffraction (XRD), field emission scanning electronic microscopy (FESEM), transmission electron microscope (TEM), energy dispersive X-ray analysis (EDX), and vibrating sample magnetometer (VSM) studies were used to look at the morphology, structure, and composition of  $\text{SiO}_2$ -MNP. The findings indicate that silica has been successfully coated on the MNPs core-shell. The effects of different experimental parameters, including sample volume, extraction time, desorption time, pH value, amounts of sorbent, and kinds and amount of desorption solvents, were examined.

The phenol recovery in water samples (84.8% to 92.4%) is considered high with simple and quick steps without the usage of the complex apparatus. At the same time, the results on water samples without phenol were detected in the concentration range of 3.82 ppm to 7.58 ppm.

## EXPERIMENTAL

### Reagents

RandM Chemicals Company provided the ferric (II) chloride tetrahydrate ( $\text{FeCl}_2 \cdot 4\text{H}_2\text{O}$ ), and ferric (III) chloride hexahydrate ( $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ ) (Essex, UK). Phenol was provided by Merck (Darmstadt, Germany). Acetonitrile was obtained from J.T. Baker (Philipsburg, New Jersey). Methanol and ethanol from HmbG Company (Hamburg, Germany). Stocks of the standard analytical solution were created using acetonitrile at concentration of 1000 g/mL and kept at 4 °C. Prior to usage, acetonitrile was brought to room temperature. Sodium hydroxide and hydrochloric acid were provided by Merck (Germany) and were used to regulate the pH of the solution throughout experiment. Stock solutions were thoroughly diluted with acetonitrile or Milli-Q ultrapure water to create working solutions.

### Instrumentation

FT-IR Spectrometer (Thermo Nicolet, Nexus 670) by ATR technique in the absorption mode with 32 scans and a resolution of  $\pm 4 \text{ cm}^{-1}$ , and a wavenumber range of 4000 to  $450 \text{ cm}^{-1}$  with diamond as a detector was used to confirm the functional group's as-synthesized nanoparticles. Using a Bruker D8 Advance X-ray powder diffraction (XRD) diffractometer (CuK, radiation,  $\alpha$  at 40 kV/30mA), at a scanning speed of  $1^\circ/\text{min}$  and  $0.02^\circ/\text{min}$  from  $10^\circ$  to  $80^\circ$  ( $2\theta$ ), it was determined how the produced nanoparticles behaved in terms of crystallinity. Field emission scanning electronic microscopy (FESEM), model VP (ZEISS SUPRA 55VP, evolution 0.2 nm@120 kV), outfitted with energy dispersive

X-ray spectrometry, was used for the elemental analysis. Talos L120C evolution 0.2 nm @ 120 kV transmission electron microscope (TEM) was utilized. The magnetic behaviour of the prepared nano-adsorbents was characterized by using a vibrating sample magnetometer (VSM Lake Shore 7404 Series, Tokyo, Japan). Data were recorded with 80 points/loop with speed of 10s/drop. The separation and quantification of the tested phenol was achieved by using an UV-Vis analysis.

### Preparation of Silica (SiO<sub>2</sub>) from Rice Husk

The main component was rice husk, and examples came from Tanjung Karang Bernas Company in Kuala Selangor. To remove sand and dirt stuck to the surface of the rice husk, it was first scrubbed and washed with distilled water (Rafiee *et al.* 2012). The samples took 10 min to dry in the oven. After that, 1.0 M of hydrochloric acid (HCl) was added, and the process to remove metal impurities was continued by soaking at 90 °C for 1 h in water bath. Prior to filtering and overnight oven drying, samples were washed with distilled water (Chakraverty and Kaleemullah 1991; Noushad *et al.* 2012). In addition, this technique was undertaken to boost the purity of silica (Norsuraya *et al.* 2016).

The dried rice husk was steeped in 1.0 M NaOH to form a sodium silicate solution. The silica component in the rice husk dissolved as a result of the reaction between the rice husk and NaOH, creating a sodium silicate solution (Patil *et al.* 2014). The solution was immersed in water bath for 1 h while being agitated with stirrer. The colour of the solution altered after 1 h and took on a blackish hue as a result of the sodium silicate reaction.

Whatman filter paper No. 41 was used to filter the solution, separating the rice husk residue from the dark-coloured solution. The silica supernatant solution was collected. HCl was drip-fed while swirling on a heated plate until pH 7 to 8 was achieved. After 45 min of stirring, the solution was allowed to mature for around 2 weeks. The precipitate was centrifuged, and the precipitate collected was calcined for 4 h at 800 °C. A white silica powder was produced. A white silica powder was produced and stored for use in the procedure that followed. The approximate yields of white silica powder produced was 0.7 gram.

### Preparation of SiO<sub>2</sub> Coated with Magnetic Nanoparticle (MNPs)

First, 450 mL of deionized water was used to dilute 0.7 g of extracted SiO<sub>2</sub> powder. In the meantime, a 2:1 aqueous solution of ferric chloride (Fe<sup>3+</sup>) and ferrous chloride (Fe<sup>2+</sup>) were made. The solution was gradually incorporated into a room temperature SiO<sub>2</sub>. The pH of the solution was raised to 10 by slowly adding ammonia solution. The solution was continuously stirred while being heated at 90 °C for 4 h. The solution was filtered after being cooled to room temperature. The dark precipitate was repeatedly washed with distilled water and acetonitrile. The precipitate was dried overnight at 70 °C.

### Extraction Procedure

SiO<sub>2</sub>-MNPs of 0.1 g was placed in a vial, and 8 mL of prepared spiked phenol solution (pH 8) was transferred into the same vial. The solution was shaken for 10 min. The adsorbent was isolated from the sample solution using an external magnet, and then the sample solution was decanted. The adsorbed phenol on SiO<sub>2</sub>-MNP was eluted by sonication method with 8.0 mL of desorption solvent (acetonitrile) for 15 min. The eluent was collected and stored in a vial for UV-Vis analysis.

## Optimization of MSPE

Several factors that affect extraction efficiency, such as solution pH value, adsorbent amount, types of desorption solvent, volume of desorption solvent, extraction time, desorption time and sample volume, were studied.

## Samples

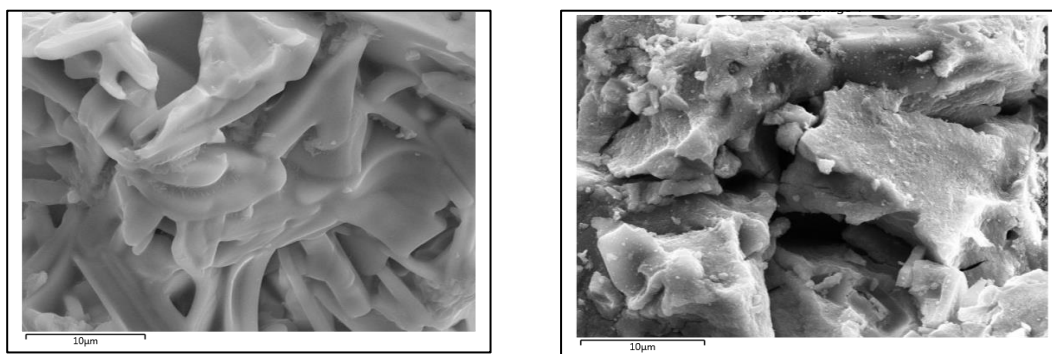
River water and lake water were selected as real water samples for investigation. River water samples were collected from Selangor, Malaysia. The specific river water locations were Sungai Kuyoh, Sungai Buah, Sungai Kantan, and Sungai Langat (in the area of Universiti Kebangsaan (UKM)). Lake waters were collected from Selangor, Malaysia. The specific location of lake water was collected from Taman Idaman, Tasik KBG (FRIM), Taman Tasik Cempaka, Tasik Sri Serdang, Tasik Faculty of Engineering (FKAB, UKM), and Tasik Bandar Tun Hussein Onn. All water samples were filtered through a 0.25  $\mu\text{m}$  membrane and stored at 4  $^{\circ}\text{C}$ .

## RESULTS AND DISCUSSION

### Characterization of $\text{SiO}_2$ and $\text{SiO}_2$ -MNP

#### Morphology analysis

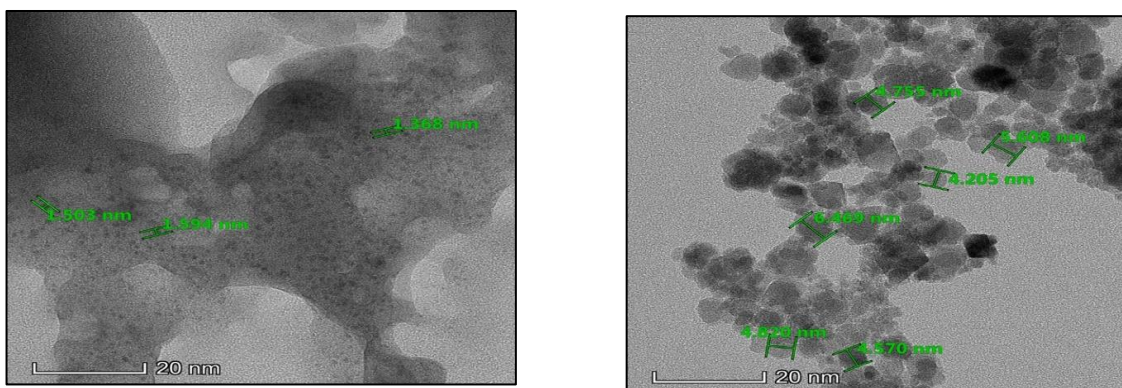
Figure 2 shows the particle size of  $\text{SiO}_2$  and  $\text{SiO}_2$ -MNP.  $\text{SiO}_2$  was depicted as having a rod-like shape and  $\text{SiO}_2$ -MNP as having an erratic arrangement. The  $\text{SiO}_2$  shown here was rice-husk silica. The samples were irregular in organization and ranged in size 3 nm to 10 nm. According to earlier research, the solvent utilized significantly impacted the molecules' size and arrangement (Ariffin *et al.* 2019). The size of the magnetic silica nanoparticles increases with the length of the alcohol chain, resulting in a rougher and bigger surface on the sample, as seen in Fig. 2(b). Blobs result from the high surface area and the resulting high surface energy area.



**Fig. 2.** Analysis FESEM of (a) Silica (rice-husk silica), (b) Silica magnetic nanoparticle (silica from rice-husk)

The outcomes of the study using transmission electron microscopy (TEM) are shown in Fig. 3. The structure of  $\text{SiO}_2$ -MNP was spherical, with  $\text{SiO}_2$  covering the exterior of  $\text{Fe}_3\text{O}_4$  nanoparticles and  $\text{Fe}_3\text{O}_4$  nanoparticles inside (Lee *et al.* 2018). These findings demonstrated that  $\text{SiO}_2$ -MNPs were spherical, contained  $\text{Fe}_3\text{O}_4$  nanoparticles, and had a  $\text{SiO}_2$  covering shell around them. In addition, two independent electron density areas

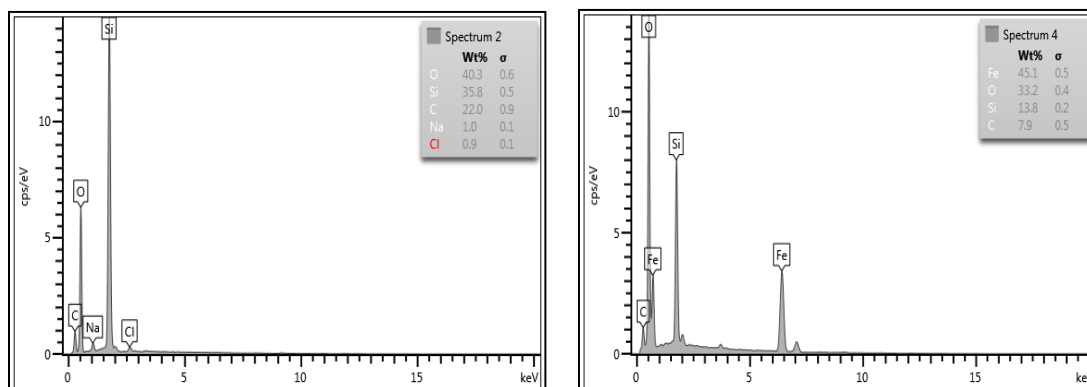
confirmed the production of silica-coated nanoparticles. The magnetic core was represented by the electron-dense region, while the silica coating was represented by the less dense translucent zone surrounding the core (Helmi Rashid Farimani *et al.* 2014).



**Fig. 3.** Analysis TEM of (a) Silica (rice-husk silica), (b) Silica magnetic nanoparticle (silica from rice-husk)

#### Elemental study

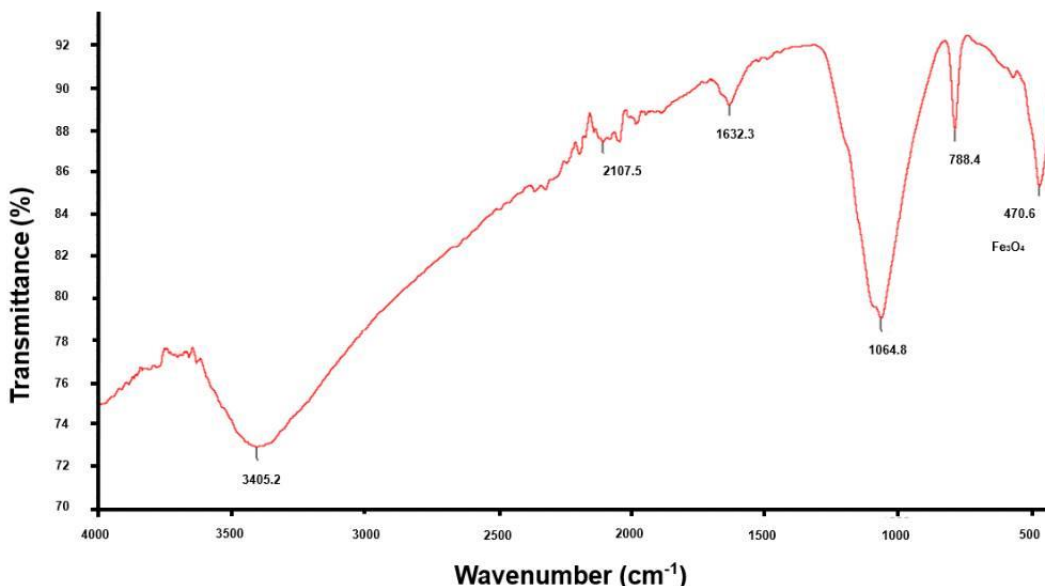
To identify the composition of components including O, Fe, Si, and C in prepared MNPs,  $\text{SiO}_2$  and  $\text{SiO}_2$ -MNPs, energy dispersive X-ray analysis (EDX) was carried out. According to Fig. 4(a) samples of MNPs, only 35.8% of Si, 40.3% of O, and 22.0% of C were presented in the MNPs. However, following coating with  $\text{SiO}_2$ , new components were presented, including 45.1% of Fe, 33.2% of O, 7.9% of C, and 13.8% of Si. The large percentages of C (22.0%), Si (2.46%) and O (40.3%) in  $\text{SiO}_2$ -MNP demonstrated that MNP was successfully introduced onto  $\text{SiO}_2$ -MNP surface, as shown in Fig. 4(b).



**Fig. 4.** Analysis EDX of (a) Silica (rice-husk silica), (b) Silica magnetic nanoparticle (silica from rice-husk)

#### Functional group analysis

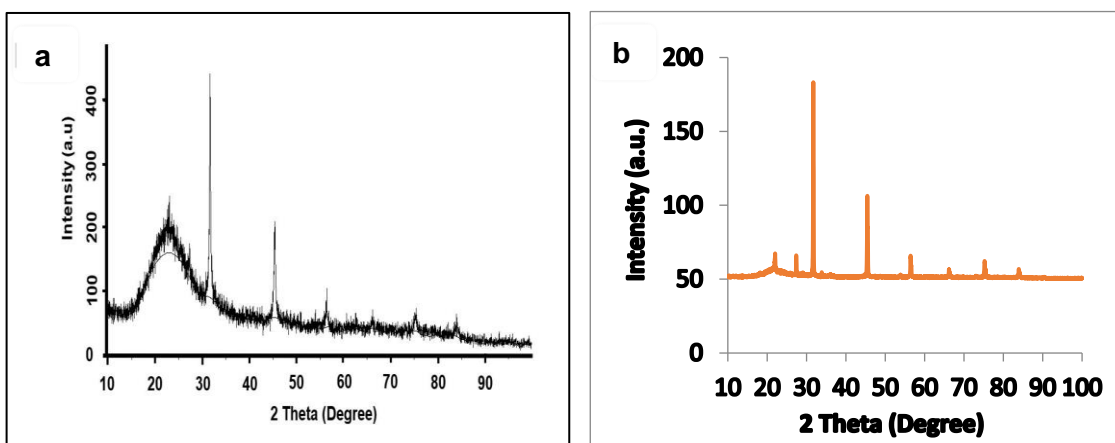
Fourier Transform Infrared Spectroscopy (FTIR) was used to characterize the surface chemical structures of  $\text{SiO}_2$ -MNP. The results are depicted in Fig. 5. The appearance of additional peaks at  $3405.2\text{ cm}^{-1}$ ,  $1064.8\text{ cm}^{-1}$ , and  $470.6\text{ cm}^{-1}$  were attributed to O-H stretching vibration, anti-symmetric Si-O-Si, symmetric Si-O-Si, and  $\text{Fe}_3\text{O}_4$  stretching, respectively. This suggests the linker  $\text{SiO}_2$  was added to the surface of MNP. These outcomes demonstrated the functionalization of MNP onto  $\text{SiO}_2$ -MNP surface.



**Fig. 5.** Analysis FTIR of silica magnetic nanoparticle (silica from rice-husk)

#### Crystallinity analysis

Figure 6 displays the XRD patterns of  $\text{SiO}_2$  and  $\text{SiO}_2$ -MNPs. Broad spectral curves with peak values between  $2\theta = 15^\circ$  up to  $30^\circ$  are known as amorphous forms (Park and Lee, 2004; Ghorbani *et al.* 2015). Figure 6(b) show peaks  $2\theta = 27.44^\circ$ ,  $31.71^\circ$ ,  $45.42^\circ$ , and  $56.41^\circ$  are the resulting nanoparticle diffraction angles. These results demonstrated the creation of highly pure crystals (Gopal *et al.* 2020; Nodeh *et al.* 2015).



**Fig. 6.** Analysis XRD of (a) Silica (rice-husk silica), (b) Silica magnetic nanoparticle (silica from rice-husk)

#### Magnetic behavior

Vibrating sample magnetometry (VSM) analysis was carried out to determine the concentration of  $\text{SiO}_2$  and  $\text{SiO}_2$ -MNPs. The magnetic behaviour of MNPs,  $\text{SiO}_2$  and  $\text{SiO}_2$ -MNP.  $\text{SiO}_2$  and  $\text{SiO}_2$ -MNP were found to be 104.1, 4.718, and 8.008 emu/g, respectively. Figure 7 illustrates the superparamagnetic behaviour of the different sorbents. The absence of hysteresis, low remanence, and coercivity demonstrates it.  $\text{SiO}_2$  coating on the outer

layer of MNP surface, which reduced interaction between the nanoparticles was evidence by the decreased magnetic value of SiO<sub>2</sub>-MNPs as compared to MNP.

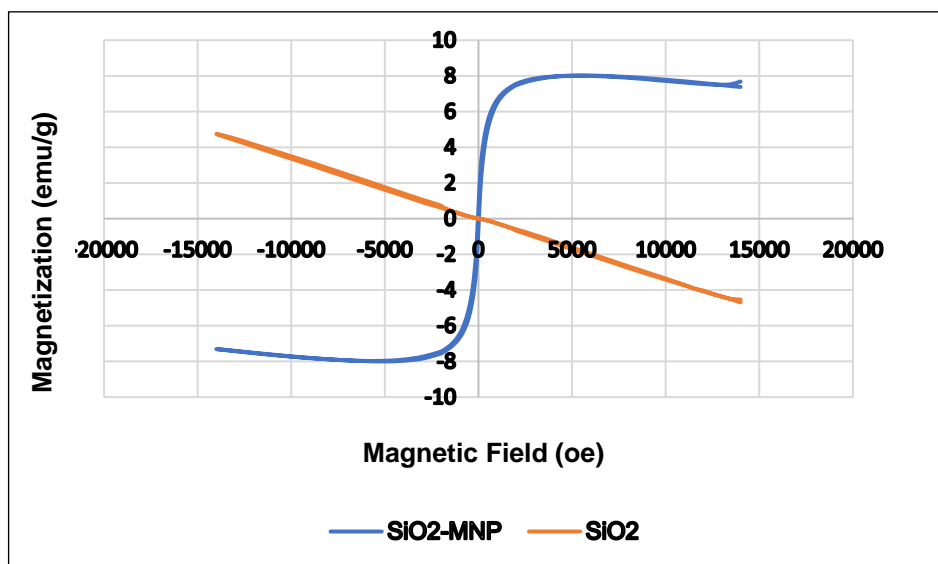


Fig. 7. Analysis VSM of (a) Silica, (b) magnetic nanoparticle, and (c) Silica magnetic nanoparticle

### Optimization of MSPE conditions

To find ideal conditions for SiO<sub>2</sub>-MNPs through the MSPE technique, key experimental parameters including pH value, amounts of sorbent, types of solvent, types of desorption solvent, extraction, and desorption time, as well as sample volume were optimized in this study.

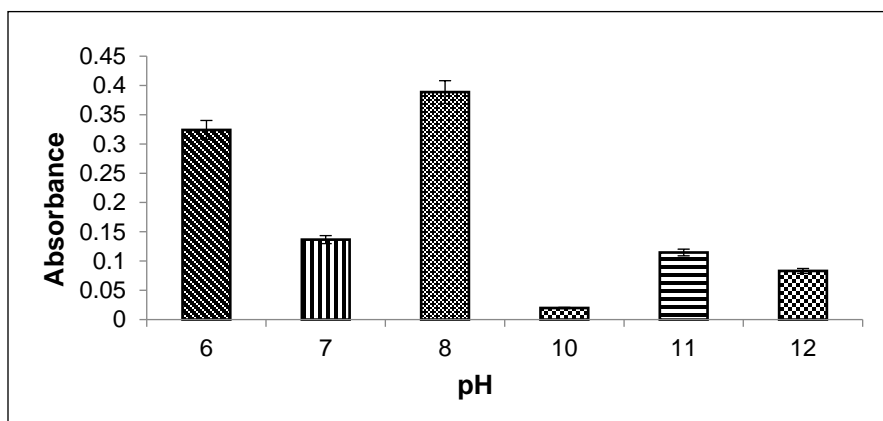


Fig. 8. Effect of pH on phenol extraction

#### *Effect of sample solution pH*

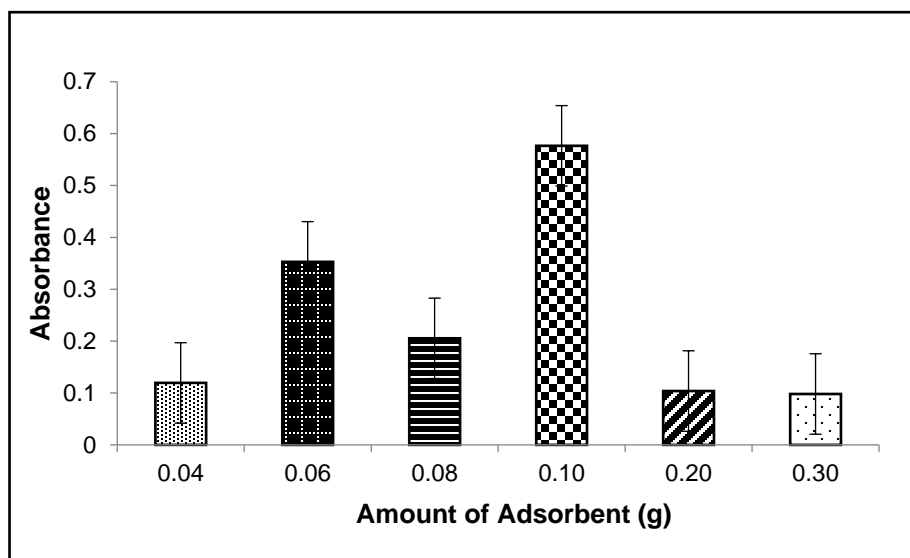
The impact of sample solution was examined from pH 6 to 12, as shown in Fig. 8. Low recovery rates were seen regardless of the pH. Thus, SiO<sub>2</sub>-MNPs charge density and phenol charge state were considered. In terms of adsorbent charge, the chosen phenol produced a neutral molecular state under acidic circumstances. Phenol extraction at pH 8



displayed the most significant peak area, so pH 8 was selected as the best pH value (Zhang *et al.* 2013). When the pH is less than 5, the solution is acidic, and the metal-organic framework may not be able to support the sorption (Wang *et al.* 2014). Phenols are more likely to ionize at high pH. As a result of the increasing quantity of charges, the absorption force weakens (Gamonchuang and Burakham 2021). Therefore, using sample solutions of 0.1 M HCl and 0.1 M NaOH, the impact of pH on extraction performances was investigated in the pH range of 6 to 12. From pH 6 to 7, the adsorption declined, and it grew more at pH 8. It started to deteriorate when the value was further raised to pH 12. This pH change may be closely correlated with the surface charge of the phenolic compound, whereby phenol is a weak acid and tend to exist in its protonated state (Gopal *et al.* 2020).

#### *Effect of sorbent amounts*

The amount of sorbent used limits the amount of phenol that can be extracted before an equilibrium is reached. According to Nodeh *et al.* (2015), an increase in the adsorbent mass results in an increase in adsorption efficiency. This is because an increase in adsorbent will result in a rise in adsorption sites, which would allow for the occupancy of more analytes. In this experiment, various dosages of the SiO<sub>2</sub>-MNP sorbents, ranging from 0.04 to 0.3 g, were applied. Figure 9 shows that when the amount of sorbent grew, the peak regions climbed sequentially from 0.04 g to 0.1 g, and the recovery plateaued after 0.1 g due to the active sites on the adsorbent SiO<sub>2</sub>-MNP being saturated, which was similar to results by Dolaksiz *et al.* (2018). Therefore, the ideal quantity of adsorbents used in subsequent tests was 0.1 g SiO<sub>2</sub>-MNP.

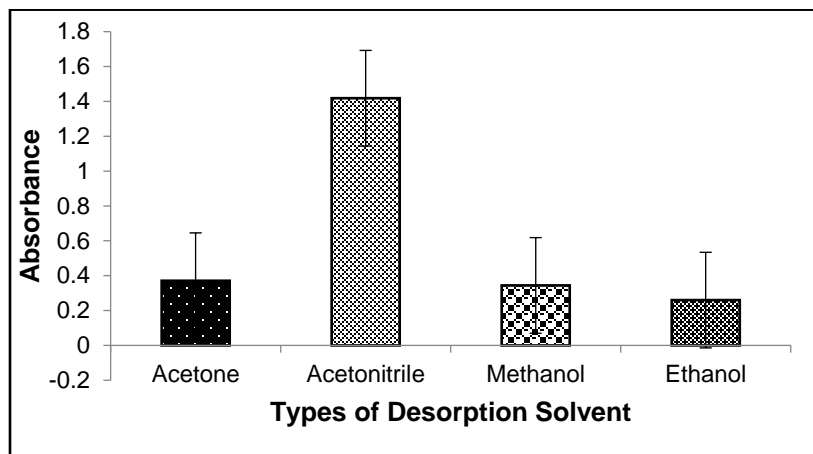


**Fig. 9.** Effect on amount adsorbent of phenol extraction

A similar study by Gopal *et al.* (2020) stated that dosage of adsorbent was a vital factor related to the extraction capacity. An increase in adsorbent dosage would increase the active binding site of magnetic to interact toward phenolic compound, eventually enhancing the extraction efficiency. Therefore, an increasing amount of adsorbent dosage resulted in saturation and aggregation of the active binding site, which resulted in further adsorption and caused the extraction efficiency to remain constant (Gopal *et al.* 2020).

*Effect of desorption solvent*

Acetone, acetonitrile, methanol, and ethanol were employed as desorption solvents. Acetonitrile yielded better recoveries for nearly all the targeted phenols and was the best eluting solvent, as demonstrated in Fig. 10. When employed as an extraction solvent, acetonitrile is an aprotic polar solvent that is unlikely to react. Acetonitrile is an aprotic polar solvent that might break the hydrogen chain between phenol and SiO<sub>2</sub>-MNP, following the earlier work. Additional investigations showed the ability to solubilize acetonitrile as a spray adsorbent solution (Van Berkel and Kertesz 2007). Therefore, acetonitrile was chosen for subsequent experiments.



**Fig. 10.** Effect of desorption solvent on phenol extraction

**Application to Real Water Sample**

The proposed method was applied to determine the targeted phenols in selected water samples with different water matrices (river water and lake water). Results ranged from 84.8% to 92.4%. Results from water samples that were unspiked with phenol showed that it could be detected at concentration between 3.82 ppm to 7.58 ppm. Table 1 examines the adsorption results from the study's use of UV-Vis spectroscopic analysis on lake and river water. Figure 11 displays the outcomes of UV-Vis phenol extraction on actual water samples. A UV-Vis spectrum from one of the outcomes from water samples is shown in Fig. 12.

**Table 1.** Phenol Analysis on River Water and Lake Water

Water Sample	Spiked Phenol	Unspiked Phenol
	%R	Concentration (ppm)
Tasik Idaman	90.96	4.52
Tasik KBG, FRIM	92.36	3.82
Taman Tasik Cempaka	90.2	4.90
Tasik Sri Serdang	90.1	4.95
Tasik FKAB	89.44	5.28
Tasik Bandar Tun Hussein Onn	92.04	3.98
Sungai Kuyoh	84.84	7.58
Sungai Buah	90.44	4.78
Sungai Kantan	87.34	6.33

A weight of 0.2 g SiO<sub>2</sub>-MNP with 4 mL volume of desorption solution produced high adsorption. In contrast, a volume content of 10 mL at the same adsorbent weight produced a lower adsorption result. This phenomenon suggested that the removal efficiency increased rapidly and significantly when more surface area and adsorption sites were used. It also remained stable to achieve saturation and a compatible balance (Mandal *et al.* 2019).

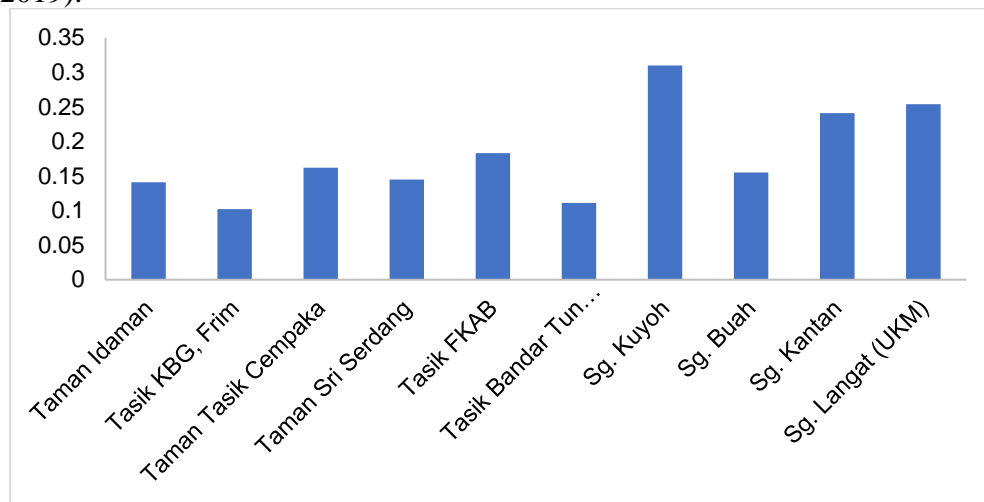


Fig. 11. Results of phenol extraction on real water samples using UV-Vis

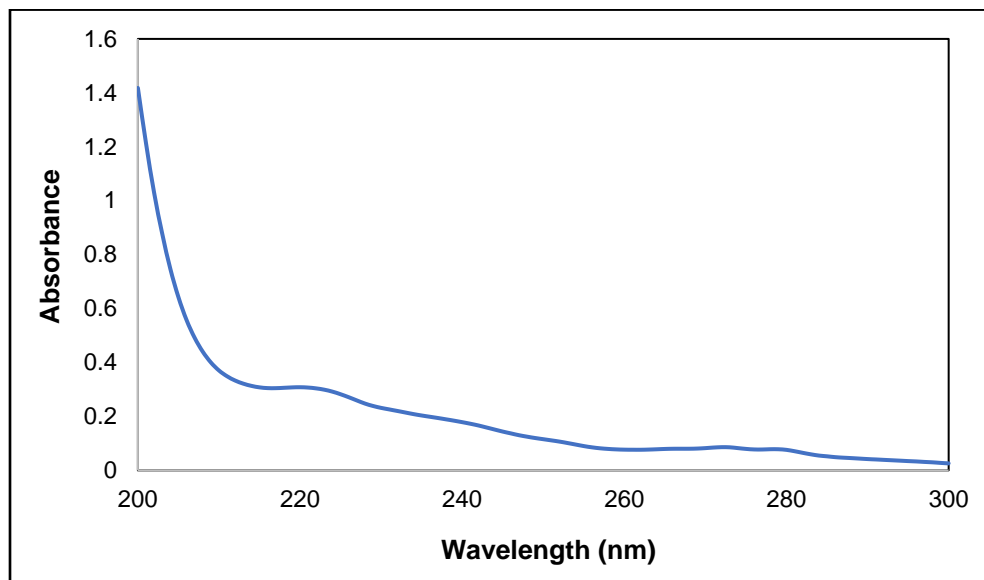


Fig. 12. UV-Vis spectrum on real water sample

Table 2 shows results from previous studies. There are many different techniques that can extract phenol from wastewater. The results of the current investigation demonstrated the superiority of the developed silica-based approach for phenol extraction from wastewater.

**Table 2.** Previous Studies on Phenolic Compound with Different Techniques

Analyte	Matric	Technique	pH and Volume of Samples	Type & Volume of Solvent	Type and Volume of Wash Solvent	Vortex & Sonication	Analysis Technique	Concentration	Reference
4-n-nonylphenol, bisphenol A, bisphenol AF	Drinking water	MSPE	25 mL	Methanol (3 mL)	Methanol	3 min	HPLC	0.08-0.21 ng.mL <sup>-1</sup>	Deng <i>et al.</i> (2019)
<i>p</i> -hydroxybenzoic acid <i>p</i> -coumaric acid	Fruit juice	MSPE	pH 7 5 mL	Ethylene glycol (20 mL)		10 min	HPLC	0.5-500 µg L <sup>-1</sup>	Soltani <i>et al.</i> (2018)
Chlorophenol	River water	SPE	2.5 (100 mL)	Acetone (2 mL) Water (2 mL)	Water (2 mL)		GC-ECD	0.0000005-0.0001	de Morais <i>et al.</i> (2012)
Phenolic group	Drinking water	MSPE	20 mL	Ethanol (6 mL) Water (24 mL)	Acetonitrile (40 µL)	1 min	HPLC	0.5-1000 µg L <sup>-1</sup>	Gamonchuang and Burakham (2021)
<i>Salicornia herbacea L.</i> , (SHL)	g-C <sub>3</sub> N <sub>4</sub> /Fe <sub>3</sub> O <sub>4</sub> (15 mg)	MSPE	Sample (5g) Water (100 mL)	Methanol/water/hydrochloric acid (1/1/0.2, v/v/v, 2x0.5 mL)	Methanol (1 mL) Water (1 mL)	5 min	HPLC	17.5-42.6 ng.mL <sup>-1</sup>	Yang <i>et al.</i> (2015)
Methylphenol	Water samples	CPE	pH 6 to 12	0.2 mL deionized water, 0.2 mL 15% additives GLDA and BMIMBr, 0.2 mL 15% surfactant non-additives and GLDA, 0.2 mL 20% surfactant aqueous for BMIMBr,		5 min	HPLC	LOD:LOQ; 0.05:0.18 ppm	Hazrina <i>et al.</i> (2018)
Phenol	Water samples	MSPE	pH 8	Acetonitrile (8 mL)	-	15 min	UV-Vis	(84.8% to 92.4%)	Current study

## Mechanism of Interaction between Silica Magnetic Nanoparticles towards Phenol in Water

A series of nanocomposites based on  $\text{Fe}_3\text{O}_4$  MNP obtained by co-precipitation and coated with  $\text{SiO}_2$  to form  $\text{SiO}_2$ -MNP were synthesized, as shown in Figs. 13 and 14. The target molecules interact with functional groups on the silica surface of the adsorbents to improve their efficiency, capacity, and selectivity. Due to their dual functionality, amino groups remove both cationic and anionic contaminants by electrostatic attraction. The amino group removes cationic contaminants such as heavy metals when the pH is higher (Hazrina *et al.* 2018; Asab *et al.* 2020).

By using a modified co-precipitation approach, magnetite ( $\text{Fe}_3\text{O}_4$ ), nanoparticles and silica coated magnetic nanoparticles ( $\text{SiO}_2$ -MNP) were created as adsorbents for phenol elimination in water. Magnetic nanoparticles were created to produce  $\text{SiO}_2$ -MNP by carefully hydrolyzing aqueous solutions of  $\text{FeCl}_2$  and  $\text{FeCl}_3$  with ammonium hydroxide (Fig. 13). Silica was attached to create the  $\text{SiO}_2$ -MNP molecule (Fig. 13).

Due to the low isoelectric point of silica surfaces, silanol groups ( $-\text{Si}-\text{O}-$ ) exist even if silanol groups ( $-\text{Si}-\text{OH}-$ ) are the prevalent type (Hozhabr and Entezari 2015). The expected H-bonds ( $-\text{Si}-\text{OH}$ ) are between the  $\text{SiO}_2$ -MNP and phenolic hydroxyl groups (Fig. 14). The expected H-bonds ( $-\text{Si}-\text{OH}$ ) are between the  $\text{SiO}_2$ -MNP and phenolic hydroxyl group (Fig. 14). The adsorption of phenol onto  $\text{SiO}_2$ -MNP involves hydrogen bonding interaction (H-bonding) through the MSPE technique (Ding *et al.* 2012). Phenol forms hydrogen bonds readily in solution (Norseyrihan *et al.* 2016). Additionally, this molecule organizes water-bridge dimers interactions and engaging assembly wherein all the H-bonding sites. Meanwhile,  $\text{SiO}_2$  is very polar because of the presence of the  $-\text{OH}$  groups that form hydrogen bonds. The presence of  $\pi-\pi$  bonds in silica magnetic nanoparticles indicates successful interaction between  $\pi$ -conjugated phenol and  $\text{SiO}_2$ -MNP.

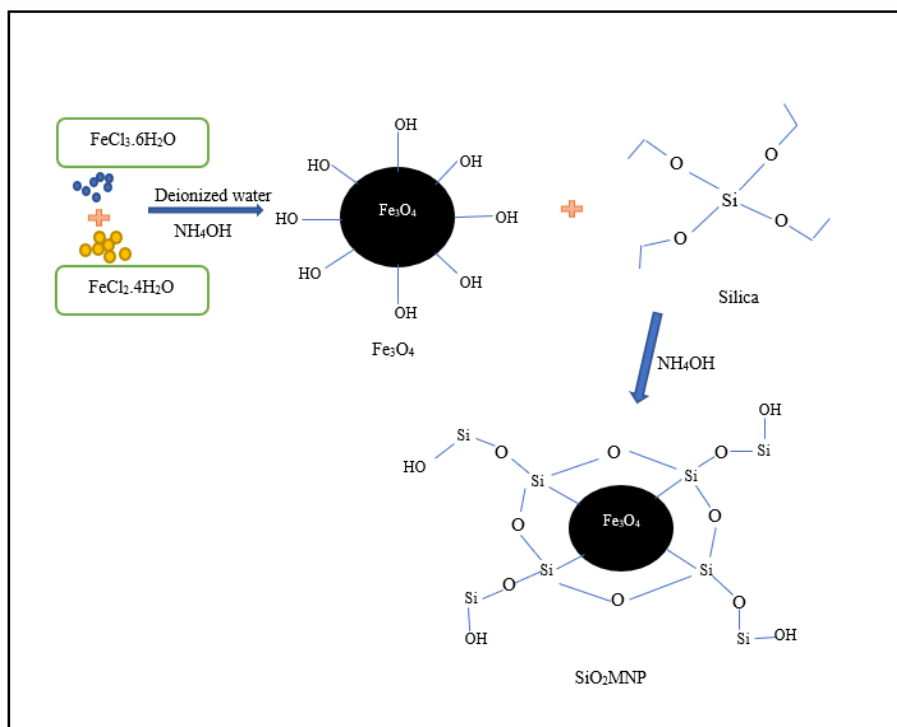


Fig. 13. Synthesis of  $\text{SiO}_2$ -MNP

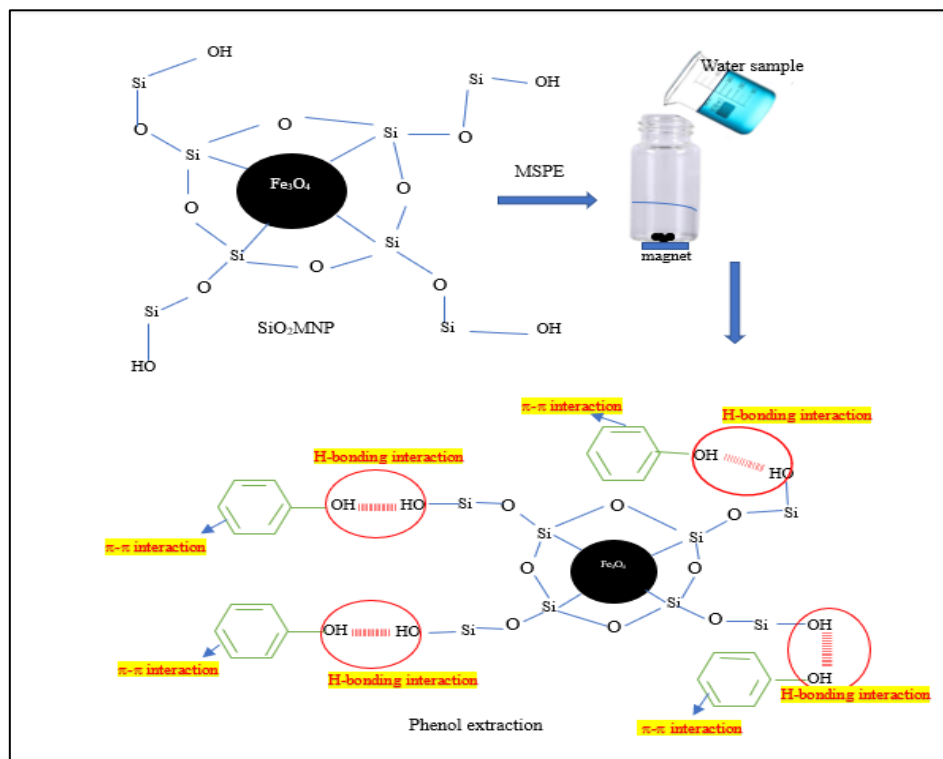


Fig. 14. Interaction of mechanism of phenol from water using SiO<sub>2</sub>-MNP

## CONCLUSIONS

1. The deposition approach can be used in this study to create silica that has been extracted from rice husk to recover phenol from water samples. In addition, magnetic nanoparticles functionalized with SiO<sub>2</sub>-magnetic nanoparticles (MNP) were also created. First, information regarding the sedimentation method of silica extraction from rice husk was gathered. Second, the micro-solid phase extraction (MSPE) approach was successfully used to synthesize the covering silica on magnetic nanoparticles. The SiO<sub>2</sub>-MNP were employed as adsorbents for phenol extraction in ambient water samples through hydrophobic interaction and hydrogen bonding.
2. The optimized parameters for adsorption were selected as 5 min and pH 8. These parameters were used to treat real water samples and the recoveries obtained ranged from 84.8% to 92.4% of phenol uptake. This approach is quicker, easier to use, more effective, and less time-consuming than other available methods.

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