# Effects of Adding Fe<sub>2</sub>O<sub>3</sub> and Fe<sub>3</sub>O<sub>4</sub> Nanoparticles to Soil on Germination and Seedling Characteristics of Oriental Beech

Hakan Sevik, a,\* Handan Ucun Ozel, Yafes Yildiz, and Halil Barış Özel C

Effects of Fe-based nanoparticles (Fe<sub>2</sub>O<sub>3</sub> and Fe<sub>3</sub>O<sub>4</sub>) on germination parameters were studied for some seedling characteristics of Oriental beech (Fagus orientalis) seeds. Fe<sub>2</sub>O<sub>3</sub> and Fe<sub>3</sub>O<sub>4</sub> nanoparticle applications were made at concentrations of 400, 800, 1200, 1600, and 2000 mg/L on Fagus orientalis seeds collected from 10 different populations, and some germination and seedling characteristics were evaluated. Preliminary results generally indicated that low-dose nanoparticle applications positively affected germination and seedling characteristics, while increases in doses led to decreases in these parameters. Values obtained from high-dose nanoparticle applications were generally lower than those from the control group. The iron nanoparticles affected the parameters to different extents, Fe<sub>2</sub>O<sub>3</sub> nanoparticles showed a significant positive effect on germination rate and radicle length, while exhibiting a significant negative effect on germination percentage and plumule length. The populations least affected by highdose iron nanoparticle applications were Bursa Inegol, Karabuk-Yenice, and Ordu Akkus, while the most affected were the Bartin-Kumluca and Kahramanmaras-Andirin populations.

DOI: 10.15376/biores.20.1.70-82

Keywords: Nanoparticle; Fe<sub>2</sub>O<sub>3</sub>; Fe<sub>3</sub>O<sub>4</sub>; Fagus orientalis; Germination

Contact information: a: Department of Environmental Engineering Faculty of Engineering and Architecture, Kastamonu University, Türkiye; b: Department of Environmental Engineering Faculty of Engineering, Architecture and Design, Bartın University, Türkiye; c: Department of Forest Engineering, Faculty of Forestry, Bartın University, Türkiye; \*Corresponding author: hakansevik@gmail.com

# INTRODUCTION

Plants are the basis of all living life on Earth. Therefore, understanding and directing plant growth and development is one of the primary study topics for human beings (Ozel *et al.* 2021; Özdikmenli *et al.* 2024). As is known, the growth and development of plants, as in all living organisms, are shaped under the influence of genetic structure (Hrivnak *et al.* 2024) and environmental conditions (Yayla *et al.* 2022; Sevik *et al.* 2024). The main environmental factors affecting plant growth are climatic and edaphic (Key *et al.* 2023; Koc *et al.* 2024). Edaphic factors include many components, such as soil nutrient content, pH, and soil texture and structure (Erdem *et al.* 2023).

The chemical structure of the soil is the most important factor affecting and shaping plant growth. The presence and amount of nutrients required for plant growth are essential. However, metals, essential as nutrients, can become lethal to plant biology at high concentrations. Therefore, it is crucial to provide plants with the optimum amounts of metals so that normal metabolic functions can be maintained without metals deficiency or

phytotoxicity (Natasha *et al.* 2022). Moreover, the optimum amount of each metal for plants needs to be determined individually for each plant. The optimum metal concentration and toxicity limit for each plant is different (Pavlovic *et al.* 2021; Kaur *et al.* 2023).

Iron is one of the metals absolutely necessary for plant growth. Iron (Fe) is an essential element for plants and plays many important roles in physiological and metabolic processes. It is a redox active, highly reactive element, and thus higher concentrations of Fe can be toxic to plants (Zuo and Zhang 2011; Kuzmina *et al.* 2023). Although Fe is present in high concentrations in soil, it is not readily taken up by plants. Fe is the third most limiting nutrient for plant growth and metabolism, primarily due to the low solubility of the oxidized ferric form in aerobic environments (Zuo and Zhang 2011). The plant growth depends on the Fe availability that is subjected to several factors including physicochemical properties of soil and microbial Fe metabolism. Plants secrete protons, phenolics, and metabolites (*e.g.* mugineic acid) to aid conversion of Fe from Fe<sup>3+</sup> to Fe<sup>2+</sup> and to increase chelation of Fe for uptake (Ishimaru *et al.* 2011). The optimum amount of Fe and the level of toxicity have been determined in many cases. For example, for *Triticum aestivum*, 10 to 50 mg L<sup>-1</sup> level is optimum, while 250 mg L<sup>-1</sup> level is toxic and 1000 mg L<sup>-1</sup> level is lethal (Kaur *et al.* 2023). In *Oryza sativa*, Fe levels that cause toxicity range from as low as 10 mg Fe L<sup>-1</sup> to 500 mg Fe L<sup>-1</sup> or higher (Sahrawat 2005).

In plant development and especially in cultivated plants, the elimination of elemental deficiency in the soil due to long-term production is of great importance in terms of ensuring product continuity. Fertilization for this purpose is vital for plant development (Erdem *et al.* 2023). Soil fertilization with nanoparticles shows promising results for agriculture, especially for crop production (Mielcarz-Skalska *et al.* 2021). In recent years, nanoparticles have been used intensively in many fields, not only for fertilization (Özel *et al.* 2024).

Among the most widely used nanoparticles, Fe<sub>2</sub>O<sub>3</sub> and Fe<sub>3</sub>O<sub>4</sub> nanoparticles are used for the recovery of contaminated water and soil, offering great promise for use in biomedicine and water treatment due to their superparamagnetic and adsorptive properties. Recent studies suggest that Fe-based nanoparticles may also be beneficial for improving plant growth and Fe nutrient accumulation in agricultural plants. However, the potential adverse effects of nanoparticle applications on other organisms remain a serious concern, despite the fact that it generally has positive effect on agricultural plants (Tombuloglu et al. 2022). As a result of the reaction of iron (III) and oxygen, there are about 16 different iron oxide species in nature (Nanography 2024). Iron oxide NPs are often reported in literature using the formula Fe<sub>3</sub>O<sub>4</sub>, but they also can have different forms such as magnetite (Fe<sub>3</sub>O<sub>4</sub>), maghemite ( $\gamma$ -Fe<sub>2</sub>O<sub>3</sub>), hematite ( $\alpha$ -Fe<sub>2</sub>O<sub>3</sub>), FeO,  $\epsilon$ -Fe<sub>2</sub>O<sub>3</sub>, and  $\beta$ Fe<sub>2</sub>O<sub>3</sub>. The different forms possess different magnetic behavior. For instance, magnetite and maghemite are ferromagnetic or superparamagnetic; however, hematites are weakly ferromagnetic or antiferromagnetic (Tombuloglu et al. 2022). For example, magnetic Fe<sub>3</sub>O<sub>4</sub> nanoparticles are superparamagnetic below the size of 20 nm. As the nanoparticle size decreases, this property tends towards paramagnetic or superparamagnetic magnetization (Ajinkya et al. 2020). This magnetic property is very valuable for many applications when produced with controlled size and crystal structure.

Nanoparticle compounds can show different properties from those in nature (Nanography 2024). Nanoparticles can be defined as nano-sized powder grains or particles having dimensions in the range of 1 to 100 nm. Compared to their bulk structures, nanoscale materials exhibit different properties due to quantum size effects, size

dependence of the electronic structure, and the number of surface atoms. Nanoparticles have enormous surface energies due to their enormous surface area to volume ratio. In this way, while their chemical and physical properties change, their functionality also changes. Therefore, naturally occurring compounds and nanoparticles are compounds of different character (Seyhan 2022).

Iron nanoparticles can have adverse effects, especially on soil organisms, stimulate abundant ROS (reactive oxygen species) production in plants, cause oxidative stress damage, and thus cause inhibition of plant growth. They can cause serious disruptions in photosynthesis, especially in chlorophyll synthesis, leading to adverse effects on plants (Tao *et al.* 2023). It has also been determined that cell density decreases dose-dependently (Ameen *et al.* 2021).

Despite having various studies conducted in agricultural areas, the number of studies on the effects of nanoparticles on forest ecosystems and forest elements is much more limited. In this study, it was aimed to identify the preliminary effects of Fe<sub>2</sub>O<sub>3</sub> and Fe<sub>3</sub>O<sub>4</sub> nanoparticles on the germination parameters of seeds collected from different locations of oriental beech (*Fagus orientalis*), which is one of the important native tree species in the authors' country, at different doses.

Iron-based nanoparticles can have different effects in soluble ionic forms, and this effect may differ depending on the plant species. Gui *et al.* (2015) observed that inhibition of rice root phytohormones under hydroponic conditions was positively correlated with nFe<sub>2</sub>O<sub>3</sub> concentration up to 200 mg/L. However, nFe<sub>2</sub>O<sub>3</sub> was reported to cause oxidative stress in maize and roots of Citrus maxima and also reduced leaf chlorophyll content (Li *et al.* 2016; Hu *et al.* 2017). Yang *et al.* (2020) reported that exposure of soybean to nFe<sub>2</sub>O<sub>3</sub> did not cause any toxicity stress or physiological disorders, on the contrary, exposure to nFe<sub>2</sub>O<sub>3</sub> significantly improved physiological performance by causing increases in chlorophyll content, plant biomass and root growth indices.

However, high amounts of nanoparticles may result in the presence of higher concentrations of iron ions of various types adjacent to the nanoparticles in moist soil due to diffusion mechanism, which may negatively affect plant growth. Therefore, the main hypothesis of the study was "Increasing dosage of Fe-based nanoparticles negatively affects germination and seedling characters in *Fagus orientalis* seeds".

### **EXPERIMENTAL**

# **Materials and Method**

The seeds collected within the scope of the study were obtained from the oriental beech forests of Türkiye. Information regarding the regions and populations, from which the seeds were collected, is given in Table 1.

After being subjected to health tests and showing no issues in their embryo and endosperm parts, and demonstrating healthy and normal development performance, the collected seeds were used in germination tests. One of the main objectives of the research was to identify the effects of iron nanoparticles on the germination parameters of oriental beech seeds as well as to establish toxic threshold values. For this purpose, under sterile and hygienic conditions in the laboratory, five different nanoparticle concentrations for both Fe<sub>2</sub>O<sub>3</sub> (average diameter: <30 nm, purity: >99.5%) and Fe<sub>3</sub>O<sub>4</sub> (average diameter: <20 nm, purity: >99.5%) nanoparticles were prepared in the amounts of 400, 800, 1200,

1600, and 2000 mg/L, thus a total of ten concentrations were then stored in sterile concentration bottles to be applied to the seeds.

Table 1. Populations from Which the Seeds Were Collected

	Region	Coordi	inates	Altitude (m)	Exposure	Stand Type	
4	Adapazari Karaau	41º 01' 13"	30° 44' 45"	200 to 500	Northwest		
1	Adapazari-Karasu	41° 00' 38"	30° 45′ 01″	300 to 500	Northwest	Knd <sub>2</sub>	
2	Balikesir-Dursunbey	39° 27' 35"	28° 33′ 54″	800 to	North	Knd₁	
	Dalikesii-Dursumbey	39° 27' 30"	28° 32' 44"	1000	NOTH	Kiiu <sub>1</sub>	
3	Bartin-Kumluca	41º 29' 43"	32° 26′ 32″	450 to 700	North	Knd₃	
3	Dartin-Kumiuca	41° 29' 55"	32° 27' 54"	450 10 700	NOTH	KHU3	
4	Bursa-Inegol	39° 58' 44"	29° 29' 04"	700 to	Northwest	Knd₃	
4	Bursa-megor	39° 58' 02"	29º 28' 46"	1200	Northwest	KHU3	
5	Canakkale-Kalkim	39° 46′ 56″	27º 09' 46"	400 to 600	North	$Knd_2$	
3	Carlakkale-Kalkiiii	39º 46' 11"	27º 09' 10"	400 10 000	NOTH	MIU <sub>2</sub>	
6	Duzce-Yigilca	31° 25′ 35″	31° 25′ 35″	700 to	Northwest	Knd₃	
0	Duzce- rigilca	31º 25' 35"	31° 25′ 35″	1100	Northwest	KHU3	
7	Kahramanmaras-	37º 46' 12"	36º 22' 10"	1400 to	North	Knd₃	
	Andirin	37º 44' 52"	36° 23′ 34″	1800	NOTH	Miu3	
8	Karabuk-Yenice	41° 09' 08"	32º 16' 37"	600 to 900	North	Knd₁	
0	Narabuk-Terrice	41° 08' 52"	32º 15' 53"	800 10 900	NOTH	Kiiu <sub>1</sub>	
9	Ordu-Akkus	40° 47' 25"	36° 58' 50"	1100 to	North	$Knd_2$	
9	Oldu-Akkus	40° 46' 56"	36° 59' 05"	1400	NOTH	rtiiu <sub>2</sub>	
10	Zonguldak-Devrek	41º 12' 27"	31º 53' 15"	500 to 700	Northwest	Knd₁	
10	Zoriguluak-Deviek	41º 16' 13"	32° 12' 52"	300 10 700	Northwest	rtiiu <sub>1</sub>	

The beech seeds treated with prepared concentrations in five different doses were placed in single-use sterile petri dishes prepared with quantitative filter papers to be used in germination tests. Five repetitions were performed for each dose, and nanoparticle treatment was applied to 150 seeds for each dose, with 30 healthy seeds per repetition. A total of 900 seeds, including the control group, were used in the germination tests. Germination tests were conducted in a 3M Climacell brand germination cabinet. The temperature of the germination medium in the cabinet was set to 20 °C, relative humidity to 70%, and the lighting duration to 12 h.

Oriental beech seeds, placed in 100-mL petri dishes in a way not to touch each other, were monitored for germination by exposing them to 10 mL of nanoparticle solution daily. On the 7<sup>th</sup> day of application, the number of germinated seeds was counted to calculate the germination rate (GR). The applications continued for 35 days, and at the end of the 35<sup>th</sup> day, the seedling height (SH), root collar diameter (RCD), plumule length (PL), radicle length (RL), and radicle thickness (RT) were measured using a digital micro-meter compass. Non-germinated seeds were cut to check if they were healthy, and the germination percentage (GP) was calculated by comparing the total number of germinated seeds to the total number of healthy seeds. Similarly, the germination rate was calculated as the ratio of the number of germinated seeds on the 7<sup>th</sup> day to the number of healthy seeds. The data obtained were analysed using SPSS 22.0 software package, and variance analysis and Duncan test were applied to the data.

# **RESULTS**

Data showing the changes in germination rate is presented in Table 2.

**Table 2.** Changes in Germination Percentage (%)

Pop.	Fe <sub>3</sub> O <sub>4</sub>							Fe <sub>2</sub> O <sub>3</sub>					
	Cont.	400	800	1200	1600	2000	Cont.	400	800	1200	1600	2000	
1	5.9	5.6	5.5	5.0	5.0	4.3	5.9	7.0	6.7	6.7	6.3	5.5	
2	5.4	5.7	5.6	5.3	5.1	4.7	5.4	7.1	6.7	6.3	6.6	5.6	
3	5.6	5.2	5.1	4.9	4.6	4.5	5.6	6.9	6.6	5.9	6.0	5.1	
4	5.2	5.9	5.8	5.5	5.3	5.0	5.2	7.1	6.8	6.7	6.6	6.0	
5	5.8	5.4	5.4	4.9	4.6	4.6	5.8	7.0	6.4	6.1	6.1	5.4	
6	5.3	5.6	5.4	4.9	4.9	4.8	5.3	6.6	6.7	6.4	6.2	6.0	
7	5.4	5.0	5.1	4.7	4.6	4.3	5.4	6.4	6.2	6.1	6.1	5.2	
8	5.2	5.8	5.5	5.2	5.1	5.1	5.2	6.9	6.8	6.6	6.3	5.9	
9	5.6	6.1	5.8	5.5	5.3	5.2	5.6	7.4	6.9	6.6	6.5	6.3	
10	5.3	5.4	5.3	5.1	4.7	4.5	5.3	6.7	6.7	6.4	6.0	5.1	
Av.	5.5	5.6	5.5	5.1	4.9	4.7	5.5	6.9	6.7	6.4	6.3	5.6	
StD.	0.2	0.3	0.2	0.3	0.3	0.3	0.2	0.3	0.2	0.3	0.2	0.4	

Pop.; Population number, Cont.; Control, Av.; Average, StD.; Standard deviation

When the changes in germination percentage were examined, it was observed that in both nanoparticle applications, the germination percentage significantly increased at low doses and began to decrease with increasing doses. The germination at a dose of 400 mg/L increased by approximately 50% in the Fe<sub>3</sub>O<sub>4</sub> nanoparticle application, while it increased more than twice in the Fe<sub>2</sub>O<sub>3</sub> nanoparticle application. Afterwards, the germination started to decrease with increasing doses, but even at the highest dose application of 2000 mg/L, it did not fall below the values in the control group.

**Table 3.** Changes in Germination Percentage (%)

Pop			Fe <sub>3</sub> C	)4			Fe <sub>2</sub> O <sub>3</sub>						
	Control	400	800	1200	1600	2000	Control	400	800	1200	1600	2000	
1	68.4	84.0	80.7	78.9	77.2	76.3	68.4	71.1	71.9	67.1	64.2	63.1	
2	71.6	84.7	85.5	79.6	77.1	77.4	71.6	75.1	75.4	73.0	70.4	69.5	
3	59.3	78.9	77.3	74.9	73.9	73.5	59.3	66.5	64.6	60.8	59.5	59.4	
4	75.2	85.5	84.1	80.9	79.5	79.0	75.2	80.9	79.7	74.9	73.6	72.8	
5	66.0	80.9	79.2	77.0	75.4	75.8	66.0	70.6	70.4	65.1	64.0	63.4	
6	69.9	85.0	83.5	79.0	76.3	77.0	69.9	74.1	72.5	68.7	65.6	65.7	
7	57.7	77.6	77.0	74.8	73.0	72.9	57.7	65.5	64.8	58.8	56.7	55.1	
8	74.6	83.5	85.6	80.8	79.7	78.9	74.6	78.2	79.9	74.3	71.6	71.1	
9	78.8	87.4	86.4	81.4	78.9	77.0	78.8	82.3	81.9	77.0	72.5	72.0	
10	59.8	82.0	80.1	78.8	75.9	74.2	59.8	69.7	68.7	63.9	62.4	62.6	
Av.	68.1	83.0	81.9	78.6	76.7	76.2	68.1	73.4	73.0	68.4	66.1	65.5	
StD.	7.3	3.1	3.5	2.4	2.3	2.1	7.3	5.8	6.2	6.3	5.8	5.8	

When the changes in germination percentage was examined based on population, it was observed that the highest germination rates were obtained in the P4 and P9 populations, while the lowest germination rates were obtained in the P7 population. The trend showing the changes in germination percentage is presented in Table 3.

It was observed that the application of Fe nanoparticles significantly affected the germination percentage. In both nanoparticle applications, the germination percentage increased significantly at low doses, with this increase being higher in the Fe<sub>3</sub>O<sub>4</sub> nanoparticle application. However, in both applications, the germination percentage decreased significantly at the application of 1200 mg/L, and it continued to decrease with increasing doses. While the germination percentage in Fe<sub>3</sub>O<sub>4</sub> nanoparticles was generally higher than that in the control group even at high doses, the germination percentages obtained at 2000 mg/L application in Fe<sub>2</sub>O<sub>3</sub> nanoparticles were generally lower than those in the control group.

When the changes in germination percentage were examined based on population, it was observed that the highest germination percentages were obtained in the P4, P8, and P9 populations. The lowest germination percentages were obtained in the P3 and P7 populations. The graph showing the changes in seedling height is presented in Table 4.

Pop				Fe <sub>3</sub> O <sub>4</sub>				Fe <sub>2</sub> O <sub>3</sub>					
	Cont.	400	80	120	160	200	Cont.	400	80	120	160	200	
			0	0	0	0			0	0	0	0	
1	7.1	7.4	7.4	7.2	6.9	6.8	7.1	7.1	7.0	6.8	6.5	6.1	
2	7.2	7.7	7.6	7.3	7.1	7.1	7.2	7.4	7.3	7.0	6.6	6.4	
3	6.4	7.2	7.2	6.9	6.8	6.5	6.4	6.9	6.7	6.3	6.2	5.9	
4	7.2	7.9	7.9	7.6	7.3	7.2	7.2	7.7	7.5	7.2	7.0	6.7	
5	7.0	7.5	7.3	7	6.8	6.6	7.0	7.1	6.9	6.7	6.4	6.1	
6	7.1	7.6	7.6	7.3	7.1	7.0	7.1	7.4	7.3	6.9	6.5	6.2	
7	6.4	7.1	7.0	6.7	6.6	6.4	6.4	6.8	6.6	6.2	6.1	5.9	
8	7.2	7.9	7.8	7.3	7.2	7.1	7.2	7.6	7.4	7.3	6.8	6.7	
9	7.3	8.2	8.1	7.8	7.5	7.2	7.3	8.3	7.6	7.4	6.8	6.4	
10	6.6	7.3	7.2	6.9	6.7	6.5	6.6	7.0	6.8	6.6	6.4	6.0	
Av.	7.0	7.6	7.5	7.2	7.0	6.8	7.0	7.0	7.3	7.1	6.8	6.5	
StD.	0.3	0.3	0.4	0.3	0.3	0.3	0.3	0.3	0.5	0.4	0.4	0.3	

Table 4. Changes in Seedling Height (cm)

Similar to germination values, it was determined that Fe nanoparticle applications significantly affected the seedling height as well. In both nanoparticle applications, seedling height increased significantly at low doses, with this increase being higher in the Fe<sub>3</sub>O<sub>4</sub> nanoparticle application. However, seedling height decreased with increasing nanoparticle levels. In the Fe<sub>2</sub>O<sub>3</sub> nanoparticle application, it was determined that the decrease in seedling height with increasing doses was at higher levels.

Seedling height was also observed to be varying significantly on a population basis. The highest seedling heights were observed in the P4, P8, and P9 populations, while the lowest seedling heights were observed in the P3, P7, and P10 populations. The graph showing the changes in root collar diameter is presented in Table 5.

75

Pop			Fe <sub>3</sub> (	<b>D</b> <sub>4</sub>			Fe <sub>2</sub> O <sub>3</sub>						
	Cont.	400	800	1200	1600	2000	Cont.	400	800	1200	1600	2000	
1	0.8	0.9	0.8	0.9	0.9	0.8	0.8	0.9	0.8	0.9	0.8	0.8	
2	0.9	0.9	1.0	0.8	0.9	0.9	0.9	1.0	1.0	0.9	0.9	0.8	
3	0.8	0.9	0.9	0.9	0.8	8.0	0.8	0.8	0.9	0.9	0.9	8.0	
4	0.9	0.9	0.9	1.0	0.9	0.9	0.9	1.0	0.9	0.9	0.9	0.9	
5	0.8	0.9	0.9	0.8	0.9	0.8	0.8	0.9	0.9	0.9	0.8	0.8	
6	0.8	0.9	0.9	0.9	0.9	0.8	0.8	1.0	0.9	0.9	0.8	0.8	
7	0.8	0.9	0.8	0.9	0.8	0.7	0.8	0.9	0.9	0.9	0.8	0.8	
8	0.9	1.0	1.0	0.8	0.9	0.9	0.9	1.0	0.9	0.9	0.9	1.0	
9	0.9	1.0	0.9	1.0	0.9	0.9	0.9	0.9	1.0	0.9	0.9	0.9	
10	0.8	0.9	0.8	0.9	0.8	0.8	0.8	0.9	0.8	0.8	0.8	0.8	
Av.	0.8	0.9	0.9	0.9	0.9	0.8	0.8	0.9	0.9	0.9	0.9	0.8	
StD.	0.1	0.0	0.1	0.1	0.0	0.1	0.1	0.1	0.1	0.0	0.1	0.1	

**Table 5.** Changes in Root Collar Diameter (mm)

As shown in Table 5, the values of root collar diameter generally ranged between 0.8 and 1.0 mm, and it was not possible to say that there was a significant change depending on both the population and the nanoparticles applied. According to these results, it can be said that the root collar diameter did not change noticeably depending on the nanoparticle applications. The graph showing the changes in plumule length is presented in Table 6.

Table 6. Changes in Plumule Length (cm)

Pop			Fe	O <sub>4</sub>			Fe <sub>2</sub> O <sub>3</sub>					
	Cont.	400	800	1200	1600	2000	Cont.	400	800	1200	1600	2000
1	1.1	1.0	0.9	1.0	1.0	0.9	1.1	0.9	1.0	0.8	0.8	0.8
2	1.1	1.0	1.0	1.0	1.0	1.0	1.1	1.0	0.9	0.9	0.9	0.9
3	0.9	1.0	0.9	0.9	0.9	0.9	0.9	0.9	0.9	0.8	8.0	0.8
4	1.1	1.0	1.0	1.0	1.0	1.0	1.1	0.9	1.0	1.0	0.9	0.8
5	1.0	1.0	0.9	1.0	0.9	0.9	1.0	0.9	0.9	0.8	0.8	0.8
6	1.1	1.0	1.0	1.0	1.0	0.9	1.1	0.9	0.9	0.9	0.9	0.8
7	0.9	0.9	1.0	0.9	0.9	0.8	0.9	0.8	0.9	0.9	8.0	0.8
8	1.1	1.0	1.0	1.0	1.0	1.0	1.1	1.0	1.0	1.0	0.9	0.8
9	1.1	1.1	1.1	1.0	1.1	1.0	1.1	1.0	1.0	1.0	0.9	0.9
10	1.0	1.0	0.9	0.9	0.9	0.9	1.0	0.9	0.8	0.9	8.0	0.8
Av.	1.0	1.0	1.0	1.0	1.0	0.9	1.0	0.9	0.9	0.9	0.9	0.8
StD.	0.1	0.0	0.1	0.0	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.0

It can be said that the changes in plumule length were not significant in the  $Fe_3O_4$  application, while they tended to decrease with increasing doses in the  $Fe_2O_3$  application. It was observed that the plumule length, which was in the range of 0.9 to 1 mm in the control group for the  $Fe_2O_3$  application, dropped to the level of 0.8 mm in most populations

with the application of Fe<sub>2</sub>O<sub>3</sub>. The graph showing the changes in radicle thickness is presented in Table 7.

**Table 7.** Changes in Radicle Thickness (mm)

Pop			Fea	O <sub>4</sub>			Fe <sub>2</sub> O <sub>3</sub>					
	Cont.	400	800	1200	1600	2000	Cont.	400	800	1200	1600	2000
1	1.2	1.0	1.0	1.0	0.9	0.9	1.2	1.0	0.9	0.9	0.8	0.9
2	1.3	1.1	1.0	0.9	1.0	0.9	1.3	1.2	1.0	1.0	0.9	0.9
3	1.1	1.0	1.0	1.0	0.8	0.8	1.1	1.1	1.0	0.9	0.9	0.9
4	1.3	1.1	1.0	1.0	1.0	0.9	1.3	1.0	0.9	0.9	1.0	0.9
5	1.1	1.1	1.0	1.0	1.0	0.9	1.1	1.1	1.0	0.9	0.9	0.9
6	1.2	1.0	1.1	1.0	0.9	0.9	1.2	1.0	0.9	0.9	1.0	0.9
7	1.0	0.9	1.0	0.9	0.9	8.0	1.0	0.9	1.0	0.8	0.8	8.0
8	1.2	1.0	1.0	1.0	1.1	0.9	1.2	1.1	1.0	0.9	1.0	0.9
9	1.3	1.1	1.1	1.1	1.0	0.9	1.3	1.1	1.0	0.9	1.0	0.9
10	1.1	1.0	1.0	0.9	0.9	0.9	1.1	0.9	1.0	8.0	0.9	0.9
Av.	1.2	1.0	1.0	1.0	1.0	0.9	1.2	1.0	1.0	0.9	0.9	0.9
StD.	0.1	0.1	0.0	0.1	0.1	0.0	0.1	0.1	0.0	0.1	0.1	0.0

As shown in Table 7, the radicle thickness significantly decreased after the application of Fe nanoparticles, when it was 1 mm or more in the control group, and it continued to decrease with increasing doses. This decrease was much faster in the  $Fe_2O_3$  application. In the  $Fe_2O_3$  application, from the dose of 1200 mg/L onwards, the average radicle length in most populations is observed to be around 0.8 mm. The graph showing the changes in radicle length is presented in Table 8.

 Table 8. Changes in Radicle Length (cm)

Pop			Fe	O <sub>4</sub>		Fe <sub>2</sub> O <sub>3</sub>						
	Cont.	400	800	1200	1600	2000	Cont.	400	800	1200	1600	2000
1	5.9	5.6	5.5	5.0	5.0	4.3	5.9	7.0	6.7	6.7	6.3	5.5
2	5.4	5.7	5.6	5.3	5.1	4.7	5.4	7.1	6.7	6.3	6.6	5.6
3	5.6	5.2	5.1	4.9	4.6	4.5	5.6	6.9	6.6	5.9	6.0	5.1
4	5.2	5.9	5.8	5.5	5.3	5.0	5.2	7.1	6.8	6.7	6.6	6.0
5	5.8	5.4	5.4	4.9	4.6	4.6	5.8	7.0	6.4	6.1	6.1	5.4
6	5.3	5.6	5.4	4.9	4.9	4.8	5.3	6.6	6.7	6.4	6.2	6.0
7	5.4	5.0	5.1	4.7	4.6	4.3	5.4	6.4	6.2	6.1	6.1	5.2
8	5.2	5.8	5.5	5.2	5.1	5.1	5.2	6.9	6.8	6.6	6.3	5.9
9	5.6	6.1	5.8	5.5	5.3	5.2	5.6	7.4	6.9	6.6	6.5	6.3
10	5.3	5.4	5.3	5.1	4.7	4.5	5.3	6.7	6.7	6.4	6.0	5.1
Av.	5.5	5.6	5.5	5.1	4.9	4.7	5.5	6.9	6.7	6.4	6.3	5.6
StD.	0.2	0.3	0.2	0.3	0.3	0.3	0.2	0.3	0.2	0.3	0.2	0.4

The Fe nanoparticle applications were observed to significantly affect radicle length. Particularly, in the low-dose  $Fe_2O_3$  application, radicle length increased significantly; however, in both applications, radicle length decreased significantly with increasing doses. Especially, in the  $Fe_2O_3$  nanoparticle application at 2000 mg/L, radicle length showed a significant decrease.

When the changes in radicle length were examined the basis of population, it was observed that the highest values were obtained in the P4, P6, P8, and P9 populations. The lowest germination percentages were obtained in the P3 and P7 populations.

### DISCUSSION

The study results showed that iron nanoparticle applications affected almost all of the measured characteristics. Generally, germination and seedling characteristics were positively affected at low nanoparticle doses, but decreases began in these parameters with increasing doses. Values obtained from high-dose nanoparticle applications were generally lower than those in the control group. Therefore, it was found that low nanoparticle applications had a positive effect on germination and seedling characteristics, but they had a negative effect as the dose increases. This result means that the main hypothesis of the study is accepted. This finding is in compliance with the information obtained in the literature.

Recent studies suggest that Fe-based nanoparticles may also be beneficial for improving plant growth and Fe accumulation in plants. For instance, it is stated that the addition of 2 mg kg<sup>-1</sup> Fe<sub>2</sub>O<sub>3</sub> nanoparticles can increase chlorophyll content, regulate phytohormones, and increase Fe content. In contrast, the negative effects of Fe-based nanoparticles have also been reported in many studies, and their use in such applications in the future has been discussed. For example, Fe<sub>3</sub>O<sub>4</sub> nanoparticles have been reported to cause oxidative stress in ryegrass and pumpkin grown hydroponically. Additionally, it is stated that Fe<sub>2</sub>O<sub>3</sub> nanoparticles at 50 mg L<sup>-1</sup> concentration can disrupt photosynthesis by reducing chlorophyll content, while nanoparticles at a concentration of 500 mg L<sup>-1</sup> can cause excessive accumulation of ROS in cells, leading to DNA damage and a decrease in mitotic index in plant cells (Li *et al.* 2021). Low concentrations of nZVI nanoparticles were found to have positive effects on *Oryza sativa* seeds and seedlings, such as increased root and shoot length, biomass and photosynthetic pigment content. However, seedlings prepared with 160 mgL<sup>-1</sup> nZVI were subjected to oxidative stress and SEM micrographs also revealed root tissue damage at this concentration (Guha *et al.* 2018).

The parameters under study were determined to be affected by iron nanoparticles at different levels. While Fe<sub>2</sub>O<sub>3</sub> nanoparticles showed a significant positive effect on germination rate and radicle length, a significant negative effect was observed on germination percentage and plumule length. The studies conducted on the subject indicate that nanoparticles of different sizes and concentrations affect germination and seedling characteristics at different levels. In a study, the effects of Fe<sub>3</sub>O<sub>4</sub> nanoparticles were investigated on seed germination in tobacco plant (*Nicotiana tabacum*), and it was found that the radicle lengths of seeds treated with 5 nm (30 mg/L concentration) and 10 nm (10 and 30 mg/L concentration) were significantly shorter than those of control seeds. In contrast, the radicle lengths of seeds treated with 10 nm (3 mg/L concentration) and 20 nm (10 mg/L concentration) were found to be significantly longer than those of control seeds.

Most of the seeds treated with nanoparticles show significantly higher seed germination percentages (Alkhatib *et al.* 2021).

In a study conducted by Asadi-Kavan *et al.* (2020), the effects of iron oxide nanoparticles (Fe<sub>2</sub>O<sub>3</sub>) on the germination and seedling growth of *Oenothera biennis* were determined, and it was found that Fe<sub>2</sub>O<sub>3</sub> increased the germination percentage 89.2%, the germination tolerance index 53.4%, and the root tolerance index 82.2%. The study concluded that the use of Fe<sub>2</sub>O<sub>3</sub> at low and medium concentrations did not have toxic effects on this plant. Gupta *et al.* (2022) reported that Fe<sub>3</sub>O<sub>4</sub> nanoparticle application in *Cucumis sativus* increased seed yield 52.2%, total chlorophyll content 17.3%, germination percentage 17.0%, and seedling vigor 72.6%. A significant increase also was observed in starch, soluble proteins, soluble sugars, and fat content in seeds obtained from plants treated with Fe<sub>3</sub>O<sub>4</sub> nanoparticles. It was determined that Fe<sub>2</sub>O<sub>3</sub> and Fe<sub>3</sub>O<sub>4</sub> nanoparticles significantly increased the germination rate (approximately 37% for Fe<sub>2</sub>O<sub>3</sub>; approximately 63% for Fe<sub>3</sub>O<sub>4</sub>), plant biomass and pigmentation in *Hordeum vulgare* (Tombuloglu *et al.* 2022).

In *Nicotiana tabacum*, it is found that Fe<sub>3</sub>O<sub>4</sub> nanoparticles accumulate only in clusters around the root cell wall, indicating that Fe<sub>3</sub>O<sub>4</sub> nanoparticles move through the apoplastic pathway in roots (Yuan *et al.* 2018; Alkhatib *et al.* 2021). It is stated that Fe<sub>3</sub>O<sub>4</sub> nanoparticles are efficiently taken up by roots and transported to leaves regardless of their size, but small-sized Fe<sub>3</sub>O<sub>4</sub> nanoparticles can be more reactive due to their size properties, which then lead to cell stress and membrane damage (Tombuloglu *et al.* 2024). In seeds treated with Fe<sub>3</sub>O<sub>4</sub> nanoparticles, it is indicated that chlorophyll is completely consumed and albinism occurs, potentially causing a lethal effect on chlorophyll synthase (Alkhatib *et al.* 2021). Germination is found to be decreasing in *Helianthus annuus* seeds treated with Fe nanoparticles, indicating a decrease in the content of elements associated with this process (Ca, Mg, K, P, and Na), and it is suggested that these elements are adsorbed by Fe nanoparticles (Kornarzyński *et al.* 2020). As can be seen, the effects of nanoparticles on plants vary depending on size and dose, but generally, nanoparticle applications that increase efficiency at low doses are found to be harmful with increasing doses.

As a result of the study, the effects of iron nanoparticle applications on germination and seedling characteristics of beech seeds were evaluated at the population level, and it was determined that populations P4, P8, and P9 were the least affected by high-dose iron nanoparticle applications, while populations P3 and P7 were the most affected. When these populations were examined, it was observed that the least affected populations were mostly located at moderate altitudes, while the most affected populations were located at low or high altitudes, *i.e.*, at extreme altitudes.

## CONCLUSIONS

- 1. The results of the study revealed that high doses of Fe<sub>2</sub>O<sub>3</sub> and Fe<sub>3</sub>O<sub>4</sub> nanoparticles negatively affected germination and seedling development. However, it was determined that 400 mg/L nanoparticle application positively affected almost all characteristics. For this reason, 400 mg/L Fe<sub>2</sub>O<sub>3</sub> and Fe<sub>3</sub>O<sub>4</sub> nanoparticles are recommended, especially in seedling production studies.
- 2. The study results indicate that some populations are less affected by iron-based nanoparticles. These populations can be used as a priority seed source for seedling

production. However, it is recommended that studies on this subject to focus on more advanced stages, such as the seedling stage, before this stage.

# REFERENCES CITED

- Ajinkya, N., Yu, X., Kaithal, P., Luo, H., Somani, P., and Ramakrishna, S. (2020). "Magnetic iron oxide nanoparticle (IONP) synthesis to applications: Present and future" *Materials* 13(20), article 4644. DOI: 10.3390/ma13204644
- Alkhatib, R., Alkhatib, B., and Abdo, N. (2021). "Effect of Fe<sub>3</sub>O<sub>4</sub> nanoparticles on seed germination in tobacco," *Environmental Science and Pollution Research* 28(38), 53568-53577. DOI: 10.1007/s11356-021-14541-x
- Ameen, F., Alsamhary, K., Alabdullatif, J. A., and ALNadhari, S. (2021). "A review on metal-based nanoparticles and their toxicity to beneficial soil bacteria and fungi," *Ecotoxicology and Environmental Safety* 213, article 112027. DOI: 10.1016/j.ecoenv.2021.112027
- Asadi-Kavan, Z., Khavari-Nejad, R. A., Iranbakhsh, A., and Najafi, F. (2020). "Cooperative effects of iron oxide nanoparticle (α-Fe<sub>2</sub>O<sub>3</sub>) and citrate on germination and oxidative system of evening primrose (*Oenthera biennis* L.)," *Journal of Plant Interactions* 15(1), 166-179.
- Erdem, R., Aricak, B., Cetin, M., and Sevik, H. (2023). "Change in some heavy metal concentrations in forest trees by species, organ, and soil depth," *Forestist* 73(3), 257-263. DOI: 10.5152/forestist.2023.22069
- Guha, T., Ravikumar, K. V. G., Mukherjee, A., Mukherjee, A., and Kundu, R. (2018). "Nanopriming with zero valent iron (nZVI) enhances germination and growth in aromatic rice cultivar (*Oryza sativa* cv. Gobindabhog L.)," *Plant Physiology and Biochemistry* 127, 403-413. DOI: 10.1016/j.plaphy.2018.04.014
- Gui, X., Deng, Y., Rui, Y. Gao, B., Luo, W., Chen, S., Nhan, L,V., Li, X., Liu, S., Han, Y., Liu, L., and Xing, B,S. (2015). "Response difference of transgenic and conventional rice (*Oryza sativa*) to nanoparticles (γFe<sub>2</sub>O<sub>3</sub>)," *Environ. Sci. Pollut. Res.* 22, 17716-17723. DOI: 10.1007/s11356-015-4976-7
- Gupta, N., Jain, S. K., Tomar, B. S., Anand, A., Singh, J., Sagar, V., Kumar, R., Singh, V., Chaubey, T., Abd-Elsalam, K. A., *et al.* (2022). "Impact of foliar application of ZnO and Fe<sub>3</sub>O<sub>4</sub> nanoparticles on seed yield and physio-biochemical parameters of cucumber (*Cucumis sativus* L.) seed under open field and protected environment *vis a vis* during seed germination," *Plants* 11(23), article 3211. DOI: 10.3390/plants11233211
- Hrivnák, M., Krajmerová, D., Paule, L., Zhelev, P., Sevik, H., Ivanković, M., Goginashvili, N., Paule, J., and Gömöry, D. (2024). "Are there hybrid zones in *Fagus sylvatica* L. sensu lato?," *European Journal of Forest Research* 143, 451-464. DOI: 10.1007/s10342-023-01634-0
- Hu, J., Guo, H., Li, J., Gan, Q., Wang, Y., and Xing, B. (2017). "Comparative impacts of iron oxide nanoparticles and ferric ions on the growth of *Citrus maxima*," *Environ. Pollut.* 221, 199-208. DOI: 10.1016/j.envpol.2016.11.064
- Ishimaru, Y., Bashir, K., and Nishizawa, N. K. (2011). "Zn uptake and translocation in rice plants," *Rice* 4, 21-27. DOI: 10.1007/s12284-011-9061-3

- Kaur, H., Kaur, H., and Srivastava, S. (2023). "The beneficial roles of trace and ultratrace elements in plants," *Plant Growth Regulation* 100(2), 219-236. DOI: 10.1007/s10725-022-00837-6
- Key, K., Kulaç, Ş., Koç, İ., and Sevik, H. (2023). "Proof of concept to characterize historical heavy-metal concentrations in atmosphere in North Turkey: Determining the variations of Ni, Co, and Mn concentrations in 180-year-old *Corylus colurna* L. (Turkish hazelnut) annual rings," *Acta Physiologiae Plantarum* 45(10), article 120. DOI: 10.1007/s11738-023-03608-6
- Koc, İ., Canturk, U., Isinkaralar, K., Ozel, H. B., and Sevik, H. (2024). "Assessment of metals (Ni, Ba) deposition in plant types and their organs at Mersin City, Türkiye," *Environmental Monitoring and Assessment* 196(3), article 282.
- Kornarzyński, K., Sujak, A., Czernel, G., and Wiącek, D. (2020). "Effect of Fe<sub>3</sub>O<sub>4</sub> nanoparticles on germination of seeds and concentration of elements in *Helianthus annuus* L. under constant magnetic field," *Scientific Reports* 10(1), article 8068. DOI: 10.1038/s41598-020-64849-w
- Kuzmina, N., Menshchikov, S., Mohnachev, P., Zavyalov, K., Petrova, I., Ozel, H. B., Aricak, B., Onat, S. M., and Sevik, H. (2023). "Change of aluminum concentrations in specific plants by species, organ, washing, and traffic density," *BioResources* 18(1), 792-803. DOI: 10.15376/biores.18.1.792-803
- Li, J. L., Hu, J., Ma, C., Wang, Y., Wu, C., Huang, J., and Xing, B. S. (2016). "Uptake, translocation and physiological effects of magnetic iron oxide (γ-Fe<sub>2</sub>O<sub>3</sub>) nanoparticles in corn (*Zea mays* L.)," *Chemosphere* 159, 326-334. DOI: 10.1016/j.chemosphere.2016.05.083
- Li, M., Zhang, P., Adeel, M., Guo, Z., Chetwynd, A. J., Ma, C., Bai, T., Hao, Y., and Rui, Y. (2021). "Physiological impacts of zero valent iron, Fe<sub>3</sub>O<sub>4</sub> and Fe<sub>2</sub>O<sub>3</sub> nanoparticles in rice plants and their potential as Fe fertilizers," *Environmental Pollution* 269, article ID 116134. DOI: 10.1016/j.envpol.2020.116134
- Mielcarz-Skalska, L., Smolińska, B., and Włodarczyk, K. (2021). "Nanoparticles as potential improvement for conventional fertilisation in the cultivation of *Raphanus sativus* var. *sativus*," *Agriculture* 11(11), article 1067. DOI: 10.3390/agriculture11111067
- Nanografi (2024). "Demir oksit" (https://shop.nanografi.com.tr/blografi/demir-oksit-ozellikleri-uretimi-ve-uygulamalari-nanografi/), Accessed 18 August 2024.
- Natasha, N., Shahid, M., Bibi, I., Iqbal, J., Khalid, S., Murtaza, B., Bakhat, H. F., Farouk, A. B. U., Amjad, M., Hammad, H. M., and Arshad, M. (2022). "Zinc in soil-plant-human system: A data-analysis review," *Science of the Total Environment* 808, article 152024. DOI: 10.1016/j.scitotenv.2021.152024
- Ozel, H. B., Abo Aisha, A. E. S., Cetin, M., Sevik, H., and Zeren Cetin, I. (2021). "The effects of increased exposure time to UV-B radiation on germination and seedling development of Anatolian black pine seeds," *Environmental Monitoring and Assessment* 193(7), article 388. DOI: 10.1007/s10661-021-09178-9
- Özdikmenli, G., Yiğit, N., Özel, H. B., and Şevik, H. (2024). "Altitude-dependent variations in some morphological and anatomical features of Anatolian chestnut," *BioResources* 19(3), 4635-4651. DOI: 10.15376/biores.19.3.4635-4651
- Özel, H. B., Şevik, H., Yıldız, Y., and Çobanoğlu, H. (2024). "Effects of silver nanoparticles on germination and seedling characteristics of Oriental beech (*Fagus orientalis*) seeds," *BioResources* 19(2), 2135-2148. DOI: 10.15376/biores.19.2.2135-2148

- Pavlovic, J., Kostic, L., Bosnic, P., Kirkby, E. A., and Nikolic, M. (2021). "Interactions of silicon with essential and beneficial elements in plants," *Frontiers in Plant Science* 12, article 697592. DOI: 10.3389/fpls.2021.697592
- Sahrawat, K. L. (2005). "Iron toxicity in wetland rice and the role of other nutrients," *Journal of Plant Nutrition* 27(8), 1471-1504, DOI: 10.1081/PLN-200025869
- Seyhan, K. (2022). *Biosynthesis, Characterization and in Vitro Evaluation of Iron Oxide Nanoparticles*, Master's Thesis, Necmettin Erbakan University, Konya, Türkiye.
- Sevik, H., Koç, İ., and Cobanoglu, H. (2024). "Determination of some exotic landscape species as biomonitors that can be used for monitoring and reducing Pd pollution in the air," *Water Air Soil Pollut*. 235, article 615. DOI: 10.1007/s11270-024-07429-2
- Tao, Z., Zhou, Q., Zheng, T., Mo, F., and Ouyang, S. (2023). "Iron oxide nanoparticles in the soil environment: Adsorption, transformation, and environmental risk," *Journal of Hazardous Materials* 2023, article 132107. DOI: 10.1016/j.jhazmat.2023.132107
- Tombuloglu, H., Albenayyan, N., Slimani, Y., Akhtar, S., Tombuloglu, G., Almessiere, M., Baykal, A., Ercan, I., Sabit, H., and Manikandan, A. (2022). "Fate and impact of maghemite (γ-Fe2O3) and magnetite (Fe3O4) nanoparticles in barley (*Hordeum vulgare* L.)," *Environmental Science and Pollution Research* 29(3), 4710-4721. DOI: 10.1007/s11356-021-15965-1
- Tombuloglu, G., Aldahnem, A., Tombuloglu, H., Slimani, Y., Akhtar, S., Hakeem, K. R., Almessiere, M. A., Baykal, A., Ercan, I., and Manikandan, A. (2024). Uptake and bioaccumulation of iron oxide nanoparticles (Fe<sub>3</sub>O<sub>4</sub>) in barley (*Hordeum vulgare* L.): effect of particle-size," *Environmental Science and Pollution Research* 31, 22171-22186. DOI: 10.1007/s11356-024-32378-y
- Yang, X., Alidoust, D., and Wang, C. (2020). "Effects of iron oxide nanoparticles on the mineral composition and growth of soybean (*Glycine max* L.) plants," *Acta Physiologiae Plantarum* 42, 1-11. DOI: 10.1007/s11738-020-03104-1
- Yayla, E. E., Sevik, H., and Isinkaralar, K. (2022). "Detection of landscape species as a low-cost biomonitoring study: Cr, Mn, and Zn pollution in an urban air quality," *Environmental Monitoring and Assessment* 194(10), article 687. DOI: 10.1007/s10661-022-10356-6
- Yuan, J., Chen, Y., Li, H., Lu, J., Zhao, H., Liu, M., Nechitaylo, G. S., and Glushchenko, N. N. (2018). "New insights into the cellular responses to iron nanoparticles in *Capsicum annuum*," *Scientific Reports* 8(1), 1-9. DOI: 10.1038/s41598-017-18055-w
- Zuo, Y., and Zhang, F. (2011). "Soil and crop management strategies to prevent iron deficiency in crops," *Plant and Soil* 339, 83-95. DOI 10.1007/s11104-010-0566-0

Article submitted: July 5, 2024; Peer review completed: August 1, 2024; Revised version received: August 18, 2024; Accepted: October 24, 2024; Published: November 4, 2024. DOI: 10.15376/biores.20.1.70-82